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What's already known about this topic?

- Incidental diagnoses of an occult maternal malignancy have been reported upon aberrant routine non-invasive prenatal testing (NIPT).
- The presence of tumor-derived cell-free DNA in the maternal circulation can skew the NIPT profile.

What does this study add?

- Pregnant women with a confirmed neoplastic disease should not have NIPT testing for fetal aneuploidy screening since NIPT results cannot accurately be applied to assess the fetal chromosomal constitution in this condition.

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Data sharing statement: The data that support the findings of this study are available from the corresponding author upon reasonable request.

Ethics: Plasma samples for NIPT testing were collected between August 2014 and November 2018. The study was approved by the ethics committee of University Hospitals Leuven (S/57197). Written informed consent was obtained from all participants.

Main text

Noninvasive prenatal testing (NIPT), using massively parallel sequencing of plasma cell-free DNA (cfDNA), has been adopted worldwide for prenatal screening of common fetal aneuploidies¹. It is based on the analysis of fetal cfDNA fragments, derived from the placenta and freely circulating in the maternal bloodstream. Two basic sequencing approaches are currently in use to analyse circulating fetal cfDNA, namely random (whole-genome) and targeted sequencing, being outlined in¹. In the genomewide method, chromosomal ratios are calculated based on the number of sequencing reads of the chromosome of interest (e.g. chromosome 21 in the case of Down syndrome) relative to the reads of a reference chromosome in a set of normal (diploid) samples. From these ratios one z-score per chromosome is calculated to determine fetal aneuploidy. A z-score of three is commonly used as a risk threshold above which a trisomy might be suspected. Because the fraction of placenta-derived ‘fetal’ cfDNA exists against a high background of maternal plasma cfDNA, NIPT profiling not only examines fetal but also maternal cfDNA, implying that maternal chromosomal abnormalities can be detected as well². Since the introduction of NIPT in prenatal diagnostics, incidental findings of an occult maternal malignancy following a ‘false-positive’ NIPT test have been reported repeatedly. Common cancer types encountered in pregnancy (such as breast cancer, lymphoma and leukemia), but also other cancers (like ovarian cancer, multiple myeloma, digestive cancers, malignant melanoma or sarcomas) and benign tumors (uterine leiomyomas) have been accidentally identified upon aberrant NIPT testing (³⁻¹⁰ and unpublished results). From these cases, it is now appreciated that the presence of tumor-derived cfDNA can skew the NIPT profile and confound its interpretation. Three particular scenarios might be encountered. Firstly, when the observed imbalances are incompatible with fetal development, a maternal malignancy might be invoked. In a second scenario, where such imbalances are compatible with fetal development, a false positive prenatal diagnosis could be made¹⁰. This is illustrated in Table 1, representing NIPT data from a series of 26 pregnant cases that had a known diagnosis of breast cancer (n=24), colon cancer (n=1) or lymphoma (n=1), prior to participating to a research study in which genomewide NIPT testing in this cancer-in-pregnancy setting was evaluated. In six out of the 26 cases, an aberrant NIPT output with chromosomewide z-scores higher than 3 for chromosomes 21, 18 and/or 13 was observed, suggesting a fetal trisomy for (one of) the respective chromosomes. However, upon low-pass sequencing of tumor biopsy specimens of these women, it was clear that the observed gains of chromosomes 21, 18 and 13 in cfDNA were derived from tumor DNA. This resulted in false

positive scores of 15,4%, 15,4% and 19,2% for trisomy 21, 18 and 13 respectively, in this study group of pregnant cancer patients. Figure 1 visualizes the NIPT output for one of these six cases, i.e. a woman that was diagnosed with a stage II, triple negative breast cancer when being 8 weeks pregnant. When limiting the analyses to the commonly tested fetal chromosomes, z-scores higher than 3 were observed for chromosomes 21, 18 and 13. A genomewide inspection however, showed the presence of chromosomal imbalances in almost all 22 autosomes. Upon comparison with the copy number profile of matched tumor biopsy DNA, the (sub)chromosomal CNAs and aneuploidies observed in cfDNA, were shown to originate from tumor DNA. This woman gave birth to a baby boy with a normal neonatal outcome. Finally, also a NIPT outcome with an apparently normal result (for the investigated fetal chromosomes) cannot accurately be applied to assess the fetal chromosomal constitution as (i) z-scores of particular fetal chromosomes or chromosomal fragments might be skewed due to excessive presentation of highly amplified tumoral chromosomes or chromosome arms or (ii) chromosomal amplifications and deletions in the fetal and tumoral cfDNA may cancel each other out resulting in a neutral z-score for a particular chromosome. In our study cohort of pregnant cancer patients, five women had a negative z-score ($z \leq -3$) for chromosomes 21, 18, 13 or a combination of these chromosomes. Except for 1 case, all observed aneuploidies in cfDNA were shown to reflect true monosomies in the tumor DNA (Table 1). Four of these five women gave birth to a child with no congenital malformations; for the fifth case no information was available on fetal outcome. If, however, one of these children would have been affected by a true fetal trisomy (characterized by a z-score ≥ 3), then the monosomies in the tumor DNA would have neutralized the final z-score for the respective chromosomes, resulting in a false negative NIPT output. The theoretical risk of such a false negative NIPT score in our patient cohort ranged from 7,7% to 15,4% for chromosomes 21, 18 and 13 (Table 1).

Together, these examples illustrate that the presence of tumor-derived cfDNA can induce an aberrant NIPT result masking the fetal chromosomal profile. Therefore, we here advocate excluding pregnant women with a confirmed neoplastic disease from NIPT for fetal aneuploidy screening. Particular difficulties might arise with targeted NIPT assays, where information about genomewide distribution of cfDNA fragments is lacking to aid in the interpretation of deviating results of chromosomes 21, 18 and/or 13. However, even with full genome information, correct interpretation of the fetal genetic constitution might be disturbed, as shown above. Hence, NIPT testing as a screening tool for fetal aneuploidies is contraindicated in cases with a known neoplastic disease. With future novel algorithms taking into account the origin of cfDNA, advanced approaches to measure fetal fraction and improved algorithms for

aneuploidy detection, it may well become possible to identify and exclude analysis of tumor-derived cfDNA and avoid misdiagnoses. Until that time, we argue that pregnant cancer patients should be offered a detailed structural anomaly screening by ultrasound and an amniocentesis for karyotyping if certainty on chromosomal abnormalities is desired. Though not offered anymore in some centres¹¹, a combined first trimester screening can be performed to screen for trisomy 21, 13 and 18 in case of cancer diagnosis before 14 weeks.

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Table 1. Risk of false positive and false negative NIPT scores for chromosomes 21, 18 and 13 in a cohort of pregnant women with a known maternal malignancy (n=26)

	NIPT profile in plasma cfDNA			Copy number profile in tumor DNA ^(†)		
	chr21	chr18	chr13	chr21	chr18	chr13
Number of cases with normal $z > -3$ and $z < 3$	22	22	20	na	na	na
Number of cases with $z \geq 3$	2	2	2	2/2	2/2	2/2
Number of cases with $z \leq -3$	2	2	4	2/2	2/2	3/4
Percentage of false positive NIPT scores (%)	15,4%	15,4%	19,2%			
Theoretical risk of false negative NIPT scores (%)	7,7%	7,7%	15,4%			

cfDNA, cell-free DNA; chr, chromosome; na, not applicable; z, z-score. ^(†)Low-pass sequencing (0,1x coverage) of matched tumor biopsy DNA.

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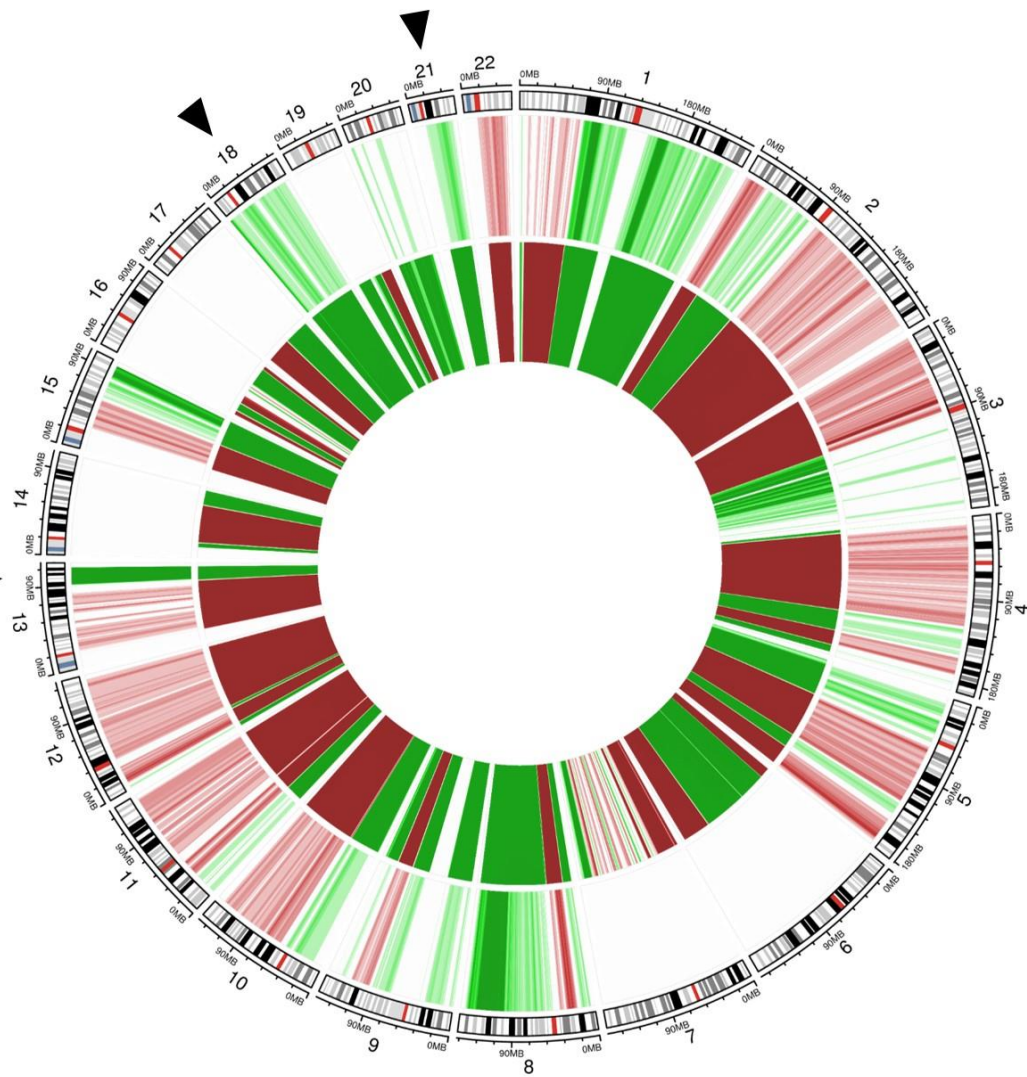


Figure 1. Circos plot showing chromosomal anomalies detectable in plasma cell-free (cfDNA) and tumor DNA of a pregnant women being 8 weeks pregnant and with a known breast cancer diagnosis. The genomic representation profile of the autosomal chromosomes is shown in clockwise order, aligned with chromosomal ideograms (outer circle). Chromosomal anomalies with a chromosomal z-score $\geq 3,0$ (suggesting gain) are indicated in green; those with a z-score $\leq 3,0$ (suggesting loss) are shown in red. Colour grades are used to indicate four z-score intervals of length 1,5 ranging from 3,0 ($-3,0$) to 9,0 (-9). The fifth darkest colour is reserved for values greater than 9 or less than -9 . The middle circle depicts the genomewide NIPT profile in plasma cfDNA with elevated z-scores for chromosomes 21, 18 and 13 (indicated by black arrows). Upon a genomewide view, (sub)chromosomal imbalances across multiple autosomal chromosomes can be observed. The inner circle shows the copy number profile of matched tumor DNA extracted from formalin-fixed paraffin-embedded tumor biopsy material (whole-genome low-pass sequencing, 0,1x coverage). Comparison of both profiles reveals that the (sub)chromosomal CNAs and aneuploidies observed in plasma cfDNA are derived from tumor DNA. Details about the NIPT data analysis pipeline can be found elsewhere¹².