

## Research Paper

**Cite this article:** Kallow S, Davies R, Panis B, Janssens SB, Vandeloos F, Mertens A, Swennen R, Tahir MB, Dickie J (2021). Regulation of seed germination by diurnally alternating temperatures in disturbance-adapted banana crop wild relatives (*Musa acuminata*). *Seed Science Research* 1–11. <https://doi.org/10.1017/S0960258520000471>

Received: 9 January 2020

Revised: 1 December 2020

Accepted: 9 December 2020

**Key words:**

alternating temperatures; banana; crop wild relatives; forest disturbance; gap detection; *Musa*; seed germination

**Correspondence:** Simon Kallow,  
E-mail: [s.kallow@kew.org](mailto:s.kallow@kew.org)

# Regulation of seed germination by diurnally alternating temperatures in disturbance-adapted banana crop wild relatives (*Musa acuminata*)

Simon Kallow<sup>1,2,3</sup> , Rachael Davies<sup>1</sup> , Bart Panis<sup>4</sup> , Steven B. Janssens<sup>3,5</sup>, Filip Vandeloos<sup>3</sup> , Arne Mertens<sup>2,3,5</sup> , Rony Swennen<sup>2,4,6</sup> , Maimun Binti Tahir<sup>7</sup> and John Dickie<sup>1</sup>

<sup>1</sup>Royal Botanic Gardens Kew, Millennium Seed Bank, Wakehurst, Ardingly, Sussex RH17 6TN, UK; <sup>2</sup>Department of Biosystems, Katholieke Universiteit Leuven, Willem de Croylaan 42, Leuven 3001, Belgium; <sup>3</sup>Meise Botanic Garden, Nieuwelaan 38, Meise 1860, Belgium; <sup>4</sup>Bioversity International, Willem de Croylaan 42, Leuven 3001, Belgium; <sup>5</sup>Plant Conservation and Population Biology, Katholieke Universiteit Leuven, Kasteelpark Arenberg 31, Leuven 3001, Belgium; <sup>6</sup>International Institute of Tropical Agriculture, c/o Nelson Mandela African Institution of Science and Technology, Arusha, Tanzania and <sup>7</sup>Malaysian Agricultural Research and Development Institute, Persiaran MARDI-UPM, Serdang 43400, Selangor, Malaysia

**Abstract**

Seed conservation of banana crop wild relatives (*Musa* L. spp.) is limited because of lack of knowledge about their germination ecology. *Musa acuminata* Colla, the most important banana crop wild relative, is distributed in tropical and subtropical Asian and Pacific rainforests and colonizes disturbed sites. The role of temperature in stimulating/inhibiting germination to detect disturbance when canopy gaps are formed is not well known. We assessed seed germination thermal requirements of three subspecies of *M. acuminata* using nine seed accessions which had been stored in the Millennium Seed Bank. Diurnally alternating temperature cycles were almost completely essential for germination compared with constant temperatures. Germination was optimal when the upper temperature of a diurnal cycle was at 35°C; the lower temperature of the cycle was less important. Subspecies occurrence coordinates were used to extract climate temperature data which were then compared against the temperature requirements for germination from our experiment results. Maximum temperatures of the warmest month across subspecies ranges were close to but below optimal germination temperatures, as were diurnal ranges, suggesting soil-warming at the micro-climate level following gap creation is important for *M. acuminata* seed germination. Additionally, pre-treatment for 3 months at 60% relative humidity at constant 25°C improved germination from 14 ± 10 (mean, standard deviation) to 41 ± 29% suggesting a period in the soil seed bank under the canopy may increase sensitivity to alternating temperature cycles. Overall viability was low (49 ± 28%), and considerable variance was caused by the different accessions. Germination remained somewhat inconsistent.

**Introduction**

Multiple biotic and abiotic issues threaten the future of banana production (Ramirez et al., 2011; Ploetz and Evans, 2015), the potential consequences of which will likely have a large impact on the nutrition and livelihoods of many millions of people (Ploetz et al., 2015). The limited gene pool of clonal crops like bananas (Perrier et al., 2011) means they have evolved little resistance to many pathogens, making them particularly vulnerable (Strange and Scott, 2005; McKey et al., 2010). Thus, there is urgent need for the genetic resources present in banana crop wild relatives (CWRs) (*Musa* L. spp.) to be conserved and made available to breeders (Dempewolf et al., 2017).

Storing and accessing genetic resources as seeds is an efficient method for *ex situ* conservation (Li and Pritchard, 2009; Convention on Biological Diversity, 2012; FAO, 2012; Laliberté, 2016). The management of seed genetic resources requires their routine germination for viability monitoring (FAO, 2014), as does accessing plants for phenotyping or breeding; for bananas, low and inconsistent germination rates are a considerable limitation to these (Laliberté, 2016; Brown et al., 2017). Improvement in the ability to germinate banana seeds will therefore have important applications for food security and plant genetic resources conservation (Panis et al., 2020).

There are around 80 species in the genus *Musa* (Govaerts and Häkkinen, 2006); cultivated bananas mostly derive from two of these: *Musa acuminata* Colla and *Musa balbisiana* Colla

© The Author(s), 2021. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution, and reproduction in any medium, provided the original work is properly cited.

(Boonruangrod *et al.*, 2008; De Langhe *et al.*, 2009; Perrier *et al.*, 2009, 2011). Overall, *M. acuminata* is the most important contributor of genetic material for cultivated bananas (Raboin *et al.*, 2005; Martin *et al.*, 2020); for instance, they are the sole contributor to the Cavendish group which accounts for approximately 47% of global production (FAO, 2019). There are ten accepted subspecies (used hereafter to also include varieties and subspecies) of *M. acuminata* (Govaerts and Häkkinen, 2006), which are distributed in Asia, northern Australasia and Melanesia (Perrier *et al.*, 2009).

In contrast to *M. balbisiana* (McGahan, 1961a, 1961b; Stotzky *et al.*, 1961, 1962; Stotzky and Cox, 1962; Bhat *et al.*, 1994), surprisingly little attention has been paid to *M. acuminata* seeds. In the 1950s, Simmonds (1952, 1959) examined both *M. acuminata* and *M. balbisiana* with various non-invasive treatments (looking at ripeness, maturity and moisture content) and invasive treatments (using chipping, scorching, soaking and acid treatments). The physical permeability of *M. acuminata* seed coats to water has received most attention but still remains unresolved (Darjo and Bakry, 1990; Wattanachaiyingcharoen and Turner, 1990; Fortescue and Turner, 2011; Puteh *et al.*, 2011). Significantly, the thermal requirements for germination of *M. acuminata* seeds have not been investigated.

*Musa* spp. are tall forest herbs of tropical to subtropical forests. Their inflorescences are pollinated by bats, birds and probably insects (Itino *et al.*, 1991; Liu *et al.*, 2002), and seeds are dispersed by bats, birds and mammals (Ge *et al.*, 2005; Tang *et al.*, 2005, 2007; Marod *et al.*, 2010). They are observed in disturbed sites such as farm or plantation edges, craggy cliffs and steep mountain gullies, or roadside and track edges. Specifically, *M. acuminata* is known to colonize and dominate disturbed sites (Zhang *et al.*, 2000; Shi *et al.*, 2002; Meng *et al.*, 2012). *Musa* are therefore widely recognized as disturbance-adapted 'jungle weeds' (Simmonds, 1959).

Several adaptations may be expected in *M. acuminata* seeds to facilitate germination following disturbance. Firstly, an environmentally regulated gap detection mechanism may stimulate germination immediately following disturbance and, conversely, prevent germination prior to disturbance (Raich and Khoon, 1990; Vázquez-Yanes and Orozco-Segovia, 1993). For larger-seeded disturbance-adapted species, germination may be stimulated by changes in micro-climate temperature (Pearson *et al.*, 2002; Aud and Ferraz, 2012). This may involve changes in diurnal temperature fluctuations, as soil is exposed to sunlight when a canopy gap forms and therefore warming and cooling fluctuations are altered (Vázquez-Yanes and Orozco-Segovia, 1982; Aud and Ferraz, 2012; Poschlod *et al.*, 2013; Jaganathan, 2018). Sensitivity to differences in alternating temperature range plays an important role in limiting seedling loss during unfavourable conditions (Kos and Poschlod, 2007; Saatkamp *et al.*, 2011). Secondly, the hard seed coat of *Musa* spp. may limit imbibition and delay germination by physical dormancy. This may be relieved according to environmental factors in relation to disturbance, including temperature and moisture regimes, as well as changes in the ecological community such as predators and dispersers. Alternatively, after-ripening or stratification (a pre-treatment period between dispersal and germination) may delay germination immediately after dispersal as observed for some seeds of tropical rainforests (Vázquez-Yanes and Orozco-Segovia, 1993, and references therein).

The aim of this study is to test hypotheses that *M. acuminata* seed germination is regulated in accordance with expectations of

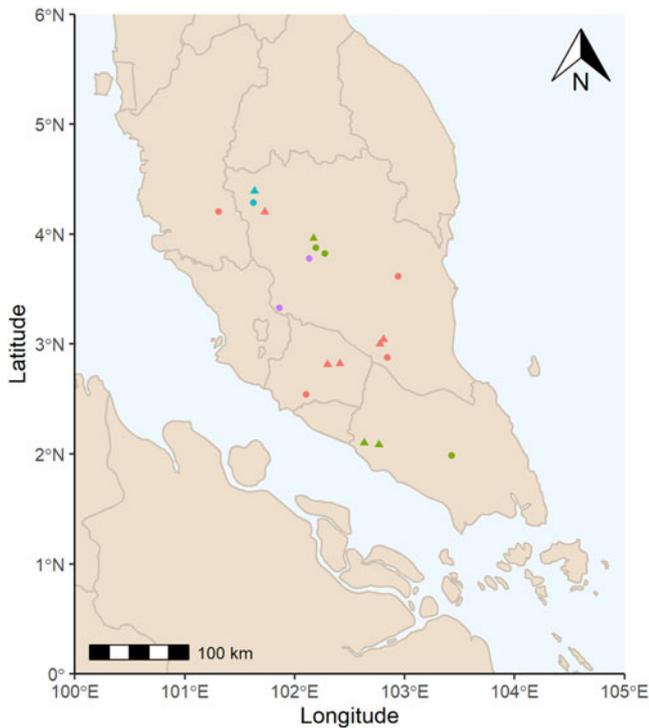
disturbance-adapted tropical species. In particular, we focus on germination responses to temperature, including diurnally alternating temperature regimes as a mechanism for gap detection. Therefore, we (1) describe the morphology and mass of the seeds used in this study, (2) assess whether seed coats are permeable to water, (3) investigate optimal temperature regimes for seed germination, (4) investigate whether stratification relieves dormancy, and finally (5) evaluate the optimal temperature regimes from our germination tests against the macro-climate temperatures of wider distributions of the subspecies under investigation. However, because the seeds used in this study have undergone processes which may influence behaviour, namely drying and sub-zero temperature storage (Baskin *et al.*, 2006), a true definition of germination and dormancy class is not possible, but rather the aim is application to seed conservation.

## Materials and methods

### Plant material

Nineteen seed accessions of four *M. acuminata* subspecies were used for the initial viability assessment (Fig. 1 and Supplementary Table S1). Following the viability assessment described below, nine of these accessions, which consist of three subspecies, were selected for a series of experiments (*M. acuminata* subsp. *acuminata*, *M. acuminata* subsp. *malaccensis* and *M. acuminata* subsp. *microcarpa*; Fig. 1 and Table 1). Seeds were collected from 'Malaysian peninsular rainforest' and 'montane rainforest' ecoregions in the 'tropical and subtropical moist broadleaf forest' biome (Olson *et al.* 2001). Mean annual temperature and mean annual precipitation at the collecting locations are  $26.4 \pm 0.7^\circ\text{C}$  and  $2332 \pm 342$  mm (mean and standard deviation), respectively (Fick and Hijmans, 2017). Seeds were collected in 2015 and 2016 and provided to the Millennium Seed Bank (MSB) by the Malaysian Agricultural Research and Development Institute.

Seeds derive from one or two plants per accession. Populations were wild in origin and occurred either in secondary rainforest, or oil palm/orchard plantation edges. Bunches were only collected from light green to yellow fruits. Fruits from unhealthy or injured plants were avoided. Cut tests were carried out at the time of collection to assess seed maturity, and only bunches with mature seeds were collected. Seeds were considered mature if the endosperm was dry and powdery, as opposed to wet or milky, and embryos were well developed with a mushroom-like capitate shape. Seeds were extracted by hand from ripe fruit on return to the laboratory, and the pulp containing seeds was washed thoroughly using running tap water and sieves, until all the flesh was removed. Unripe fruits were left in the laboratory at room temperature ( $\sim 25^\circ\text{C}$ ) to ripen until they began to yellow and soften and then seeds were removed in the manner described above. After extraction, seeds were air-dried in the laboratory at room temperature and then further dried in a large sealed plastic drum containing silica gel to a maximum of 25% equilibrium relative humidity (eRH). Seeds were then packed in sealed aluminium envelopes and stored for an average of 6 months at  $4^\circ\text{C}$  until air shipped to the MSB. On arrival at the MSB seeds were further dried at 15% eRH and  $18^\circ\text{C}$  in a dry room to approximately 7% moisture content. Seed mass of dried seeds was measured by weighing five replicates of 50 seeds. Seeds were then sealed in airtight glass containers and stored at  $-20^\circ\text{C}$  for 12–18 months prior to use in this study. Before use, selected accessions were removed from cold storage for at least 24 h to



**Fig. 1.** Collecting locations of *M. acuminata* seed accessions. Red = *M. acuminata* subsp. *acuminata*, green = *M. acuminata* subsp. *malaccensis*, blue = *M. acuminata* subsp. *microcarpa*, purple = *M. acuminata* subsp. *truncata*; triangles = selected accessions following viability assessment, circles = accessions not selected. Collection points adjusted to reduce overlap.

equilibrate to 18°C in a dry room at 15% relative humidity (RH). Morphological observations were recorded by dissecting ten seeds per accession and inspecting using a binocular microscope. Seed mass was measured using five samples of 50 seeds.

### Viability

Post-storage seed viability was assessed using two methods to improve estimation methods for this and future studies. The first method was the tetrazolium chloride test (TTC), following the approach of Leist and Krämer (2011). Seeds were imbibed on agar for 3 d at 20°C. Then, a proportion of the testa was removed using a scalpel on two lateral sides to expose the endosperm. Seeds were then soaked in 1% buffered 2,3,5-triphenyl tetrazolium chloride (pH6–8) for 2 d at 30°C in the dark. Staining patterns were then recorded. Embryos that completely stained dark red, or that showed dark red staining at the embryonic axis were considered viable; light pink staining or white embryos were considered unviable. Fifty seeds per accession were tested.

Secondly, embryo rescue (ER) was carried out as a viability test. In a laminar flow, seeds were sterilized by soaking them in 96% ethanol for 3 min, followed by soaking in 1% NaOCl (diluted commercial bleach 5% containing one drop of detergent per 100 ml) for 20 min. Seeds were then rinsed three times in sterilized water. Continuing in the laminar flow with sterile forceps and scalpel, embryos were extracted from seeds. This was done using an incision in the seed coat next to the micropyle and manipulating the seed in order to split the testa, the embryo was then gently removed. Embryos were subsequently transferred onto autoclaved half MS medium (Murashige and Skoog, 1962) in

tubes using long forceps with the haustorium in contact with medium and the embryonic axis upwards. Tubes containing embryos were incubated in the dark at 27°C for 14 d after which they were put in a growth chamber in the light at 27°C for an additional 14 d. Six possible observations were recorded: shoot, callus, blackened colouration, contamination, no change or no embryo as observed during extraction. Ten seeds per accession were tested.

### Imbibition

Seed coat permeability to moisture was tested by assessing mass change of dry seeds during imbibition. Seeds were either left intact and incubated at 25°C, or scarified by removing a sliver of lateral seed coat with a scalpel in order to expose the white seed endosperm ( $3.36 \pm 1.35$  mg) and incubated at 25°C, both with a 12 h light/dark regime. To imbibe seeds, they were placed on agar with the micropyle in contact with 1% agar in Petri dishes (100 × 150 mm). Seeds were weighed individually on days 0, 1, 2, 5, 7 and 21. Prior to measurement, surface moisture and agar were blotted off with tissue paper. Twenty-five seeds from accession *M. acuminata* subsp. *malaccensis* (882899) were used for each treatment. This accession was selected because it had the most seeds. The increase in mass percentage was calculated individually for each seed and plotted against time in order to estimate equilibrium.

### Germination and temperature

Nine accessions were selected for germination tests based on adequate viability and availability (Table 1 and Supplementary Table S1). Incubator temperature regimes were based on the climate at the location of collection. Temperatures were extracted from WorldClim version 2.0 (Fick and Hijmans, 2017) using seed accession collection coordinates to extract climate data at 30 arc seconds resolution. The collecting locations' mean monthly temperature to the nearest 5°C was selected for a constant incubation regime: 25°C; whilst alternating temperatures were based on the monthly maximum and minimum temperatures where the upper temperature was rounded both up and down: 35/20 and 30/20°C. Minimal temperature seasonality occurred in the climate data for the collecting sites (mean coefficient of variability  $1.81 \pm 0.16$ ), so no seasonal differences were included in the design.

Seeds were placed on moist sand (100 g fine silica sand and 14 ml of deionized water) in sealed square plastic boxes (100 × 100 × 20 mm) which were then sealed in clear plastic sealable bags to minimize moisture loss and contamination. These were then placed in the corresponding incubators with 12 h at each temperature. Photoperiod was also on 12 h cycles, light was during the warmer period and darkness during the cooler; light quantity was approximately  $7 \mu\text{mol m}^{-2} \text{s}^{-1}$ . Twenty-three to fifty-three seeds were used in one to three replicates per accession. Sample sizes were constrained by limited seed availability. Germination tests were scored every 7 d, and tests were continued for 6 months to account for potential long sporadic germination times previously observed and to ensure reasonable maximum germination. Germination was defined by radicle emergence. Germinated seeds were removed at scoring.

### Thermogradient

For one accession of *M. acuminata* subsp. *malaccensis* (accession 882899) (the accession with the most seeds), 25 temperature regimes were tested using a thermogradient plate (GRD1, Grant Instruments Ltd, Cambridge, UK). All combinations of constant

**Table 1.** *M. acuminata* seed accessions selected for use in germination tests

Accession	Subspecies	State	Mother plants	Date collected	Date donated
882741	<i>malaccensis</i>	Pahang	1	20/10/2015	23/06/2016
882763	<i>acuminata</i>	Pahang	1	21/10/2015	23/06/2016
882800	<i>acuminata</i>	Negeri Sembilan	2	11/05/2015	23/06/2016
882811	<i>acuminata</i>	Negeri Sembilan	2	11/05/2015	23/06/2016
882833	<i>acuminata</i>	Selangor	1	19/11/2015	23/06/2016
882844	<i>acuminata</i>	Selangor	1	19/11/2015	23/06/2016
882888	<i>malaccensis</i>	Johor	1	30/12/2015	23/06/2016
882899	<i>malaccensis</i>	Johor	1	30/12/2015	23/06/2016
928500	<i>microcarpa</i>	Pahang	1	20/07/2016	31/03/2017

The full set of accessions used for viability testing is in Supplementary Table S1. Date donated is accessioning date on arrival at the MSB.

and alternating temperatures from 15 to 35°C with an interval of 5°C were used. An additional six temperature regimes were tested in incubators to 40°C, with all combinations of 15–40°C under 5°C temperature intervals. Temperature was cycled on a 6/18 hourly basis to allow for a comparison of short and long periods at the warmer/cooler part of the cycles estimated from local temperature data (Meteoblue, 2020). Accordingly, for each temperature regime, there was a sample that had the hotter temperature at 6 h and one at 18 h, except for the samples at 40°C which only had the hotter temperature for 6 h because of a limitation of incubators. Incubation was carried out in the dark because light could not be controlled differently in the thermogradient plate for the short and long cycles, and seeds were previously observed to germinate in the dark. For each condition, 30 seeds were placed on 10 g of silica sand with 8 ml of deionized water in a circular Petri dish (100 × 15 mm), and dishes were sealed with film to avoid moisture loss. An additional 1–2 ml of water was added if the colour of the sand lightened due to water loss. Tests were scored in daylight every 7 d. These germination tests were continued for 70 d.

### Germination and pre-treatment

To simulate the transition of seeds in the soil seed bank before and after gap formation, seeds were incubated ('stratified') at constant 25°C and then moved to alternating 30/20°C after 3 months. There were two levels of stratification at 25°C, seeds were either placed on moist sand as described above (at 100% RH), or suspended in sealed jars over lithium chloride solutions controlled to be at 60% RH (60 g LiCl in 200 ml deionized water), for these a humidity meter was placed into the jar with the seeds for 7 d to ensure the correct RH. After 3 months, seeds were then incubated for three further months at 30/20°C light/dark as previously described. Scoring of germination tests was carried out as previously described. All germination tests were carried out at the MSB.

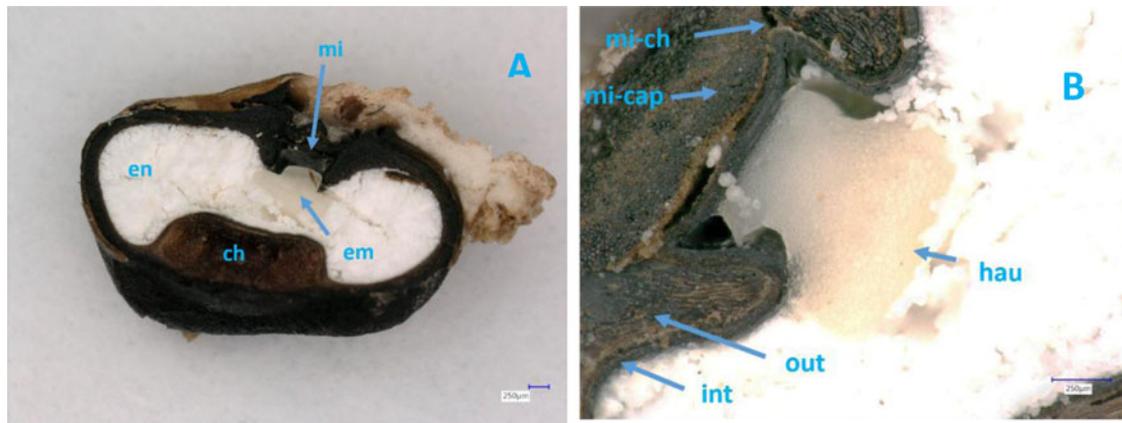
### Macro-climate assessment

We composed a dataset of occurrence records of wild *M. acuminata* subspecies from 16 sources including recent field missions (Supplementary Fig. S1 and Supplementary Table S2). Accurate locality descriptions without coordinates were

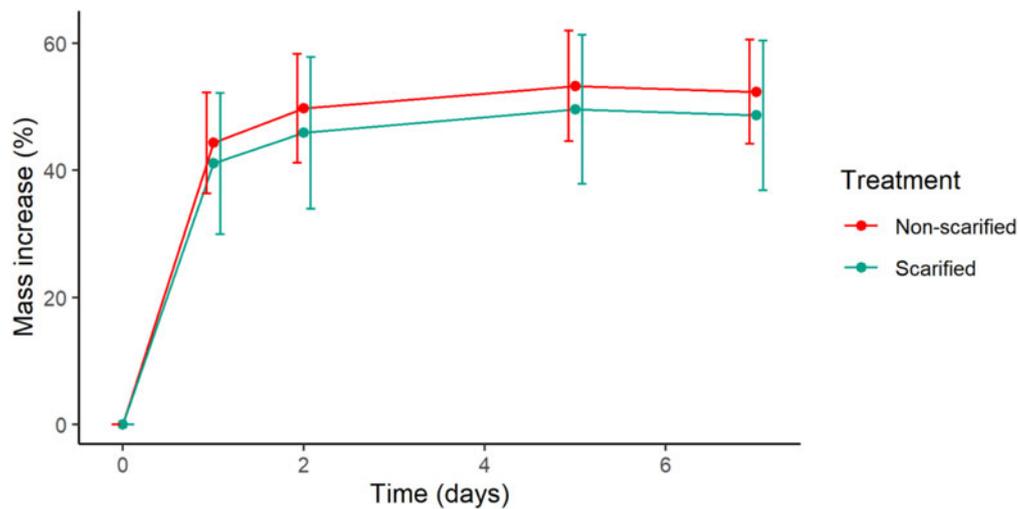
georeferenced using Google Earth pinpoints (Google Earth, 2018). Duplicate records, outliers and zero coordinates were detected and removed with the online tool CoordinateCleaner (Zizka *et al.*, 2019). This resulted in a dataset consisting of 222 occurrence records of nine (of the ten) *M. acuminata* subspecies. Coordinates from these were used to extract temperature-related bioclimatic data from WorldClim version 2.0 (Fick and Hijmans, 2017) to compare to the optimal germination temperatures from the previous germination tests, and to look for variation between subspecies. Additionally, precipitation across subspecies distributions was extracted to assess further climate variation by taxa.

### Statistical analysis

Seed mass increase during imbibition was plotted against time, and a point was selected where equilibrium with the environment was reached across treatments. Percentage mass increase at this point for the two treatments was compared using a two-sample *t*-test. Descriptive indices were calculated for the germination data in the R package GerminaR (Isla *et al.*, 2019). These included germination percentage (Labouriau and Valadares, 1983), mean germination rate defined as the reciprocal of the average time to germination (Ranal and Santana, 2006), mean time to germination (Czabator, 1962) and synchrony index (Primack, 1980; Ranal and Santana, 2006). Generalized linear models (GLMs) were made to analyze final germination with binary responses. Models had quasibinomial error structure (to account for overdispersion) and logit link functions. Overdispersion of models were assessed by simulating residuals from the fitted models and comparing the standard deviation of the simulated and actual data using the DHARMA package in R (Hartig, 2019). Maximum models, with fixed factors of accession, subspecies, viability and a random factor of temperature treatment, were fitted. Minimum adequate models were achieved by removing factors according to ANOVA and Chi-squared tests. *Post hoc* contrast analysis was carried out between treatments and controls using multiple comparisons of means and Tukey contrasts. In all cases, effective sample sizes were used which corrected sample sizes according to the viability estimates from the TTC test result. This viability measure was used because it was not possible to correlate the two viability measures (ER and TTC) and the sample size used in the TTC was greater than for ER (50 rather than



**Fig. 2.** (A) Longitudinal section of a *M. acuminata* seed, showing micropyle (mi), embryo (em), chalazal mass (ch) and endosperm (en). (B) Detail showing micropyle cap (mi-cap), micropyle channel (mi-ch), haustorium (hau), inner integument (int) and outer integument (out). Image taken with Keyence VHX5000. Images: S. Kallow.



**Fig. 3.** Increase in seed mass of scarified and non-scarified *M. acuminata* subsp. *malaccensis* seeds during imbibition from 7 to 36% moisture content. Seeds weighed individually ( $n = 25$ ). Bars represent standard deviation.

10 seeds). All statistical analysis was carried out using R v 3.6.0 (R Core Team, 2019).

## Results

### Morphology and seed mass

Observations showed that *M. acuminata* seeds have two chambers within a two-layer integument (Fig. 2A). The larger first chamber contains the embryo and endosperm. Embryos are undifferentiated and capitate, 1–2 mm in size and the embryonic axis extends into the micropylar collar (Fig. 2B). The cotyledonary haustorium of the embryo extends below the embryonic axis and is surrounded by a powdery white endosperm. Above the embryo is a micropyle, which is filled by a micropyle plug and forms an operculum. The second chamber consists of a chalazal mass. Seed mass was  $40.45 \pm 6.80$  mg (mean and standard deviation, used from hereon).

### Viability

Viability was  $49 \pm 28\%$  according to the TTC test and  $28 \pm 29\%$  for the embryo recue (i.e. the 'shoot' category; Supplementary Table S1). No linear relationship could be found between the two parameters of viability ( $r^2 = 0.055$ ,  $P = 0.811$ ). Embryos that did not germinate mainly remained white (Supplementary Fig. S2). This means that viability was low because embryos were most likely dead, rather than infested by insects, fungal/bacterial contaminated or lacking embryos. The combined results of the viability tests allowed us to select the nine accessions used for germination tests.

### Imbibition

During imbibition, both scarified and non-scarified seeds rapidly increased mass in 1 d and reached equilibrium after 5 d or earlier (Fig. 3). The difference in percentage mass increase was not statistically significant between scarified ( $49.6 \pm 11.8\%$ ) and non-scarified seeds ( $53.3 \pm 8.7\%$ , two-sample  $t$ -test,  $t = 1.1189$ ,  $df = 33.145$ ,  $P = 0.27$ ). The moisture content of non-scarified seeds increased from 7% on day 0 to 36% after 21 d imbibition.

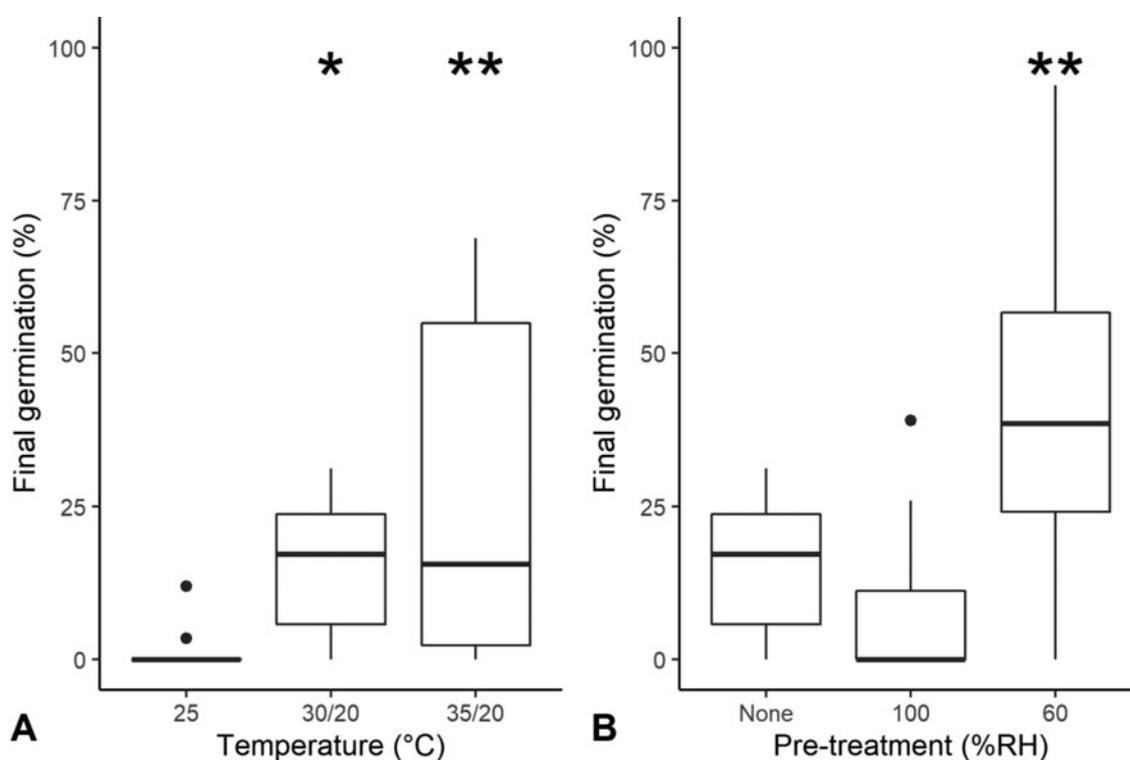
**Table 2.** Indices from germination results from selected *M. acuminata* accessions

Treatment (°C)	Final germination (%)		Time to germination (d)		Germination rate		Synchrony index	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
25	1.19	3.40	85	67.88	0.02	0.01	1.00	NA
30/20	14.38	10.20	41.17	43.87	0.04	0.01	0.57	0.36
35/20	25.63	27.47	55.69	39.18	0.03	0.01	0.39	0.32
100% RH <sup>a</sup>	8.22	12.79	41.44	46.90	0.05	0.03	0.72	0.42
60% RH <sup>a</sup>	41.54	29.26	25.62	12.06	0.04	0.01	0.51	0.21
60% RH control <sup>b</sup>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00

Germination time is mean time to germination in days (Czabator, 1962), germination rate defined as the reciprocal of the average time to germination (Ranal and Santana, 2006), and synchrony index defined by Primack (1980) and Ranal and Santana (2006). Alternating temperatures were for 12 h thermo and photo cycles. Germination tests were continued for 6 months. Results were corrected to take into account viability assessment with previous TTC test ( $n = 23\text{--}53$  seeds in 1–3 replicates).

<sup>a</sup>Pre-treated by incubation at constant 25°C at two levels of RH for 3 months prior to 30/20°C incubation for 3 months at 100% RH.

<sup>b</sup>Incubated only at constant 25°C.



**Fig. 4.** Final germination percentages after 6 months incubation at different conditions for nine *M. acuminata* accessions (subsp. *acuminata*, *malaccensis* and *microcarpa*). (A) Alternating temperature regimes compared with control of constant temperature (30/20,  $P = 0.031$ ; 35/20,  $P = 0.006$ ). (B) Pre-treatment for 3 months at 25°C (60% RH,  $P = 0.007$ ) prior to incubation for 3 months at 30/20°C. Incubation was on moist sand (100% RH) unless otherwise stated. Alternating temperatures (30/20 or 35/20°C) were on 12 hourly cycles. Stars indicate  $P$ -values ( $* = <0.05$ ,  $** = <0.01$ ) from a GLM with quasibinomial error structure and logit link using the number of seeds germinated and the number of seeds that did not germinate, against the control. Final germination percentages are corrected to take into account viability assessment with previous TTC test ( $n = 23\text{--}53$  seeds in 1–3 replicates).

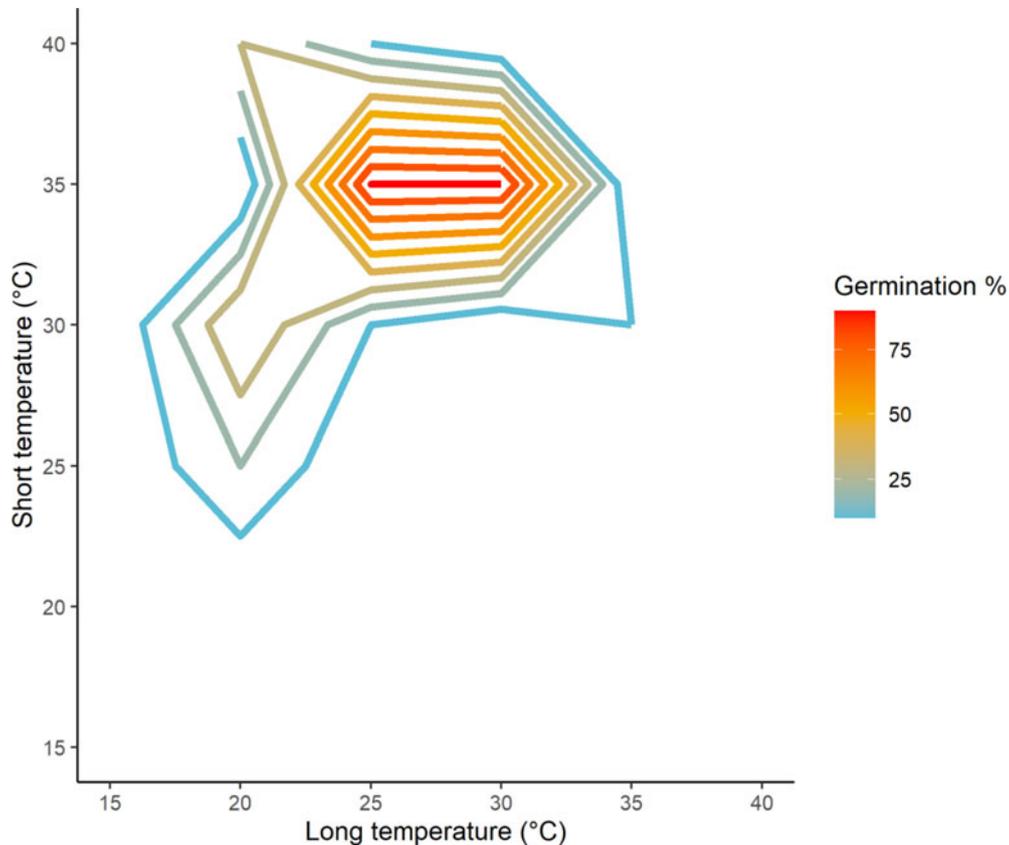
### Germination and temperature

Seeds germinated to higher levels at alternating temperatures compared with constant temperature (Table 2 and Fig. 4). Optimal germination was at 35/20°C ( $P = 0.006$ ) followed by 30/20°C ( $P = 0.031$ ) compared with constant temperature at 25°C; however, there was no significant difference between 35/20 and 30/20 ( $P = 0.164$ ) following multiple comparison of means using Tukey contrasts. The minimum adequate

GLM included only treatment temperatures, as germination was highly variable between accessions in all tests. Mean time to germination under optimal temperature (35/20°C) was 56 d, and this was highly variable ( $\pm 39$  d).

### Thermogradient

Maximum germination occurred equally at 35/25°C (6/18 h) and 35/30°C, followed by 30/20°C (90 and 40%, respectively; Fig. 5).



**Fig. 5.** Contour plot of final germination percentage for *M. acuminata* subsp. *malaccensis* (accession 882899) following incubation at all temperature combinations at 5°C intervals between 15 and 40°C, short temperature was for 6 h and long for 18 h in 24-h cycles, using a thermogradient plate. Results were after 70 d. Germination percentages corrected to take into account estimated viability ( $n = 30$  for each temperature combination).

No germination occurred at any constant temperature, or at temperatures below 20°C. There was no germination when the higher temperature was for the longer period (18 h), apart from at 30/35°C. In the GLM, only the short hotter temperature had a significant positive effect on the number of seeds germinated ( $P < 0.001$ ), and all other parameters (long temperature, the average temperature or the temperature differential) could be removed from the model without reducing the explanatory power.

#### Pre-treatment

Pre-treating the seeds for 3 months at 60% RH increased germination to  $41 \pm 29\%$  from  $14 \pm 10\%$  ( $P = 0.007$ ; Table 2 and Fig. 4), but the same treatment at 100% RH did not have an effect ( $P = 0.423$ ). The minimal adequate model only included the treatment, and there was therefore no discernible effect of subspecies.

#### Macro-climate

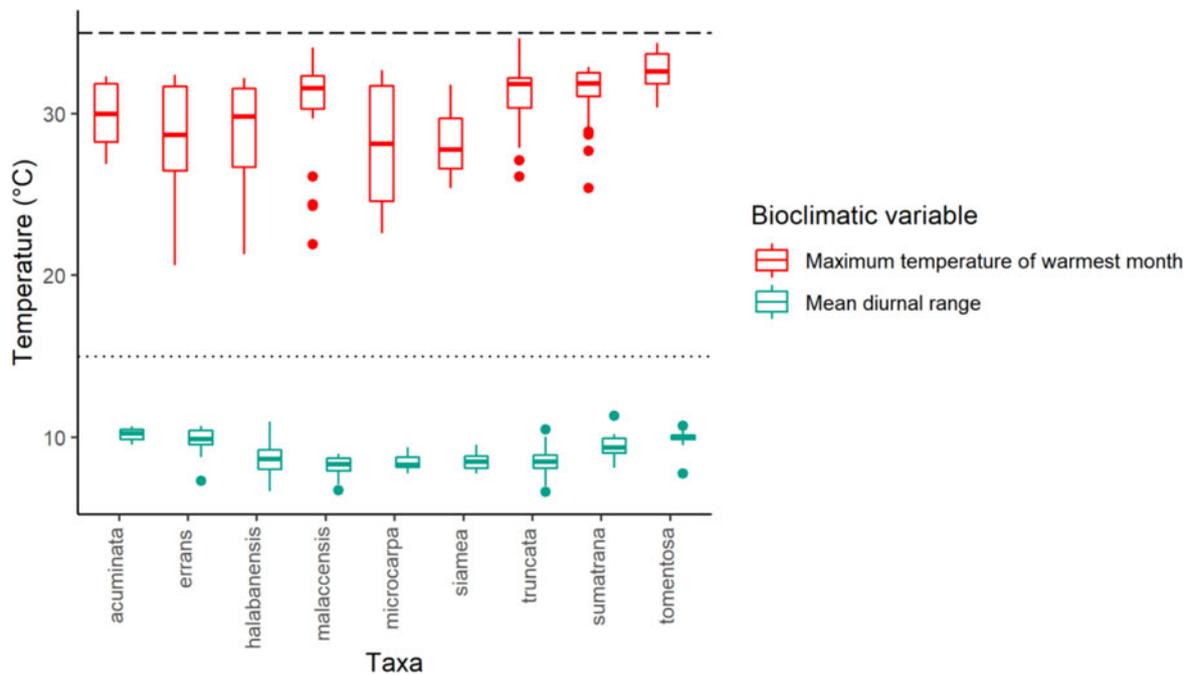
Based on our combined germination test results compared with temperature extracted from climate data, we see that the germination temperature requirements are warmer and have a wider diurnal range than estimated from subspecies distributions (Fig. 6). For instance, the temperature of the hotter part of the diurnal cycle from the germination experiments was 35°C; this is close to but warmer than the maximum temperature of the warmest month for all taxa. There were no records at all as high as 35°C, and a few records were close.

For diurnal range, no records were as high as the 15°C we observed from the germination test results. Mean diurnal range across taxa distributions was  $8 \pm 1^\circ\text{C}$  (Fig. 6), and the widest range was 11°C.

#### Discussion

Stored *M. acuminata* seeds had an almost absolute requirement for alternating temperatures. Within a seed accession, there was non-uniform sensitivity to alternating temperatures, such that by pre-treating seeds with a period of constant temperature, higher final germination percentages were achieved than without such a pre-treatment. Seeds had hard coats, but we found that these were permeable to water.

Our results were comparable to many tropical disturbance-adapted species (reviewed by Vázquez-Yanes and Orozco-Segovia, 1993; Baskin and Baskin, 2014). Within the Musaceae, *M. acuminata* thermal requirements were similar to *M. balbisiana* (Stotzky et al., 1961; Stotzky and Cox, 1962). In these studies, there was also an almost absolute requirement for alternating temperatures, with maximum germination at 35°C for 5 h (mean 59%), and low temperature was 15°C for 19 h (mean 70%). Additionally, low levels of germination of *M. balbisiana* occurred following short and even singular exposure to alternating temperature, but continuous cycles were required for maximum germination (Stotzky and Cox, 1962). As a combination, germination was optimal (80%) at 5 h of 35°C and 19 h of 12°C. The cooler lower temperature requirement for *M. balbisiana* reflects



**Fig. 6.** Bioclimatic variables at occurrence locations for *M. acuminata* taxa extracted from WorldClim 2.0 (Fick and Hijmans, 2017). Dashed line represents optimal temperature for the warm part of the diurnal cycle (35°C) from the germination test results; dotted line represents optimal diurnal range (15°C) from the germination test results.

the subtropical and at times higher elevation distribution of *M. balbisiensis* – East India to Yunnan, China.

At the macro-climate scale, temperature and diurnal ranges were rather similar across subspecies distributions (Fig. 6 and Supplementary Fig. S3). In general, temperatures estimated from the climate model (WorldClim) were cooler than in our germination tests; diurnal range was also smaller than may be optimal for germination. This suggests that micro-climate, rather than solely macro-climate, is important in germination of *M. acuminata*. This is because soil temperature, when exposed to direct sunlight, may heat considerably more than the air temperature usually used in climate data and modelling. Furthermore, this observation supports our expectations that seeds are adapted to specifically detect gaps in the forest canopy following disturbance.

Specific optimum temperature fluctuations were different between our two experiments:  $5.7 \pm 3.9^\circ\text{C}$  for the thermogradient plate experiment and  $15^\circ\text{C}$  in the other germination tests, suggesting a degree of plasticity in fluctuation requirement. Pearson *et al.* (2002) proposed four categories of response to alternating temperature dynamics according to seed mass. Using these same categories, our results place *M. acuminata* into category three, where there is a positive response to increasing temperature fluctuation in the range of  $0\text{--}16.7^\circ\text{C}$ , but there is no dramatic optimum or cut-off point. For this group, seed mass was the heaviest category in their sample ( $20.9 \pm 14.2$  mg, mean and standard error), *M. acuminata* seeds were even heavier than this ( $40.45 \pm 6.80$  mg).

At the micro-climate scale, soil mean temperature and diurnal fluctuation are dependent on several factors, including topography, canopy, litter, air temperature and solar irradiance (Kang *et al.*, 2000; Saatkamp *et al.*, 2011; Bilgili *et al.*, 2013). Additionally, mean temperature is correlated to distance from

the edge as well as the size of the gap (Pearson *et al.*, 2002; Saner *et al.*, 2009; Takada *et al.*, 2015). Temperature fluctuations are also dependent on the burial depth of the seed in the soil (Pearson *et al.*, 2002). Finally, the composition of the forest effects both the mean temperature and range of the diurnal fluctuation, for instance, *M. acuminata* seeds would be inhibited from germinating in the temperatures of old growth forests, but may occur in oil palm plantations (Hardwick *et al.*, 2015).

Our results show that, after storage, most seeds from the same inflorescence are non-dormant but some seeds are; germination is increased following a period of stratification or after-ripening. Heterogeneity in dormancy could be part of a bet-hedging strategy whereby seeds have variable levels of dormancy to aid dispersal and maximize seedling establishment (Ng, 1980; Tielborger *et al.*, 2012; Gremer *et al.*, 2016).

We found *M. acuminata* seed coats did not limit imbibition. However, for seeds that have not been dried and frozen, physical dormancy cannot be completely ruled out; especially as drying increases imbibition rates in *Musa* seeds (Puteh *et al.*, 2011). Furthermore, *Musa* seeds clearly invest in physical defences, which may correlate with physical dormancy. In other species, alternating temperatures have also demonstrated removal of physical dormancy (De Souza *et al.*, 2012; Jaganathan *et al.*, 2019), and this in relation to disturbance (Jaganathan, 2018).

Much of the variation in our results was the result of the variation between accessions. This is despite seeds being treated and collected in the same way. Additionally, the overall viability of our accessions was low, and *Musa* seeds have been shown to have variable levels of desiccation sensitivity depending on species and maturity at collection (Kallow *et al.*, 2020). The loss of viability observed here may also be the result of immature seeds not having fully developed desiccation tolerance (Ellis *et al.*, 1991;

Hay and Probert, 1995; Leprince et al., 2017). Seed maturity within and between bunches can be important for traits such as desiccation tolerance and germination potential. Despite our efforts, it is possible that immature seeds were collected during field expeditions and this may help explain why it is then difficult to obtain consistent results for seed germination. When collecting bananas in the wild, it is difficult to access mature seeds as it is rare to find ripe or mature bunches in the forest, presumably because of predation and frugivory.

*M. acuminata* seeds demonstrate sensitivity to alternating temperatures for seed germination suitable to detect gaps in forest canopies following disturbance. These results can directly be applied to the management of banana seeds for conservation, and to more easily access material for phenotyping and breeding.

**Supplementary material.** To view supplementary material for this article, please visit: <https://doi.org/10.1017/S0960258520000471>.

**Acknowledgements.** The authors thank the Malaysian Agricultural Research and Development Institute for providing seeds, in particular Anuar Rasyidi, M. N., Ahmad Syahman, M. D., Mohd Shukri, M. A., Suryanti, B. At Royal Botanic Gardens, Kew, thanks the Seed Collections team at the Millennium Seed Bank for curating the seeds and carrying out the Tetrazolium Chloride tests; Kirstine Manger and Kay Singleton for assistance with the germination tests; John Adams and Pablo Barreiro for thermogradient plate and incubator set up. At KU, Leuven thanks for Tom Vanderstraeten and Kevin Longin for the embryo rescue. The authors are also grateful to all donors who supported this work through their contributions to the CGIAR Fund (<https://www.cgiar.org/funders/>) and in particular to the CGIAR Research Program for Roots, Tubers and Bananas (CRP-RTB).

**Financial support.** This work was funded as a subgrant from the University of Queensland from the Bill & Melinda Gates Foundation project 'BBTV mitigation: Community management in Nigeria, and screening wild banana progenitors for resistance' [OPP1130226]. The Royal Botanic Gardens Kew is part supported by a grant in aid from the UK Department of Environment Food and Rural Affairs.

#### Author contributions.

Conceptualization: S.K., R.D. and J.D.; methodology: S.K., R.D. and J.D.; software: S.K.; validation: S.K., R.D., B.P. and F.V.; formal analysis: S.K.; investigation: S.K. and R.D.; resources: B.P., S.B.J., A.M., R.S., M.B.T. and J.D.; data curation: S.K., R.D. and A.M.; writing original draft: S.K.; writing reviewing and editing: S.K., B.P., S.B.J., F.V., R.S. and J.D.; visualization: S.K.; supervision: B.P., S.B.J., R.S. and J.D.; project administration: J.D.; funding acquisition: B.P., S.B.J., R.S. and J.D.

**Conflict of interest.** None.

## References

- Aud FF and Ferraz IDK** (2012) Seed size influence on germination responses to light and temperature of seven pioneer tree species from the central Amazon. *Anais da Academia Brasileira de Ciencias* **84**, 759–766.
- Baskin CC and Baskin JM** (2014) *Seeds: ecology, biogeography and evolution of dormancy and germination*. San Diego, CA, Academic Press.
- Baskin CC, Thompson K and Baskin JM** (2006) Mistakes in germination ecology and how to avoid them. *Seed Science Research* **16**, 165–168.
- Bhat SR, Bhat KH and Chandel KPS** (1994) Studies on germination and cryopreservation of *Musa balbisiana* seed. *Seed Science and Technology* **22**, 637–640.
- Bilgili M, Sahin B and Sangun L** (2013) Estimating soil temperature using neighboring station data via multi-nonlinear regression and artificial neural network models. *Environmental Monitoring and Assessment* **185**, 347–358.
- Boonruangrod R, Desai D, Fluch S, Berenyi M and Burg K** (2008) Identification of cytoplasmic ancestor gene-pools of *Musa acuminata* Colla and *Musa balbisiana* Colla and their hybrids by chloroplast and mitochondrial haplotyping. *Theoretical and Applied Genetics* **118**, 43–55.
- Brown A, Tumuhimbise R, Amah D, Uwimana B, Nyine M, Mduma H, Talengera D, Karamura D, Kuriba J and Swennen R** (2017) Bananas and plantains (*Musa* spp.), pp. 219–240 in Campos H and Caligari PDS (Eds.) *Genetic improvement of tropical crops*. Switzerland, Cham, Springer.
- Convention on Biological Diversity** (2012) *Global strategy for plant conservation: 2011–2020*. Richmond, UK, Botanic Gardens Conservation International.
- Czabator FJ** (1962) Germination value: an index combining speed and completeness of pine seed germination. *Forest Science* **8**, 386–396.
- Darjo P and Bakry F** (1990) Conservation and germination of banana seeds. *Fruits* **45**, 103–113.
- De Langhe E, Vrydaghs L, De Maret P, Perrier X and Denham T** (2009) Why bananas matter: an introduction to the history of banana domestication. *Ethnobotany Research and Applications* **7**, 165–177.
- Dempewolf H, Baute G, Anderson J, Kilian B, Smith C and Guarino L** (2017) Past and future use of wild relatives in crop breeding. *Crop Science* **57**, 1070–1082.
- De Souza TV, Voltolini CH, Santos M and Paulilo MTS** (2012) Water absorption and dormancy-breaking requirements of physically dormant seeds of *Schizolobium parahyba* (Fabaceae – Cesalpinioideae). *Seed Science Research* **22**, 169–176.
- Ellis RH, Hong TD and Roberts EH** (1991) An intermediate category of seed storage behaviour. 2, Effects of provenance, immaturity and imbibition on desiccation-tolerance in coffee. *Journal of Experimental Botany* **42**, 653–657.
- FAO** (2012) *Second global plan of action for plant genetic resources for food and agriculture in Commission on Genetic Resources for Food and Agriculture*. Rome, Italy, Commission on Genetic Resources for Food and Agriculture.
- FAO** (2014) *Genebank standards for plant genetic resources for food and agriculture*. Rome, Italy.
- FAO** (2019) *Banana facts and figures*. Retrieved from: <http://www.fao.org/economics/est-est-commodities/bananas/bananafacts/en/#.XtpQTigzY2w>.
- Fick SE and Hijmans RJ** (2017) WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. *International Journal of Climatology* **25**, 1965–1978.
- Fortescue J and Turner D** (2011) Reproductive Biology. in M Pillay and A Tenkouano (eds), *Banana Breeding Progress and Challenges*. vol. 1, CRC Press, Florida, pp. 145–180.
- Ge XJ, Liu MH, Wang WK, Schaal BA and Chiang TY** (2005) Population structure of wild bananas, *Musa balbisiana*, in China determined by SSR fingerprinting and cpDNA PCR-RFLP. *Molecular Ecology* **14**, 933–944.
- Google Earth** (2018) GOOGLE EARTH. Available at: <http://www.google.com/earth/index.html>.
- Govaerts R and Häkkinen M** (2006) *World checklist of Musaceae*. Kew, Facilitated by the Royal Botanic Gardens. Available at: <http://wcp.science.kew.org/> (retrieved 25 June 2020).
- Gremer JR, Kimball S and Venable DL** (2016) Within-and among-year germination in Sonoran Desert winter annuals: bet hedging and predictive germination in a variable environment. *Ecology Letters* **19**, 1209–1218.
- Hardwick SR, Toumi R, Pfeifer M, Turner EC, Nilus R and Ewers RM** (2015) The relationship between leaf area index and micro-climate in tropical forest and oil palm plantation: forest disturbance drives changes in micro-climate. *Agricultural and Forest Meteorology* **201**, 187–195.
- Hartig F** (2019) DHARMA: RESIDUAL DIAGNOSTICS FOR HIERARCHICAL (MULTI-LEVEL/MIXED) REGRESSION MODELS. R package version 0.3.2.0. Available at: <http://florianhartig.github.io/DHARMA/>.
- Hay FR and Probert RJ** (1995) Seed maturity and the effects of different drying conditions on desiccation tolerance and seed longevity in foxglove (*Digitalis purpurea* L.). *Annals of Botany* **76**, 639–647.
- Isla FL, Alfaro OB, de Santana DG, Ranal MA and Pompelli MF** (2019) GERMINAR: GERMINATION INDICES AND GRAPHICS FOR SEED GERMINATION IN ECOPHYSIOLOGICAL STUDIES. R package version 1.4.2. Available at: <https://CRAN.R-project.org/package=Germinar>.
- Itino T, Kato M and Hotta M** (1991) Pollination ecology of the two wild bananas, *Musa acuminata* subsp. *halabanensis* and *M. salaccensis* – chiropterophily and ornothophily. *Biotropica* **23**, 151–158.
- Jaganathan GK** (2018) Physical dormancy alleviation and soil seed bank establishment in *Cassia roxburghii* is determined by soil microsite characteristics. *Flora* **244**, 19–23.

- Jaganathan GK, Han YY, Song DP, Selvam P and Liu BL (2019) Maternal and bural environment determine physical dormancy release in tropical *Senna auriculata* (Fabaceae) seeds. *Journal of Forestry Research* **31**, 1343–1351.
- Kallow, S, Longin, K, Fanega Sleziak, N, Janssens, SB, Vandeloek, F, Dickie, J, Swennen, R, Paofa, J, Carpentier, S and Panis, B (2020) Challenges for *ex situ* conservation of wild bananas: seeds collected in Papua New Guinea have variable levels of desiccation tolerance. *Plants* **9**, 1243.
- Kang S, Kim S, Oh S and Lee D (2000) Predicting spatial and temporal patterns of soil temperature based on topography, surface cover and air temperature. *Forest Ecology and Management* **136**, 173–184.
- Kos M and Poschold P (2007) Seeds use temperature cues to ensure germination under nurse-plant shade in xeric Kalahari savannah. *Annals of Botany* **99**, 667–675.
- Labouriau LG and Valadares MEB (1983) *The germination of seeds*. Washington, DC, OEA.
- Laliberté B (2016) Global strategy for the conservation and use of *Musa* (banana) genetic resources: a consultative document prepared by the Global Musa Genetic Resources Network (MusaNet). Rome, Bioversity International.
- Leist N and Krämer S (2011) *Supplements 2011 to ISTA working sheets on tetrazolium testing*. ISTA.
- Leprince O, Pellizzaro A, Berriri S and Buitink J (2017) Late seed maturation: drying without drying. *Journal of Experimental Botany* **68**, 827–841.
- Li DZ and Pritchard DW (2009) The science and economics of *ex situ* plant conservation. *Trends in Plant Science* **14**, 614–621.
- Liu AZ, Li DZ, Wang H and Kress WJ (2002) Ornithophilous and chiropterophilous pollination in *Musa itinerans* (Musaceae), a pioneer species in tropical rain forests of Yunnan, southwestern China. *Biotropica* **34**, 254–260.
- Marod D, Pinyo P, Duengkai P and Hiroshi T (2010) The role of wild banana (*Musa acuminata* Colla) on wildlife diversity in mixed deciduous forest, Kanchanaburi Province, Western Thailand. *Kasetsart Journal, Natural Sciences* **44**, 35–43.
- Martin G, Cardi C, Sarah G, Ricci S, Jenn C, Fondi E, Perrier X, Glaszmann JC, D'Hont A and Yahiaoui N (2020) Genome ancestry mosaics reveal multiple and cryptic contributors to cultivated banana. *Plant Journal* **102**, 1008–1025.
- McGahan MW (1961a) Studies on the seed of banana. I. Anatomy of the seed and embryo of *Musa balbisiana*. *American Journal of Botany* **48**, 230–238.
- McGahan MW (1961b) Studies on the seed of banana. II. The anatomy and morphology of the seedling of *Musa balbisiana*. *American Journal of Botany* **48**, 630–637.
- McKey D, Elias M, Pujol B and Duputie A (2010) The evolutionary ecology of clonally propagated domesticated plants. *New Phytologist* **186**, 318–332.
- Meng LZ, Gao XX, Chen J and Martin K (2012) Spatial and temporal effects on seed dispersal and seed predation of *Musa acuminata* in southern Yunnan, China. *Integrative Zoology* **7**, 30–40.
- Meteoblue (2020) WEATHER CLOSE TO YOU. Available at: [meteoblue.com/](https://meteoblue.com/) (accessed 25 June 2020).
- Murashige T and Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. *Physiologia Plantarum* **15**, 473–497.
- Ng FSP (1980) Germination ecology of Malaysian woody plants. *Malaysian Forester* **43**, 406–437.
- Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GV, Underwood EC, D'Amico JA, Itoua I, Strand HE, Morrison JC and Loucks CJ (2001) Terrestrial Ecoregions of the World: A New Map of Life on Earth A new global map of terrestrial ecoregions provides an innovative tool for conserving biodiversity. *BioScience* **51**, 933–938.
- Panis B, Kallow S and Janssens SB (2020) Seed germination, preservation and population genetics of wild *Musa* germplasm in Kema GHK and Drenth A (Eds.) *Achieving sustainable cultivation of bananas v2: germplasm and genetic improvement*. Cambridge, UK, Burleigh Dodds.
- Pearson TRH, Burslem D, Mullins CE and Dalling JW (2002) Germination ecology of neotropical pioneers: interacting effects of environmental conditions and seed size. *Ecology* **83**, 2798–2807.
- Perrier X, Bakry F, Carreel F, Jenny C, Horry JP, Lebot V and Hippolyte I (2009) Combining biological approaches to shed light on the evolution of edible bananas. *Ethnobotany Research and Applications* **7**, 199–216.
- Perrier X, De Langhe E, Donohue M, Lentfer C, Vrydaghs L, Bakry F, Carreel F, Hippolyte I, Horry JP, Jenny C, Lebot V, Risterucci AM, Tomekpe K, Doutrelepont H, Ball T, Manwaring J, de Maret P and Denham T (2011) Multidisciplinary perspectives on banana (*Musa* spp.) domestication. *Proceedings of the National Academy of Sciences USA* **108**, 11311–11318.
- Ploetz RC and Evans EA (2015) The future of global banana production, pp. 311–351 in Janick J (Ed.) *Horticultural reviews*. Hoboken, NJ, John Wiley and Sons.
- Ploetz RC, Kema GH and Ma LJ (2015) Impact of diseases on export and smallholder production of banana. *Annual Review of Phytopathology* **53**, 269–288.
- Poschold P, Abedi M, Bartelheimer M, Drobnik J, Rosbakh S and Saatkamp A (2013) Seed ecology and assembly rules in plant communities, pp. 164–202 in van der Maarel E and Franklin J (Eds.) *Vegetation ecology* (2nd edn.). Chichester, Wiley.
- Primack RB (1980) Variation in the phenology of natural populations of montane shrubs in New Zealand. *The Journal of Ecology* **68**, 862.
- Puteh AB, Aris EM, Sinniah UR, Rahman M, Mohamad RB and Abdullah NAP (2011) Seed anatomy, moisture content and scarification influence on imbibition in wild banana (*Musa acuminata* Colla) ecotypes. *African Journal of Biotechnology* **10**, 14373–14379.
- R Core Team (2019) *R: a language and environment for statistical computing*. Vienna, Austria, R Foundation for Statistical Computing.
- Raboin LM, Carreel F, Noyer JL, Baurens FC, Horry JP, Bakry F, Montcel HTD, Ganry J, Lanaud C and Lagoda PJJ (2005) Diploid ancestors of triploid export banana cultivars: molecular identification of 2n restitution gamete donors and n gamete donors. *Molecular Breeding* **16**, 333–341.
- Raich JW and Khoon GW (1990) Effects of canopy openings on tree seed germination in a Malaysian dipterocarp forest. *Journal of Tropical Ecology* **6**, 203–217.
- Ramirez J, Jarvis A, Van den Bergh I, Staver C and Turner D (2011) Changing climates: effects on growing conditions for banana and plantain (*Musa* spp.) and possible responses. *Crop Adaptation to Climate Change* **19**, 426–438.
- Ranal MA and Santana DGD (2006) How and why measure the germination process? *Revista Brasileira de Botanica* **29**, 1–11.
- Saatkamp A, Affre L, Baumberger T, Dumas PJ, Gasmí A, Gachet S and Arene F (2011) Soil depth detection by seeds and diurnally fluctuating temperatures: different dynamics in 10 annual plants. *Plant and Soil* **349**, 331–340.
- Saner P, Lim R, Burla B, Ong RC, Scherer-Lorenzen M and Hector A (2009) Reduced soil respiration in gaps in logged lowland dipterocarp forests. *Forest Ecology and Management* **258**, 2007–2012.
- Shi JP, Zhang GM, Bai KJ and Tang JW (2002) The effects of human disturbance on biomass and plant diversity of *Musa acuminata* community. *Journal of Wuhan Botanical Research* **20**, 119–123.
- Simmonds NW (1952) The germination of banana seeds. *Tropical Agriculture* **29**, 35–49.
- Simmonds NW (1959) Experiments on the germination of banana seeds. *Tropical Agriculture* **36**, 259–273.
- Stotzky G and Cox EA (1962) Seed germination studies in *Musa*. II. Alternating temperature requirement for the germination of *Musa balbisiana*. *American Journal of Botany* **49**, 763–770.
- Stotzky G, Cox E and Goos R (1961) Alternating temperature requirements for the germination of *Musa balbisiana* Colla seeds. *Plant Physiology* **36**, 21–22.
- Stotzky G, Cox EA and Goos RD (1962) Seed germination studies in *Musa*. I. Scarification and aseptic germination of *Musa balbisiana*. *American Journal of Botany* **45**, 515–520.
- Strange RN and Scott PR (2005) Plant disease: a threat to global food security. *Annual Review of Phytopathology* **43**, 83–116.
- Takada M, Yamada T, Shamsudin I and Okuda T (2015) Spatial variation in soil respiration in relation to a logging road in an upper tropical hill forest in peninsular Malaysia. *Tropics* **24**, 1–9.
- Tang ZH, Cao M, Sheng LX, Liang B and Zhang SY (2005) Fruit consumption and seed dispersal of wild banana *Musa acuminata* by short-nosed fruit bat *Cynocephalus sphinx*. *Acta Zoologica Sinica* **51**, 608–615.

- Tang ZH, Sheng LX, Ma XF, Cao M, Parsons S, Ma J and Zhang S** (2007) Temporal and spatial patterns of seed dispersal of *Musa acuminata* by *Cynopterus sphinx*. *Acta Chiropterologica* **9**, 229–236.
- Tielborger K, Petru M and Lampei C** (2012) Bet-hedging germination in annual plants: a sound empirical test of the theoretical foundations. *Oikos* **121**, 1860–1868.
- Vázquez-Yanes C and Orozco-Segovia A** (1982) Seed germination of a tropical rain forest pioneer tree (*Heliocarpus donnell-smithii*) in response to diurnal fluctuation of temperature. *Physiologia Plantarum* **56**, 295–298.
- Vázquez-Yanes C and Orozco-Segovia A** (1993) Patterns of seed longevity and germination in the tropical rainforest. *Annual Review of Ecology and Systematics* **24**, 69–87.
- Wattanachaiyingcharoen D and Turner D** (1990) Viability, germination and dormancy of banana seed (*Musa acuminata* ssp.). *Banana Newsletter* **13**, 16.
- Zhang GM, Tang JW, Shi JP and Bai KJ** (2000) Niche dynamics of dominant populations of *Musa acuminata* Colla pioneer community in Xishuangbanna, SW China. *Journal of Plant Resources and Environment* **9**, 22–26.
- Zizka A, Silvestro D, Andermann T, Azevedo J, Duarte Ritter C, Edler D, Farooq H, Herdean A, Ariza M, Scharn R, Svanteson S, Wengstrom N, Zizka V and Antonelli A** (2019) CoordinateCleaner: standardized cleaning of occurrence records from biological collection databases. *Methods Ecology Evolution* **10**, 744–751.