1	Increased daily temperature fluctuations overrule the ability of gradual thermal evolution to
2	offset the increased pesticide toxicity under global warming

- 3
- 4 Julie Verheyen, Vienna Delnat and Robby Stoks
- 5
- Evolutionary Stress Ecology and Ecotoxicology, University of Leuven, Charles Deberiotstraat 32,
 B-3000 Leuven, Belgium
- 8
- 9
- 10 Keywords: "Climate-induced toxicant sensitivity" (CITS) concept; Evolutionary ecotoxicology;
- 11 Geographic variation; Multiple stressors; "Toxicant-induced climate change sensitivity" (TICS)
- 12 concept; Upper thermal tolerance
- 13

14 Abstract:

The widespread evidence that global warming can increase species sensitivities to chemical 15 toxicants and vice versa, and the recent insight that thermal evolution may mitigate these effects is 16 crucial to predict the future impact of toxicants in a warming world. Nevertheless, a major 17 18 component of global warming, the predicted increase in daily temperature fluctuations (DTFs), has been ignored at the interface of evolutionary ecotoxicology and global change biology. We studied 19 whether 4°C warming and an 5°C DTF increase (to 10°C DTF) magnified the negative impact of 20 the insecticide chlorpyrifos (CPF) in larvae of low- and high-latitude populations of the damselfly 21 22 Ischnura elegans. While 4°C warming only increased CPF-induced mortality in high-latitude larvae, the high (10°C) DTF increased CPF-induced larval mortality at both latitudes. CPF reduced 23 the heat tolerance, however, this was buffered by latitude-specific thermal adaptation to both mean 24 25 temperature and DTF. Integrating our results in a space-for-time substitution indicated that gradual thermal evolution in high-latitude larvae may offset the negative effects of CPF on heat tolerance 26 under warming, unless the expected DTF increase is taken into account. Our results highlight the 27 crucial importance of jointly integrating DTFs and thermal evolution to improve risk assessment 28 of toxicants under global warming. 29

30 Introduction

Chemical pollution and global warming are two major threats to organisms, that moreover can interact^{1–5}. This constitutes a key challenge for a realistic ecological risk assessment under global warming^{2,6,7}. Two concepts are pivotal at this intersection of ecotoxicology and global change biology. On the one hand, higher mean temperatures can magnify the lethal effects of many pesticides^{1,2} such as organophosphates (including the insecticide chlorpyrifos^{8–10}). This is encapsulated in the "climate-induced toxicant sensitivity" (CITS) concept^{2,4,11}. On the other hand, many studies demonstrated that exposure to chemical toxicants can decrease the heat tolerance of
ectotherms¹²⁻¹⁸. The latter is captured by the "toxicant-induced climate change sensitivity" (TICS)
concept^{1,2,4}.

While there is increasing awareness that to better understand and predict the impact of 40 toxicants¹⁹ and of warming²⁰ an evolutionary perspective is pivotal, this has largely been ignored 41 in studies on the CITS and TICs concepts. Despite emerging insights that populations along 42 geographical gradients may differ both in toxicant sensitivity²¹ and in thermal sensitivity²⁰, studies 43 on the CITS and TICS concepts almost exclusively focused on single populations. This limits our 44 45 understanding of the role of thermal adaptation in shaping geographic variation in the strength of 46 both concepts. Moreover, this restricts our ability to predict the impact of gradual thermal 47 evolution in shaping the impact of toxicants under global warming. Indeed, in case of latitude-48 associated thermal adaptation one can use the current sensitivity to high temperatures and to chemical toxicants at high temperatures in the populations at warmer, lower latitudes to predict the 49 50 future sensitivity to warming and chemical toxicants under warming in high-latitude populations (for example^{22,23}). This so-called space-for-time substitution approach thereby assumes gradual 51 52 thermal evolution of the high-latitude populations toward the phenotype of the low-latitude populations 20,24,25 . While powerful to predict the effects of micro-evolution 26 , this approach has 53 54 never been used to address how gradual thermal evolution may shape the strength of the TICS and CITS concepts under future warming. 55

Besides increases in mean temperature, increases in daily temperature fluctuations (DTFs) are a largely overlooked key component of global warming²⁷. Recent ecological studies showed that increases in DTFs may impact fitness more severely compared to increases in mean temperature^{27,28}. Moreover, space-for-time substition studies only take increases in mean temperature into account and not increases in DTF. Recent evidence, however, showed that DTFs

may switch predictions of the impact of increases in mean temperature²⁹. DTFs have only recently 61 been integrated in ecotoxicology. Yet, the few studies all demonstrated DTF can increase chemical 62 toxicity (for pesticides $^{30-32}$; for metals 33), highlighting the importance of integrating DTFs in the 63 CITS concept. This all indicates DTFs should be considered in studies at the interface of 64 evolutionary ecotoxicology and global change biology. Furthermore, the few studies so far 65 66 considered the effect of DTF on toxicity at a single mean temperature, yet the effects of DTFs on fitness, hence likely also on sensitivity to chemical toxicants, may strongly differ in magnitude 67 depending on the mean temperature³⁴. 68

Although both the CITS and TICS concepts are necessary to understand and predict the effects of chemical toxicants under global warming, they have been consistently studied in isolation. Yet, it can be expected that both concepts are interconnected making studies of single concepts potentially misleading. A notable exception is the study by Op de Beeck et al.¹⁸ showing that exposure to an increase in mean temperature changed how the pesticide chlorpyrifos reduced the heat tolerance, thereby illustrating the interdependence of both concepts. Studies how DTFs, potentially in interaction with mean temperatures, may connect both concepts are missing.

76 We investigated the single and combined effects of increases in mean temperature and in DTF on the sensitivity to the pesticide chlorpyrifos (CPF), and vice versa the effect of exposure to 77 78 CPF on the sensitivity to warming, both in low- and high-latitude populations of an aquatic insect. This approach allowed to address in an integrated way the above identified knowledge gaps in the 79 TICS and CITS concepts: the influence of gradual evolution (thermal adaptation), the role of DTFs 80 and the interdependence of both concepts. Specifically, we tested: (i) if 4°C warming and a 5°C 81 increase in DTF increase CPF toxicity (CITS concept) by studying effects on two life-history traits 82 (mortality and growth rate); (ii) if exposure to CPF decreases heat tolerance (TICS concept); (iii) if 83 4°C warming and a 5°C DTF increase negatively impacted the CPF-induced decrease in heat 84

85	tolerance (interdependence of CITS and TICS); and (iv) if gradual thermal evolution can buffer for
86	the increased CPF toxicity under global warming (space-for-time substitution approach ²⁵). The
87	organophosphate insecticide chlorpyrifos (CPF) is frequently applied worldwide ³⁵ , and is a top ten
88	chemical found in UK surface waters with the highest risk to aquatic wildlife ³⁶ . Moreover, it is
89	considered a priority pollutant by the European Water Framework Directive 2000/60/EC ³⁷ . As
90	study organisms we chose Ischnura elegans damselfly larvae which inhabit shallow freshwater
91	ponds that are subjected to large DTFs. Damselfly larvae are especially vulnerable to both
92	warming ³⁸ and toxicants ³⁹ since they cannot escape exposure to these stressors due to their
93	obligated aquatic life ⁴⁰ . We studied high-latitude (Denmark and southern Sweden) and low-
94	latitude (southern France) populations to assess the influence of thermal adaptation on the CITS
95	and CITS concepts and to assess the future stength of both concepts under global warming in the
96	high-latitude populations by applying a space-for-time substitution ²⁵ .

97 Materials and methods

98 *Study populations*

Early summer 2017 (mid of June – mid of July), 30 mated females of the damselfly I. elegans 99 were sampled at each of three populations in both the high-latitude region (Denmark and southern 100 Sweden) and the low-latitude region (southern France) of the species' European range⁴¹. All six 101 populations were located at shallow ponds; these were likely not directly exposed to pesticides 102 because they were situated >100 m from agricultural fields⁴², however, we cannot exclude the 103 possibility that they were exposed to very low pesticide concentrations. Given pesticide exposure 104 is in general higher at low latitudes than at high latitudes in Europe⁴³, it is possible that 105 immigration of damselflies from sites directly exposed to pesticides into the unexposed study 106 populations may have been larger at the low latitude. Yet, this was not mirrored in our results as 107 108 for example at 20°C larval mortality in low-latitude larvae was as high as in high-latitude larvae

(and even higher at 10°C DTF) (see results). The low-latitude populations were located in southern
France: St. Martin de Crau (43°37'57.8"N; 04°46'55.1"E), Bassin de Réaltor (43°28'11.1"N;
05°19'44.1"E) and La Durance (43°43'52.5"N; 05°44'53.0"E). The high-latitude populations
were located in southern Sweden: Hovgardsdammarna (57°14'24.3"N, 12°08'28.2"E) and Kalmar
Dämme (56°40'04.6"N; 16°17'46.5"E); and in Denmark: Roskilde (55°39'09.8"N;
12°08'01.7"E).

Mated females were placed individually in plastic cups (7.5 cm height, 3.5 cm diameter) that contained wet filter papers for oviposition. Only filter papers that contained eggs were transferred to the lab in Leuven (Belgium). After arrival, the eggs were immediately stored in a temperature-controlled incubator with a constant water temperature of 22°C and a 14:10h lightdark cycle. Ten day old hatchlings were separated and kept individually in plastic cups (5 cm height, 6 cm diameter) containing 90 mL dechlorinated tap water. From this moment onwards, larvae were fed *Artemia* nauplii ad libitum, five days per week (Monday-Friday).

122 General experimental setup

123 From hatching onwards, larvae were placed at one of six combinations of two mean temperatures $(20^{\circ}\text{C and } 24^{\circ}\text{C})$ and three daily temperature fluctuation levels (DTFs: constant = 0°C , low = 5°C 124 125 and high = 10° C). One day after larvae had reached the final instar, they entered the 6-day 126 pesticide exposure period, resulting in 12 treatment combinations per latitude: 2 mean temperatures \times 3 DTFs \times 2 pesticide treatments (solvent control and chlorpryifos). Depending on 127 mortality, we started between 24 and 50 larvae per treatment combination (total of 852 larvae). 128 129 Larvae coming from the three populations per latitude were as equally divided as possible among the different treatment combinations. 130

131 The temperature treatments were chosen based on (i) the current mean water temperature
132 and (ii) the current maximum DTFs during summer in shallow (<1m) freshwater ponds that the

study species inhabits at high latitude (mean of 20°C and maximum DTF of 5°C) and low latitude 133 (mean of 24°C and maximum DTF of 10°C). These data were derived from the Flake model⁴⁴ and 134 confirmed by temperature data measured by Hobo onset data loggers (TidbiT v2 Temp logger) in 135 both high- and low-latitude shallow ponds inhabited by the study species^{45–47}. The same loggers 136 were used throughout the entire laboratory experiment to collect every 10 minutes water 137 temperature data in each temperature treatment. The collected water temperature data are shown 138 per temperature treatment in figure S1 (Appendix S1). 139 The current thermal conditions at the low latitude (mean summer water temperature of 140

141 24° C and maximum DTF of 10° C) correspond to the expected future thermal conditions at the

high latitude under global warming. This is according to the IPCC (2013)⁴⁸ Representative

143 Concentration Pathway (RCP) 8.5, which predicts by 2100 (i) a 4°C increase in mean temperature

and (ii) a 5°C increase in magnitude of $DTF^{27,48}$. The magnitude of DTF at high latitude was

estimated using daily maximum and minimum temperatures predicted by BCC_CSL 1.1 (Beijing

146 Climate Center Climate System Model 1.1) (following Paaijmans et al.⁴⁹). More details on this

147 method can be found in Verheyen and $Stoks^{29}$.

148 *Pesticide exposure*

149 When larvae moulted into the final instar they entered the 6-day pesticide exposure period. Larvae

150 were placed individually in glass jars (200 mL) that were filled with 100 mL medium (solvent

151 control or chlorpyrifos). All media were daily renewed (= static renewal). A chlorpyrifos (CPF)

152 concentration of $1 \mu g/L$ was selected based on a previous experiment where final instar larvae

were exposed to four concentrations (0, 0.75, 1.25 and 1.75 μ g/L CPF) under the same

154 experimental conditions. At 1.25 µg/L CPF larvae suffered considerable mortality (ca. 45%) while

at 0.75 µg/L CPF there was only a slight growth reduction. In edge-to-field ponds where *I. elegans*

may be very abundant⁵⁰, CPF peak concentrations up to $100 \mu g/L$ can occur through run-off⁵¹.

157	The exposure concentration was obtained from a CPF stock solution (1 mg/mL) which was
158	stored in a dark room at 4°C and renewed monthly. To prepare this CPF stock solution, CPF
159	powder (Sigma-Aldrich, purity > 99%) was dissolved in ethanol (absolute, 100%). The stock
160	solution was then further diluted to acquire the targeted CPF concentration of 1 μ g/L. A solvent
161	control was used with the same concentration (1 μ L/L) of ethanol (absolute, 100%) as in the
162	pesticide treatment. This low ethanol concentration is assumed to have no effects on the response
163	variables since it is lower (12×) than the NOEC for aquatic invertebrates ⁵² and much lower (66×)
164	than the recommended concentration of 100 μ L/L ⁵³ .
165	Water samples were taken at two moments to measure the CPF concentrations in the glass
166	jars. At the start of the exposure period, water samples were taken from six jars per mean
167	temperature. After 24 hours (just before the medium was renewed), six water samples were taken
168	at each of the six thermal treatment combinations. CPF concentrations were measured using UPLC
169	MS/MS with Triple Quadrupole Mass Spectrometry at KU Leuven. The mean start CPF
170	concentration in the glass jars at the mean temperature of 20°C was 0.98 μ g/L (SE: 0.04), while it
171	was 1.04 μ g/L (SE: 0.04) at the mean temperature of 24°C. At the mean temperature of 20°C,
172	initial CPF concentrations decreased after 24 hours to 0.59 μ g/L (SE: 0.03) at the constant (0°C)
173	DTF treatment, to 0.56 μ g/L (SE: 0.05) at the low (5°C) DTF treatment and to 0.62 μ g/L (SE:
174	0.05) at the high (10°C) DTF treatment. At the mean temperature of 24°C, initial CPF
175	concentrations decreased to 0.61 μ g/L (SE: 0.05) at the constant (0°C), to 0.53 μ g/L (SE: 0.06) at
176	the low (5°C) and to 0.63 μ g/L (SE: 0.03) at the high (10°C) DTF treatment.
177	Life-history traits

178 Mortality was checked daily during the 6-day pesticide exposure period. To obtain growth rates,

each larva was weighed twice: at the start (initial mass) and at the end (final mass) of the 6-day

pesticide exposure period. Wet masses were obtained to the nearest 0.01 mg using an electronic
balance (Mettler Toledo® AB135-S, Columbus, Ohio, USA). Larval growth rates were calculated
as [ln (final mass) – ln (initial mass)] / 6 days. Before the larvae were weighed, they were gently
blotted dry with tissue paper; this gives reliable wet mass data that strongly correlate with dry
mass⁵⁴. Sample sizes for larval growth rate varied between 23 and 37 larvae for each of the 12
treatment combinations (total of 730 larvae, see Fig. 2 for exact sample sizes).

186 *Heat tolerance: CTmax*

The heat tolerance of each larvae was assessed after the 6-day pesticide exposure period by 187 measuring the critical thermal maximum (CTmax). This is considered a good estimation to assess 188 an organism's vulnerability to warming⁵⁵. A dynamic method was used whereby larvae were 189 heated up at a constant rate until the temperature (CTmax) was reached where larvae neither 190 moved any body parts nor showed muscular spasms⁵⁶. For this, larvae were placed individually in 191 plastic vials (50 mL), which were kept floating in a temperature-controlled aquarium. Vials were 192 193 filled with dechlorinated tap water in the absence of the pesticide. The initial water temperature in 194 the vials was set at 20°C or 24°C depending on the mean water temperature of the larval treatment. All CTmax tests started between 10h45 and 13h45, in which all DTF treatments reached the mean 195 temperature of either 20°C or 24°C. 196

We used a constant heating rate of 0.3°C per minute that was realized using an aquarium
heater (TC120 optima immersion thermostat, Cambridgeshire, UK). The applied heating rate lies
in the commonly used range when quantifying the CTmax of aquatic insects (for example^{56,57}).
Furthermore, this heating rate is considered to be fast enough to avoid acclimation and slow
enough for the larvae's body temperatures to closely track the raising water temperatures⁵⁷.
When larvae reached their CTmax, they were immediately transferred to their original
temperature treatment to recover. Only very few larvae (1.62%: 12 out of 742 larvae) did not

recover within 20 minutes after the CTmax test. These were presumed to be dead, and were
excluded from the CTmax analyses. The recovered larvae were weighed after the CTmax test. In
all trials the saturation of dissolved oxygen in the water of the vials remained above 80%. Water
temperatures were measured in a randomly chosen vial during each trial to check the heating rate
in each trial. Sample sizes for heat tolerance (CTmax) varied between 23 and 37 larvae for each of
the 12 treatment combinations (total of 730 larvae, see Fig. 3 for exact sample sizes).

210 Statistical analyses

The single and combined effects of mean temperature, DTF, pesticide exposure and latitude on 211 212 larval growth rate and heat tolerance were analyzed using linear mixed models within the 'lme4' package v1.1-18.1⁵⁸, and by using the 'afex' package v0.22-1⁵⁹. Effects on mortality (binary 213 response variable) were tested using a generalized linear mixed model with a binomial error 214 distribution and the logit-link function. Since almost no larval mortality occurred in the control 215 treatments (see results), the single and combined effects of mean temperature, DTF and latitude 216 217 were analyzed separately for larvae exposed to the solvent control and to the pesticide. This allows 218 more powerful tests whether the pesticide effect is stronger at the higher mean temperature and at higher DTFs, and whether this differs between latitudes. Population nested in latitude was added in 219 each model as a random factor, although it was never significant. Furthermore, body mass was 220 221 added to the model as covariate in the analysis of heat tolerance. To meet ANOVA assumptions, 222 larval growth rates were square root transformed, while heat tolerances were boxcox transformed. R 3.4.0. for Windows was used to statistically analyze the data⁶⁰. Wald chi-square statistics 223 and p-values for fixed effects were calculated by using the 'car' package v3.0-2⁶¹. Interactions 224 between treatment combinations were further analyzed by comparing least-square means using 225 Tukey posthoc tests obtained with the 'lsmeans" package v2.27-62⁶². Given the focus on the 226 effects of the pesticide and DTF, we interpreted interactions from this perspective. 227

228 **Results**

229 *Mortality*

230 The main model indicated that exposure to CPF caused a general increase in mortality (main effect

- Pesticide: $\chi_2^2 = 23.50$, P < 0.001). Separate analyses per pesticide treatment showed that mortality
- was low (on average 3.12%) in all solvent control treatments (Fig. 1). Neither mean temperature,
- 233 DTF or latitude, nor their interactions affected mortality in the absence of the pesticide (all P >
- 234 0.69, Fig. 1). However, in pesticide-exposed larvae the 10°C DTF treatment caused in general an
- increase in CPF-induced mortality of ca. 11% compared to the 0°C and 5°C DTF treatments (DTF:
- 236 $\chi_2^2 = 6.44$, P = 0.040). Furthermore, mean temperature and latitude interacted in pesticide-exposed
- 237 larvae (MeanT × Latitude: χ_1^2 =5.78, P = 0.016, Fig. 1): CPF-induced mortality increased with ca.
- 19.4 % at 24°C compared to 20°C, but only in high-latitude larvae (Tukey: P = 0.0037).
- 239 *Growth rate*
- Exposure to the pesticide reduced the growth rate (main effect Pesticide), yet only in low-latitude
- larvae at 24°C (Tukey: P < 0.001) resulting in a significant Pest × MeanT × Lat interaction (Table
- 1, Fig. 2). In general, low-latitude larvae grew faster than high-latitude larvae (Latitude: Table 1,
- Fig. 2). The effect of DTF depended on the mean temperature and latitude (DTF \times MeanT and
- 244 DTF \times MeanT \times Lat: Table 1, Fig. 2). At a mean of 20°C, growth rates at both latitudes were not
- affected by DTF (Tukey: all P > 0.37). At a mean of 24°C, growth rate in high-latitude larvae
- decreased at 5°C (Tukey: P = 0.0010) and 10°C DTF (Tukey: P = 0.0057) compared to 0°C DTF;
- 247 while growth rate in low-latitude larvae did not decrease when temperatures fluctuated (Tukey: all
- 248 P > 0.27).
- 249 *Heat tolerance (CTmax)*

250 Overall, larvae had a higher heat tolerance when reared at 24°C than at 20°C (main effect MeanT:

Table 1, Fig. 3). Exposure to the pesticide, if anything, reduced the heat tolerance (main effect

252	Pesticide) and this tended to be stronger at high DTF (Pest \times DTF: $P = 0.078$). Notably, this Pest \times
253	DTF interaction effect was further shaped by mean temperature and latitude resulting in a
254	significant Pest \times DTF \times MeanT \times Lat interaction (Table 1, Fig. 3). When the high-latitude larvae
255	were kept at their local mean summer temperature of 20°C they only showed a CPF-induced
256	reduction in CTmax at the high 10°C DTF (Tukey: $P = 0.0078$) and not at the other DTFs (both P
257	> 0.45) (Pest × DTF: χ_2^2 = 7.10, <i>P</i> = 0.029). Yet, at 24°C high-latitude larvae consistently suffered
258	a CPF-induced reduction in CTmax (Pest: $\chi_1^2 = 11.33$, $P < 0.001$; Pest × DTF: $\chi_2^2 = 0.79$, $P =$
259	0.67). Similarly, when the low-latitude larvae were kept at their local mean summer temperature of
260	24°C they only showed a CPF-induced reduction in CTmax at the high 10°C DTF (Tukey: $P =$
261	0.014) and not at the other DTFs (both $P > 0.69$) (Pest × DTF: $\chi^2_2 = 5.19$, $P = 0.074$). Yet, at 20°C
262	low-latitude larvae tended to consistently suffer a CPF-induced reduction in CTmax (Pest: χ_1^2 =
263	2.74, $P = 0.098$; Pest × DTF: $\chi_2^2 = 1.53$, $P = 0.46$).

264 Discussion

As expected, exposure to the here applied nominal concentration of the pesticide chlorpyrifos (1 265 µg/L CPF) increased mortality, and decreased growth rate and heat tolerance in *I. elegans* 266 damselfly larvae. A CPF-induced reduction in heat tolerance is as predicted by the 'toxicant-267 induced climate change sensitivity' (TICS) concept². A new key finding was that latitude-specific 268 thermal adaptation buffered for the CPF-induced reduction in thermal tolerance in both high- and 269 270 low-latitude larvae; as a result, the combination of mean temperature and DTF shaped the TICS patterns. Consistent with the 'climate-induced toxicant sensitivity' (CITS) concept², warming 271 272 increased CPF toxicity in terms of an increased mortality (high-latitude larvae) and a decreased growth rate (low-latitude larvae), and the high 10°C DTF increased CPF-induced mortality. 273

274 *General patterns in heat tolerance and growth rate in the solvent control*

Heat tolerance of larvae from both latitudes was higher when acclimated to a higher mean
temperature, supporting the general pattern that upper thermal limits increase with higher
acclimation temperatures^{57,63,64}. The higher growth rate of low-latitude compared to high-latitude
larvae matches previous findings obtained at constant^{9,65} and at fluctuating rearing temperatures²⁹.
This reflects their faster life history associated with having multiple generations per year while the
high-latitude larvae complete one generation every two years⁶⁶.

Warming decreased growth rate only in high-latitude larvae when DTFs were present (5°C 281 282 and 10°C), while in low-latitude larvae the growth rate was not affected by mean temperature and 283 DTF. This indicates that at 24°C both DTFs exceeded the optimal temperature for growth rate in 284 high-latitude larvae but not in low-latitude larvae, matching latitude-associated thermal adaptation (see also Verheven and Stoks²⁹) as local mean summer water temperatures are 20°C in the high-285 latitude populations and 24°C in the low-latitude populations^{45–47}. This DTF-induced growth 286 reduction at 24°C is probably the result of prioritizing allocation of energy to increased metabolic 287 demands for cell maintenance over allocation of energy towards growth^{2,28,67}. 288

289 Warming and DTFs increased pesticide toxicity (CITS)

Not only warming but also DTFs increased CPF toxicity in terms of mortality, highlighting that 290 both the predicted increases in mean temperature and in temperature variation under global 291 warming^{34,48} are key drivers of the CITS concept². Our results confirm the general pattern that the 292 toxicity of many pesticides (including CPF) increases at higher mean temperatures^{1,2} (for the study 293 species⁹). Two major mechanisms can explain this increase in pesticide toxicity at higher constant 294 temperatures: an increased pesticide uptake¹¹ and an increased metabolic conversion into more 295 toxic metabolites¹⁰ (for CPF the conversion to CPF oxon⁶⁸). Notably, 4°C warming only increased 296 297 CPF-induced mortality in high-latitude larvae but not in low-latitude larvae. The latter indicates

that local thermal adaptation to higher mean temperatures in low-latitude larvae was able to bufferthe negative impact of CPF on mortality.

The increased CPF toxicity under the high (10°C) DTF matches the few ecotox studies 300 301 that integrated DTFs and all showed chemical toxicants becoming more toxic under DTFs in aquatic ectotherms (for pesticides^{30–32}, for metals³³). The underlying mechanisms for a higher 302 toxicity under DTFs are poorly understood. The two above discussed major mechanisms 303 underlying the higher CPF toxicity at higher mean temperatures may play a role because during 304 305 the 10°C DTF cycle larvae were exposed for several hours to high temperatures going up to 25°C 306 at the mean temperature of 20°C and even to 29°C at the mean temperature of 24°C. Moreover, since exposure to DTFs is assumed to be energetically costly for ectotherms (reviewed in Colinet 307 et al.²⁸), less energy will be available for detoxification and damage repair⁶⁹ when exposed to a 308 309 second stressor like the pesticide CPF. Pesticide toxicity can indeed increase greatly when 310 combined with an energetically costly environmental stressor⁵.

311 Chlorpyrifos also reduced the growth rate but only at 24°C in low-latitude larvae. CPF-312 induced reductions in growth rate have been observed before in damselfly larvae⁷⁰, including the 313 study species⁹. This can be explained by an energy trade-off between growth rate and defense mechanisms against the pesticide⁶⁹. Since CPF becomes more toxic at higher temperatures¹, at 314 315 24°C larvae probably allocated more energy away from growth rate towards detoxification (e.g. upregulation of glutathione-S-transferase⁷⁰) and other defense mechanisms (e.g. upregulation of 316 heat shock protein 70⁷⁰). That we did not observe a CPF-induced reductions in growth rate in high-317 318 latitude larvae may be explained by the fact that growth rates were already low in the control treatments (because of their slower life history⁶⁶), especially when temperatures got stressful (at 319 24°C with 5°C or 10°C DTF). Moreover, survival selection may have removed the weakest (with 320

the lowest growth rates) high-latitude larvae at 24°C, thereby masking any CPF impact on larvalgrowth rate.

323 *Pesticide exposure decreased the heat tolerance (TICS)*

324 In most thermal treatment combinations, exposure to the pesticide CPF reduced the heat tolerance of both low- and high-latitude larvae. This matches the TICS concept² and is consistent with the 325 general pattern in aquatic ectotherms (in fish^{12–14}; in damselflies^{15,16}, including the study 326 species^{17,18}). Heat tolerance is determined by the mismatch between oxygen demand and supply 327 occurring at high extreme temperatures⁷¹. Under toxicant exposure basal metabolism and thereby 328 oxygen consumption are typically higher due to increased energy demands since more energy is 329 needed for detoxification and damage repair mechanisms⁷². In line with this, CPF increased energy 330 costs and negatively affected aerobic performance in the Japanese quail⁷³. Furthermore, in aquatic 331 invertebrates toxicant exposure can impair respiratory functioning which may reduce oxygen 332 uptake (reviewed in Noyes and Lema²). This is likely the case under CPF exposure as it has been 333 shown to damage gills in crabs⁷⁴ and fish⁷⁵. Taken together, the reduced heat tolerance under 334 toxicant exposure can be explained by the mismatch between oxygen demand and supply 335 occurring already at lower temperatures in case animals are exposed to chlorpyrifos⁷². 336 A key finding was that the CPF-induced decrease in thermal tolerance was buffered by 337

local thermal adaptation in both high- and low-latitude larvae. This is the first demonstration that
thermal adaptation can shape the TICS patterns by reducing the negative impact of a toxicant on
heat tolerance. Specifically, it was the combination of mean temperature and DTF that determined
the TICS patterns, thereby adding a new dimension to the importance of DTF in ecotoxicology.
Indeed, at the mean summer water temperature at the high latitude (20°C), CPF consistently
reduced the heat tolerance in low-latitude larvae across all DTFs, while in high-latitude larvae this
reduction only occurred at high (10°C) DTF. Similarly, at the mean summer water temperature at

the low latitude (24°C), CPF reduced heat tolerance across all DTFs in high-latitude larvae, while
in low-latitude larvae this reduction was only present at high (10°C) DTF. Notably, these
combined effects of mean temperature and DTF in shaping the toxicant-induced sensitivity to
climate change (TICS), also indicate that the TICS and CITS concepts are not independent (see
also Op de Beeck et al.¹⁸).

350 Implications for risk assessment under global warming

351 Both the TICS and CITS concepts are central to our understanding of the effects of toxicants in a warming world ^{2,4}. By studying in an integrated way the single and combined effects of 4°C 352 353 warming and an 5°C increase in DTF on chlorpyrifos toxicity, and vice versa the effect of exposure to CPF on the sensitivity to warming, at two strategically chosen latitudes, we added 354 important novel insights to both concepts. We provided proof-of-principle that besides an increase 355 in mean temperature also higher DTFs under global warming are not only a key driver of CITS (by 356 increasing mortality and decreasing larval growth rate), but more importantly also of the 357 358 interdependence of both concepts. Notably, other conclusions would have been reached about both 359 the strengths of both concepts when ignoring DTF. This connection between the TICS and CITS concepts differed between latitudes, reflecting a crucial role of latitude-associated thermal 360 adaptation. This highlights an overlooked geographic component in how DTF may shape toxicant 361 362 sensitivity and the connectedness of the TICS and CITS concepts.

Our results imply that taking DTF into account may be crucial to arrive at realistic predictions of the effect of global warming on toxicant sensitivity. Indeed, while our results suggest that gradual thermal evolution in high-latitude larvae may buffer the effects of CPF on heat tolerance under warming, this is no longer true when also taking the increase in DTF into account. This prediction is based on a space-for-time substitution^{20,25} taking into account that the current thermal regime at the low latitude (mean of 24°C and maximum DTF of 10°C) matches the future thermal regime at the high latitude under global warming by 2100, as predicted by the IPCC (2013)⁴⁸ RCP8.5 scenario. When assuming no thermal evolution, high-latitude larvae would suffer a CPF-induced decrease in heat tolerance under 4°C warming (24°C) at all DTFs. However, under the more realistic scenario where high-latitude larvae could gradually evolve to have the same ability as low-latitude larvae to deal with a higher mean temperature (24°C), high-latitude larvae would no longer suffer a CPF-induced reduction in heat tolerance under 4°C warming unless we also take into account the expected increase to 10°C DTF.

376 To conclude, we demonstrated that the daily temperature fluctuations which most 377 organisms encounter in nature are a key environmental factor shaping contaminant toxicity, 378 making them highly relevant for current and future risk assessment of toxicants. The novel finding 379 of latitude-associated thermal adaptation in how DTFs shape toxicity, adds an important but 380 overlooked dimension to the calls to take spatial variation in local conditions into account during risk assessment of toxicants^{21,76}. Moreover, we provided the first evidence that DTFs, that are 381 expected to increase under global warming²⁷, are highly relevant to consider at the interface of 382 383 evolutionary ecotoxicology and global change biology. Indeed, our results indicate that predictions of the future impact of CPF under global warming may be critically dependent on the inclusion of 384 DTF. Taken together, our results highlight the importance of integrating both daily temperature 385 fluctuations and a geographic component to improve both current and future risk assessment of 386 chemical toxicants under global warming^{2,7}. 387

388 Figure 1



Figure 1. Mean (± 1 SE) mortality (%) of high-latitude and low-latitude *Ischnura elegans* damselfly larvae as a function of pesticide



392

389

Figure 2



Figure 2. Mean (\pm 1 SE) growth rate of high-latitude and low-latitude *Ischnura elegans* damselfly larvae as a function of pesticide

396 exposure, daily temperature fluctuation (DTF) and mean temperature. Numbers above bars represent sample sizes.

Figure 3

400



401 Figure 3. Mean (± 1 SE) heat tolerance (measured as CTmax) of high-latitude and low-latitude *Ischnura elegans* damselfly larvae as a

402 function of pesticide exposure, daily temperature fluctuation (DTF) and mean temperature. Numbers above bars represent sample sizes.

Table 1. Results of linear mixed models testing for the effects of pesticide exposure (Pest), daily temperature fluctuation (DTF), mean temperature (MeanT) and latitude (Lat) on larval growth rate and heat tolerance (CTmax) in *Ischnura elegans* damselfly larvae. Body mass was added as covariate in the heat tolerance model. Bold *P*-values are significant (< 0.05).

	Growth rate			Heat tolerance		
Effect	χ^2	df	Р	χ^2	df	Р
Pest	10.75	1	0.0010	11.79	1	<0.001
DTF	8.43	2	0.015	5.26	2	0.072
Mean T	20.71	1	<0.001	61.80	1	<0.001
Lat	258.48	1	<0.001	43.01	1	<0.001
$DTF \times Mean T$	6.82	2	0.033	4.91	2	0.086
$\text{Pest} \times \text{Mean } T$	3.45	1	0.063	0.41	1	0.52
$\text{Pest} \times \text{DTF}$	2.39	2	0.30	5.10	2	0.078
$\text{Pest} \times \text{Lat}$	0.26	1	0.61	0.37	1	0.54
Mean $T \times Lat$	4.34	1	0.037	3.33	1	0.068
DTF × Lat	1.51	2	0.47	0.72	2	0.70
$Pest \times DTF \times Mean \ T$	2.81	2	0.24	0.51	2	0.77
$Pest \times Mean \ T \times Lat$	5.08	1	0.024	2.95	1	0.085
$Pest \times DTF \times Lat$	1.34	2	0.51	0.54	2	0.76
$DTF \times Mean \ T \times Lat$	6.75	2	0.034	4.46	2	0.11
$Pest \times DTF \times Mean \ T \times Lat$	2.39	2	0.30	7.19	2	0.028
Body mass	/	/	/	7.48	1	0.0063

404	Supporting Information
405	Appendix S1: Realized thermal regimes
406	Author Information
407	Corresponding author: Julie Verheyen; Tel.: +3216376436; fax: +3216324575
408	E-mail address: julie.verheyen@kuleuven.be
409	Authors' Contributions
410	JV and RS conceived and designed the experiments. JV performed the experiments and analyzed
411	the data. All authors wrote the manuscript and gave final approval for publication.
412	Funding Sources
413	Financial support came from FWO grants G.0524.17 and G.0956.19, and KU Leuven grant
414	C16/17/002.
415	Acknowledgements
415 416	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark.
415 416 417	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV
415 416 417 418	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV and VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authors
415 416 417 418 419	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV and VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authors declare no conflict of interest. Comments from three anonymous reviewers improved the
415 416 417 418 419 420	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV and VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authors declare no conflict of interest. Comments from three anonymous reviewers improved the manuscript.
415 416 417 418 419 420 421	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV and VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authors declare no conflict of interest. Comments from three anonymous reviewers improved the manuscript. Abbreviations
 415 416 417 418 419 420 421 422 	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV and VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authors declare no conflict of interest. Comments from three anonymous reviewers improved the manuscript. Abbreviations BCC_CSL 1.1, Beijing Climate Center Climate System Model 1.1; CITS, Climate-Induced
 415 416 417 418 419 420 421 422 423 	Acknowledgements We are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark. We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JV and VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authors declare no conflict of interest. Comments from three anonymous reviewers improved the manuscript. Abbreviations BCC_CSL 1.1, Beijing Climate Center Climate System Model 1.1; CITS, Climate-Induced Toxicant Sensitivity; CPF, chlorpyrifos; CTmax, critical thermal maximum; DTF, daily
415 416 417 418 419 420 421 422 423 424	AcknowledgementsWe are very grateful to Dinh Van Khuong for the collection of the damselfly eggs in Denmark.We thank Geert Neyens and Rony Van Aerschot for technical support during the experiment. JVand VD are PhD fellows of the Fund for Scientific Research Flanders (FWO). All authorsdeclare no conflict of interest. Comments from three anonymous reviewers improved themanuscript.AbbreviationsBCC_CSL 1.1, Beijing Climate Center Climate System Model 1.1; CITS, Climate-InducedToxicant Sensitivity; CPF, chlorpyrifos; CTmax, critical thermal maximum; DTF, dailytemperature fluctuation; IPCC, Intergovernmental Panel on Climate Change; Lat, latitude;

426 Representative Concentration Pathway 8.5; TICS, Toxicant-Induced Climate Change sensitivity.

427 **References**

- 428 (1) Noyes, P. D.; McElwee, M. K.; Miller, H. D.; Clark, B. W.; Van Tiem, L. A.; Walcott, K.
- 429 C.; Erwin, K. N.; Levin, E. D. The Toxicology of Climate Change: Environmental
- 430 Contaminants in a Warming World. *Environ. Int.* **2009**, *35*, 971–986.
- 431 https://doi.org/10.1016/j.envint.2009.02.006.
- 432 (2) Noyes, P. D.; Lema, S. C. Forecasting the Impacts of Chemical Pollution and Climate
- 433 Change Interactions on the Health of Wildlife. *Curr. Zool.* **2015**, *61*, 669–689.
- 434 https://doi.org/10.1093/czoolo/61.4.669.
- 435 (3) Holmstrup, M.; Bindesbøl, A. M.; Oostingh, G. J.; Duschl, A.; Scheil, V.; Köhler, H. R.;
- 436 Loureiro, S.; Soares, A. M. V. M.; Ferreira, A. L. G.; Kienle, C.; Gerhardt, A.; Laskowski,
- 437 R.; Kramarz, P. E.; Bayley, M.; Svendsen, C.; Spurgeon, D. J. Interactions between
- 438 Effects of Environmental Chemicals and Natural Stressors: A Review. *Sci. Total Environ.*

2010, *408*, 3746–3762. https://doi.org/10.1016/j.scitotenv.2009.10.067.

- 440 (4) Moe, S. J.; De Schamphelaere, K.; Clements, W. H.; Sorensen, M. T.; Van den Brink, P.
- 441 J.; Liess, M. Combined and Interactive Effects of Global Climate Change and Toxicants
- 442 on Populations and Communities. *Environ. Toxicol. Chem.* **2013**, *32*, 49–61.
- 443 https://doi.org/10.1002/etc.2045.
- Liess, M.; Foit, K.; Knillmann, S.; Schäfer, R. B.; Liess, H. D. Predicting the Synergy of
 Multiple Stress Effects. *Sci. Rep.* 2016, *6*, 1–8. https://doi.org/10.1038/srep32965.
- 446 (6) Landis, W. G.; Rohr, J. R.; Moe, S. J.; Balbus, J. M.; Clements, W.; Fritz, A.; Helm, R.;
- 447 Hickey, C.; Hooper, M.; Stahl, R. G.; Stauber, J. Global Climate Change and
- 448 Contaminants, a Call to Arms Not yet Heard? Integr. Environ. Assess. Manag. 2014, 10,

449

483-484. https://doi.org/10.1002/ieam.1568.

450	(7)	Van den Brink, P. J.; Boxall, A. B. A.; Maltby, L.; Brooks, B. W.; Rudd, M. A.;
451		Backhaus, T.; Spurgeon, D.; Verougstraete, V.; Ajao, C.; Ankley, G. T.; Apitz, S. E.;
452		Arnold, K.; Brodin, T.; Cañedo-Argüelles, M.; Chapman, J.; Corrales, J.; Coutellec, M.
453		A.; Fernandes, T. F.; Fick, J.; Ford, A. T.; Giménez Papiol, G.; Groh, K. J.; Hutchinson, T.
454		H.; Kruger, H.; Kukkonen, J. V. K.; Loutseti, S.; Marshall, S.; Muir, D.; Ortiz-
455		Santaliestra, M. E.; Paul, K. B.; Rico, A.; Rodea-Palomares, I.; Römbke, J.; Rydberg, T.;
456		Segner, H.; Smit, M.; van Gestel, C. A. M.; Vighi, M.; Werner, I.; Zimmer, E. I.; van
457		Wensem, J. Toward Sustainable Environmental Quality: Priority Research Questions for
458		Europe. Environ. Toxicol. Chem. 2018, 37, 2281–2295. https://doi.org/10.1002/etc.4205.
459	(8)	De Silva, P. M. C. S.; Pathiratne, A.; van Gestel, C. A. M. Influence of Temperature and
460		Soil Type on the Toxicity of Three Pesticides to Eisenia Andrei. Chemosphere 2009, 76,
461		1410-1415. https://doi.org/10.1016/j.chemosphere.2009.06.006.
462	(9)	Dinh Van, K.; Janssens, L.; Debecker, S.; Stoks, R. Temperature- and Latitude-Specific
463		Individual Growth Rates Shape the Vulnerability of Damselfly Larvae to a Widespread
464		Pesticide. J. Appl. Ecol. 2014, 51, 919–928. https://doi.org/10.1111/1365-2664.12269.
465	(10)	Harwood, A. D.; You, J.; Lydy, M. J. Temperature as a Toxicity Identification Evaluation
466		Tool for Pyrethroid Insecticides: Toxicokinetic Confirmation. Environ. Toxicol. Chem.
467		2009 , 28, 1051–1058. https://doi.org/10.1897/08-291.1.
468	(11)	Hooper, M. J.; Ankley, G. T.; Cristol, D. A.; Maryoung, L. A.; Noyes, P. D.; Pinkerton, K.
469		E. Interactions between Chemical and Climate Stressors: A Role for Mechanistic
470		Toxicology in Assessing Climate Change Risks. Environ. Toxicol. Chem. 2013, 32, 32–

- 471 48. https://doi.org/10.1002/etc.2043.
- 472 (12) Heath, S.; Heath, S.; Bennett, W. a; Bennett, W. a; Kennedy, J.; Kennedy, J.; Beitinger, T.
- 473 L.; Beitinger, T. L. Heat and Cold Tolerance of the Fathead Minnow, *Pimephales*
- 474 *Promelas*, Exposed to the Synthetic Pyrethroid Cyfluthrin. *Can. J. Fish. Aquat. Sci.* **1994**,
- 475 *51*, 437–440. https://doi.org/10.1139/f94-045.
- 476 (13) Messaad, I. A.; Peters, E. J.; Young, L. Thermal Tolerance of Red Shiner (Cyprinella
- 477 *Lutrensis*) after Exposure to Atrazine, Terbufos, and Their Mixtures. *Bull. Environ.*

478 *Contam. Toxicol.* **2000**, *64*, 748–754. https://doi.org/10.1007/s001280000067.

- 479 (14) Patra, W. P.; Chapman, J. C.; Lim, R. P.; Gehrke, P. C. The Effects of Three Organic
- 480 Chemicals on the Upper Thermal Tolerances of Four Freshwater Fishes. *Environ. Toxicol.*481 *Chem.* 2007, *26*, 1454–1459. https://doi.org/10.1897/06-156R1.1.
- 482 (15) Janssens, L.; Verberk, W.; Stoks, R. A Widespread Morphological Antipredator
- 483 Mechanism Reduces the Sensitivity to Pesticides and Increases the Susceptibility to
- 484 Warming. *Sci. Total Environ.* **2018**, *626*, 1230–1235.
- 485 https://doi.org/10.1016/j.scitotenv.2018.01.179.
- 486 (16) Janssens, L.; Stoks, R. Rapid Larval Development under Time Stress Reduces Adult
 487 Lifespan through Increasing Oxidative Damage. *Funct. Ecol.* 2018, *32*, 1036–1045.
- 488 https://doi.org/10.1111/1365-2435.13068.
- (17) Op de Beeck, L.; Verheyen, J.; Stoks, R. Competition Magnifies the Impact of a Pesticide
- in a Warming World by Reducing Heat Tolerance and Increasing Autotomy. *Environ*.
- 491 *Pollut.* **2018**, *233*, 226–234. https://doi.org/10.1016/j.envpol.2017.10.071.

492	(18)	Op de Beeck, L.; Verheyen, J.; Stoks, R. Integrating Both Interaction Pathways between
493		Warming and Pesticide Exposure on Upper Thermal Tolerance in High- and Low-Latitude
494		Populations of an Aquatic Insect. Environ. Pollut. 2017, 224, 714–721.
495		https://doi.org/10.1016/j.envpol.2016.11.014.
496	(19)	Brady, S. P.; Monosson, E.; Matson, C. W.; Bickham, J. W. Evolutionary Toxicology:
497		Toward a Unified Understanding of Life's Response to Toxic Chemicals. Evol. Appl.
498		2017 , 10, 745–751. https://doi.org/10.1111/eva.12519.
499	(20)	Stoks, R.; Geerts, A. N.; De Meester, L. Evolutionary and Plastic Responses of Freshwater

- Invertebrates to Climate Change: Realized Patterns and Future Potential. *Evol. Appl.* 2014,
 7, 42–55. https://doi.org/10.1111/eva.12108.
- 502 (21) Beketov, M. A.; Liess, M. Ecotoxicology and Macroecology Time for Integration.
 503 *Environ. Pollut.* 2012, *162*, 247–254. https://doi.org/10.1016/j.envpol.2011.11.011.
- 504 (22) Dinh Van, K.; Janssens, L.; Debecker, S.; De Jonge, M.; Lambret, P.; Nilsson-Örtman, V.;
- 505 Bervoets, L.; Stoks, R. Susceptibility to a Metal under Global Warming Is Shaped by
- 506 Thermal Adaptation along a Latitudinal Gradient. *Glob. Chang. Biol.* 2013, 19, 2625–
- 507 2633. https://doi.org/10.1111/gcb.12243.
- 508 (23) Janssens, L.; Dinh Van, K.; Debecker, S.; Bervoets, L.; Stoks, R. Local Adaptation and
- 509 the Potential Effects of a Contaminant on Predator Avoidance and Antipredator Responses
- 510 under Global Warming: A Space-for-Time Substitution Approach. *Evol. Appl.* **2014**, *7*,
- 511 421–430. https://doi.org/10.1111/eva.12141.
- 512 (24) Fukami, T.; Wardle, D. A. Long-Term Ecological Dynamics: Reciprocal Insights from
 513 Natural and Anthropogenic Gradients. *Proc. R. Soc. B Biol. Sci.* 2005, 272, 2105–2115.

514 https://doi.org/10.1098/rspb.2005.3277.

- 515 (25) De Frenne, P.; Graae, B. J.; Rodríguez-Sánchez, F.; Kolb, A.; Chabrerie, O.; Decocq, G.;
- 516 De Kort, H.; De Schrijver, A.; Diekmann, M.; Eriksson, O.; Gruwez, R.; Hermy, M.;
- 517 Lenoir, J.; Plue, J.; Coomes, D. A.; Verheyen, K. Latitudinal Gradients as Natural
- Laboratories to Infer Species' Responses to Temperature. J. Ecol. 2013, 101, 784–795.
- 519 https://doi.org/10.1111/1365-2745.12074.
- 520 (26) Wogan, G. O. U.; Wang, I. J. The Value of Space-for-Time Substitution for Studying
- 521 Fine-Scale Microevolutionary Processes. *Ecography.* **2017**, *40*, 1–12.
- 522 https://doi.org/10.1111/ecog.03235.
- 523 (27) Vasseur, D. A.; Delong, J. P.; Gilbert, B.; Greig, H. S.; Harley, C. D. G.; McCann, K. S.;
- 524 Savage, V.; Tunney, T. D.; O'Connor, M. I. Increased Temperature Variation Poses a
- 525 Greater Risk to Species than Climate Warming. *Proc. R. Soc. B* **2014**, *281*, 20132612.
- 526 https://doi.org/10.1098/rspb.2013.2612.
- 527 (28) Colinet, H.; Sinclair, B. J.; Vernon, P.; Renault, D. Insects in Fluctuating Thermal
- 528 Environments. *Annu. Rev. Entomol.* 2015, *60*, 123–140. https://doi.org/10.1146/annurev529 ento-010814-021017.
- 530 (29) Verheyen, J.; Stoks, R. Temperature Variation Makes an Ectotherm More Sensitive to
- Global Warming Unless Thermal Evolution Occurs. *J. Anim. Ecol.* published online.
 https://doi.org/10.1111/1365-2656.12946.
- 533 (30) Barbosa, M.; Inocentes, N.; Soares, A. M. V. M.; Oliveira, M. Synergy Effects of
- 534 Fluoxetine and Variability in Temperature Lead to Proportionally Greater Fitness Costs in
- 535 *Daphnia*: A Multigenerational Test. *Aquat. Toxicol.* **2017**, *193*, 268–275.

536 https://doi.org/10.1016/j.aquatox.2017.10.017.

- 537 (31) Willming, M. M.; Qin, G.; Maul, J. D. Effects of Environmentally Realistic Daily
- 538 Temperature Variation on Pesticide Toxicity to Aquatic Invertebrates. *Environ. Toxicol.*
- 539 *Chem.* **2013**, *32*, 2738–2745. https://doi.org/10.1002/etc.2354.
- 540 (32) Willming, M. M.; Maul, J. D. Direct and Indirect Toxicity of the Fungicide Pyraclostrobin
- 541 to *Hyalella Azteca* and Effects on Leaf Processing under Realistic Daily Temperature
- 542 Regimes. *Environ. Pollut.* **2016**, *211*, 435–442.
- 543 https://doi.org/10.1016/j.envpol.2015.11.029.
- 544 (33) Hallman, T. A.; Brooks, M. L. The Deal with Diel: Temperature Fluctuations,
- Asymmetrical Warming, and Ubiquitous Metals Contaminants. *Environ. Pollut.* 2015,
 206, 88–94. https://doi.org/10.1016/j.envpol.2015.06.005.
- 547 (34) Vázquez, D. P.; Gianoli, E.; Morris, W. F.; Bozinovic, F. Ecological and Evolutionary
- 548 Impacts of Changing Climatic Variability. *Biol. Rev.* **2017**, *92*, 22–42.
- 549 https://doi.org/https://doi.org/10.1111/brv.12216.
- 550 (35) Eaton, D. L.; Daroff, R. B.; Autrup, H.; Bridges, J.; Buffler, P.; Costa, L. G.; Coyle, J.;
- 551 McKhann, G.; Mobley, W. C.; Nadel, L.; Neubert, D.; Schulte-Hermann, R.; Spencer, P.
- 552 S. Review of the Toxicology of Chlorpyrifos with an Emphasis on Human Exposure and
- 553 Neurodevelopment. *Crit. Rev. Toxicol.* **2008**, *38*, 1–125.
- 554 https://doi.org/10.1080/10408440802272158.
- 555 (36) Johnson, A. C.; Donnachie, R. L.; Sumpter, J. P.; Jürgens, M. D.; Moeckel, C.; Pereira, M.
- 556 G. An Alternative Approach to Risk Rank Chemicals on the Threat They Pose to the
- 557 Aquatic Environment. *Sci. Total Environ.* **2017**, *599–600*, 1372–1381.

558 https://doi.org/10.1016/j.scitotenv.2017.05.039.

- 559 (37) Ojec, D. 2000/60/EC of the European Parliament and of the Council of 23 October 2000
- 560 Establishing a Framework for Community Action in the Field of Water Policy. *Off. J. Eur.*

561 *Communities* **2000**, 22.

- 562 (38) Hassall, C.; Thompson, D. J. The Effects of Environmental Warming on Odonata: A
 563 Review. *Int. J. Odonatol.* 2008, *11*, 131–153.
- 564 https://doi.org/10.1080/13887890.2008.9748319.
- 565 (39) Liess, M.; Von Der Ohe, P. C. Analyzing Effects of Pesticides on Invertebrate

566 Communities in Streams. *Environ. Toxicol. Chem.* **2005**, *24*, 954–965.

- 567 https://doi.org/10.1897/03-652.1.
- 568 (40) Stoks, R.; Debecker, S.; Dinh Van, K.; Janssens, L. Integrating Ecology and Evolution in
 569 Aquatic Toxicology: Insights from Damselflies. *Freshw. Sci.* 2015, *34*, 1032–1039.
 570 https://doi.org/10.1086/682571.
- 571 (41) Gosden, T. P.; Stoks, R.; Svensson, E. I. Range Limits, Large-Scale Bioeographic
- 572 Variation and Localised Evolutionary Dynamics in a Polymorphic Damselfly. *Biol. J.*

573 *Linn. Soc.* **2011**, *102*, 775–785. https://doi.org/10.1111/j.1095-8312.2011.01619.x.

- 574 (42) Hua, J.; Jones, D. K.; Mattes, B. M.; Cothran, R. D.; Relyea, R. A.; Hoverman, J. T. The
- 575 Contribution of Phenotypic Plasticity to the Evolution of Insecticide Tolerance in
- 576 Amphibian Populations. *Evol. Appl.* **2015**, *8*, 586–596. https://doi.org/10.1111/eva.12267.
- 577 (43) Kattwinkel, M.; Jan-Valentin, K.; Foit, K.; Liess, M. Climate Change, Agricultural
- 578 Insecticide Exposure, and Risk for Freshwater Communities. *Ecol. Appl.* 2011, 21, 2068–

579

584

2081. https://doi.org/10.1890/10-1993.1.

- 580 (44) Simmons, A.; Uppala, S. M.; Dee, D.; Kobayashi, S. ERA-Interim: New ECMWF
- 581 Reanalysis Products from 1989 Onwards. *ECMWF Newsl.* 2007, *110*, 25–35.
- 582 https://doi.org/10.21957/pocnex23c6.
- 583 (45) De Block, M.; Pauwels, K.; Van Den Broeck, M.; De Meester, L.; Stoks, R. Local Genetic

Adaptation Generates Latitude-Specific Effects of Warming on Predator-Prey

- 585 Interactions. *Glob. Chang. Biol.* **2013**, *19*, 689–696. https://doi.org/10.1111/gcb.12089.
- 586 (46) Dinh Van, K.; Janssens, L.; Stoks, R. Exposure to a Heat Wave under Food Limitation
- 587 Makes an Agricultural Insecticide Lethal: A Mechanistic Laboratory Experiment. *Glob*.

588 *Chang. Biol.* **2016**, *22*, 3361–3372. https://doi.org/10.1111/gcb.13415.

- (47) Debecker, S.; Stoks, R. Pace of Life Syndrome under Warming and Pollution: Integrating
 Life History, Behavior and Physiology across Latitudes. *Ecol. Monogr.* 2019, 89, e01332.
- 591 https://doi.org/10.1002/ecm.1332.
- 592 (48) IPCC. Climate Change 2013: The Physical Science Basis. Contribution of Working Group
- 593 *I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*;
- 594 Cambridge University Press, Cambridge, UK, 2013.
- 595 (49) Paaijmans, K. P.; Heinig, R. L.; Seliga, R. A.; Blanford, J. I.; Blanford, S.; Murdock, C.
- C.; Thomas, M. B. Temperature Variation Makes Ectotherms More Sensitive to Climate
 Change. *Glob. Chang. Biol.* 2013, *19*, 2373–2380. https://doi.org/10.1111/gcb.12240.
- 598 (50) Dijkstra, K.-D. B. *Field Guide to the Dragonflies of Britain and Europe*; British Wildlife
- 599 Publishing: Gillingham, Dorset, UK, 2006.

- 600 (51) Bernabò, I.; Sperone, E.; Tripepi, S.; Brunelli, E. Toxicity of Chlorpyrifos to Larval Rana
- 601 *Dalmatina*: Acute and Chronic Effects on Survival, Development, Growth and Gill
- 602 Apparatus. Arch. Environ. Contam. Toxicol. **2011**, *61*, 704–718.
- 603 https://doi.org/10.1007/s00244-011-9655-1.
- (52) United Nations Environment Program. Screening Information DataSet: Initial Assessment
 Report Ethanol Cas No: 64-17-5. 2004.
- 606 (53) OECD. Guidance Document on Aquatic Toxicity Testing of Difficult Substances and
- 607 *Mixtures*; Organisation for Economic Cooperation and Development: Paris, 2000.
- 608 (54) Stoks, R.; De Block, M.; Van De Meutter, F.; Johansson, F. Predation Cost of Rapid
- 609 Growth: Behavioural Coupling and Physiological Decoupling. J. Anim. Ecol. 2005, 74,

610 708–715. https://doi.org/10.1111/j.1365-2656.2005.00969.x.

- 611 (55) Huey, R. B.; Kearney, M. R.; Krockenberger, A.; Holtum, J. A. M.; Jess, M.; Williams, S.
- E. Predicting Organismal Vulnerability to Climate Warming: Roles of Behaviour,
- 613 Physiology and Adaptation. *Philos. Trans. R. Soc. B* **2012**, *367*, 1665–1679.
- 614 https://doi.org/10.1098/rstb.2012.0005.
- 615 (56) Verberk, W. C. E. P.; Bilton, D. T. Can Oxygen Set Thermal Limits in an Insect and Drive
- 616 Gigantism? *PLoS One* **2011**, *6*, e22610. https://doi.org/10.1371/journal.pone.0022610.
- 617 (57) Dallas, H. F.; Rivers-Moore, N. A. Critical Thermal Maxima of Aquatic
- 618 Macroinvertebrates: Towards Identifying Bioindicators of Thermal Alteration.
- 619 *Hydrobiologia* **2012**, 679, 61–76. https://doi.org/10.1007/s10750-011-0856-4.
- 620 (58) Bates, D.; Mächler, M.; Bolker, B.; Walker, S. C. Fitting Linear Mixed-Effects Models

621

- Using Lme4. J. Stat. Softw. 2015, 67, 1–48.
- 622 (59) Singmann, H.; Bolker, B.; Westfall, J.; Aust, F. Afex: Analysis of Factorial Experiments.
 623 2017.
- 624 (60) R Core Team. R: A Language and Environment for Statistical Computing, 2014.
 625 https://doi.org/ISBN 3-900051-07-0.
- 626 (61) Fox, J.; Weisberg, S. *An {R} Companion to Aplied Regression*, Second.; Sage
 627 Publications: Thousand Oaks CA, 2011.
- 628 (62) Lenth, R. V. Least-Squares Means: The R Package Lsmeans. J. Stat. Softw. 2016, 69, 1–
 629 33. https://doi.org/10.18637/jss.v069.i01.
- 630 (63) Geerts, A. N.; Vanoverbeke, J.; Vanschoenwinkel, B.; Van Doorslaer, W.; Feuchtmayr,
- H.; Atkinson, D.; Moss, B.; Davidson, T. A.; Sayer, C. D.; De Meester, L. Rapid
- Evolution of Thermal Tolerance in the Water Flea *Daphnia*. *Nat. Clim. Chang.* **2015**, *5*,

633 665–668. https://doi.org/10.1038/nclimate2628.

- 634 (64) Gunderson, A. R.; Stillman, J. H. Plasticity in Thermal Tolerance Has Limited Potential to
 635 Buffer Ectotherms from Global Warming. *Proc. R. Soc. B Biol. Sci.* 2015, 282, 20150401.
 636 https://doi.org/10.1098/rspb.2015.0401.
- 637 (65) Shama, L. N. S.; Campero-Paz, M.; Wegner, K. M.; De Block, M.; Stoks, R. Latitudinal
- and Voltinism Compensation Shape Thermal Reaction Norms for Growth Rate. *Mol. Ecol.*
- **2011**, *20*, 2929–2941. https://doi.org/10.1111/j.1365-294X.2011.05156.x.
- 640 (66) Corbet, P. S.; Suhling, F.; Soendgerath, D. Voltinism of Odonata: A Review. Int. J.
- 641 *Odonatol.* **2006**, *9*, 1–44. https://doi.org/10.1080/13887890.2006.9748261.

642	(67)	Ruel, J. J.; Ayres, M. P. Jensen's Inequality Predicts Effects of Environmental Variation.
643		Trends Ecol. Evol. 1999, 14, 361–366. https://doi.org/10.1016/S0169-5347(99)01664-X.
644	(68)	Buchwalter, D. B.; Sandahl, J. F.; Jenkins, J. J.; Curtis, L. R. Roles of Uptake,
645		Biotransformation, and Target Site Sensitivity in Determining the Differential Toxicity of
646		Chlorpyrifos to Second to Fourth Instar Chironomous Riparius (Meigen). Aquat. Toxicol.
647		2004, 66, 149–157. https://doi.org/10.1016/j.aquatox.2003.08.004.
648	(69)	Congdon, J. D.; Dunham, A. E.; Hopkins, W. A.; Rowe, C. L.; Hinton, T. G. Resource
649		Allocation-Based Life Histories: A Conceptual Basis for Studies of Ecological
650		Toxicology. Environ. Toxicol. 2001, 20, 1698–1703.
651		https://doi.org/10.1002/etc.5620200811.
652	(70)	Janssens, L.; Stoks, R. Exposure to a Widespread Non-Pathogenic Bacterium Magnifies
653		Sublethal Pesticide Effects in the Damselfly Enallagma Cyathigerum: From the
654		Suborganismal Level to Fitness-Related Traits. Environ. Pollut. 2013, 177, 143–149.
655		https://doi.org/10.1016/j.envpol.2013.02.016.
656	(71)	Verberk, W. C. E. P.; Overgaard, J.; Ern, R.; Bayley, M.; Wang, T.; Boardman, L.;
657		Terblanche, J. S. Does Oxygen Limit Thermal Tolerance in Arthropods? A Critical
658		Review of Current Evidence. Comp. Biochem. PhysiolPart A 2016, 192, 64-78.
659		https://doi.org/10.1016/j.cbpa.2015.10.020.
660	(72)	Sokolova, I. M. Energy-Limited Tolerance to Stress as a Conceptual Framework to
661		Integrate the Effects of Multiple Stressors. Integr. Comp. Biol. 2013, 53, 597-608.
662		https://doi.org/10.1093/icb/ict028.

663 (73) Narváez, C.; Ríos, J. M.; Píriz, G.; Sanchez-Hernandez, J. C.; Sabat, P. Subchronic

664		Exposure to Chlorpyrifos Affects Energy Expenditure and Detoxification Capacity in
665		Juvenile Japanese Quails. Chemosphere 2016, 144, 775–784.
666		https://doi.org/10.1016/j.chemosphere.2015.09.060.
667	(74)	Negro, C. L.; Collins, P. Histopathological Effects of Chlorpyrifos on the Gills,
668		Hepatopancreas and Gonads of the Freshwater Crab Zilchiopsis Collastinensis. Persistent
669		Effects after Exposure. Ecotoxicol. Environ. Saf. 2017, 140, 116-122.
670		https://doi.org/10.1016/j.ecoenv.2017.02.030.
671	(75)	Marigoudar, S. R.; Mohan, D.; Nagarjuna, A.; Karthikeyan, P. Biomarker and
672		Histopathological Responses of Lates Calcarifer on Exposure to Sub Lethal
673		Concentrations of Chlorpyrifos. Ecotoxicol. Environ. Saf. 2018, 148, 327-335.
674		https://doi.org/10.1016/j.ecoenv.2017.10.026.
675	(76)	Rico, A.; Van den Brink, P. J.; Gylstra, R.; Focks, A.; Brock, T. C. Developing Ecological
676		Scenarios for the Prospective Aquatic Risk Assessment of Pesticides. Integr. Environ.
677		Assess. Manag. 2016, 12, 510-521. https://doi.org/10.1002/ieam.1718.
678		
679		

680