

1 **WY 1048, a 17-methyl 19-nor D-ring analog of vitamin D₃, in combination with risedronate**
2 **restores bone mass in a mouse model of postmenopausal osteoporosis.**

3 Doms S.¹, Verlinden L.¹, Vanhevel J.¹, Janssens I.¹, Bouillon R.¹, De Clercq P.², Verstuyf
4 A.^{1*}

5 ¹Clinical and Experimental Endocrinology, Department of Chronic Diseases, Metabolism and
6 Ageing, KU Leuven, Belgium

7 ²Department of Organic and Macromolecular Chemistry, UGent, Belgium

8 *Corresponding author, mieke.verstuyf@kuleuven.be

9 **Abstract**

10 **Bisphosphonates like risedronate inhibit osteoclast-mediated bone resorption and are therefore**
11 **used in the prevention and treatment of osteoporosis. Also vitamin D₃ and calcium**
12 **supplementation is commonly used in the prevention or treatment of osteoporosis. Combined**
13 **therapy of risedronate with 1,25(OH)₂D₃, the active metabolite of vitamin D₃, may be**
14 **advantageous over the use of either monotherapy, but bears a risk of causing hypercalcemia**
15 **thereby decreasing the therapeutic window for osteoporosis treatment. In this study, we**
16 **evaluated the effect on bone mass of the combination of risedronate with the 17-methyl 19-nor**
17 **five-membered D-ring vitamin D₃ analog WY 1048 in a mouse ovariectomy model for**
18 **postmenopausal osteoporosis. Ovariectomy-induced bone loss was restored by administration**
19 **of risedronate or a combination of risedronate with 1,25(OH)₂D₃. However, the combination of WY**
20 **1048 with risedronate induced an even higher increase on total body and spine bone mineral**
21 **density and on trabecular and cortical bone mass. Our data indicate that combination therapy of**
22 **risedronate with WY 1048 was superior in restoring and improving bone mass over a**
23 **combination of risedronate with 1,25(OH)₂D₃ with minimal calcemic side effects.**

24 **Key words: vitamin D, risedronate, vitamin D analog, ovariectomy, osteoporosis**

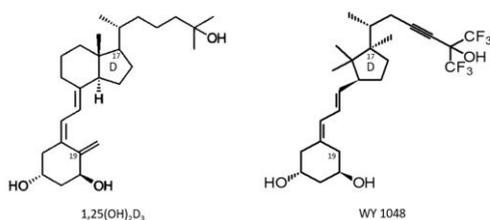
25

1 Introduction

2 Osteoporosis is a metabolic bone disease characterized by a decreased bone mass and impaired bone
3 architecture, both contributing to reduced bone strength and increased fracture risk. Although
4 osteoporosis is prevalent in both sexes, more women are affected mainly because of the loss of the
5 bone-protective effects of estrogens during menopause [1]. Among the first-line treatments for the
6 prevention and treatment of osteoporosis in post-menopausal women are the bisphosphonates such as
7 risedronate. Risedronate binds to the bone matrix where it leads to an increased osteoclast apoptosis
8 rate. Already after a few days, this increase in osteoclast apoptosis leads to decreased bone resorption
9 and a net gain in bone mass [2, 3].

10 Also the use of vitamin D₃ and calcium supplementation has been implicated in the management of
11 osteoporosis since with ageing, the capacity of the kidney to produce active vitamin D₃ declines with
12 negative consequences for calcium uptake [4]. Indeed, vitamin D₃ and, more specifically, its hormonally
13 active form, 1 α ,25-dihydroxyvitamin D₃ [1,25(OH)₂D₃], is a major regulator of calcium homeostasis by
14 stimulating intestinal calcium absorption and renal calcium re-absorption [5]. Moreover, in preclinical
15 models of postmenopausal osteoporosis active vitamin D₃ metabolites can fully prevent estrogen-
16 induced loss of trabecular and cortical bone by suppressing bone resorption [6–9]. Nevertheless, the
17 use of active vitamin D₃ metabolites in the treatment of osteoporosis remains paradoxical as they,
18 especially at higher doses, excessively stimulate intestinal calcium absorption resulting in calcemic side
19 effects [10]. In addition, during a negative calcium balance vitamin D₃ metabolites aim to maintain
20 normocalcemia by transferring calcium from the bone to the serum, which adversely impacts bone mass
21 and strength [11].

22 Interestingly, a number of recent studies have shown that the vitamin D₃ analog eldcalcitol (ED-71) is
23 a potent compound in the restoration of estrogen-induced bone loss by suppression of osteoclastic bone
24 resorption and its use for the treatment of postmenopausal osteoporosis is approved in Japan [12, 13].
25 In our laboratories, the vitamin D₃ analog WY 1048, a 17-methyl 19-nor five-membered D-ring analog
26 of 1,25(OH)₂D₃, was developed (**Figure 1**). This compound has a lower affinity (50%) for the vitamin D
27 receptor (VDR) and a weaker binding (7%) to the vitamin D binding protein compared with 1,25(OH)₂D₃.
28 Despite this lower VDR affinity, 10-fold lower concentrations of WY 1048, compared with 1,25(OH)₂D₃,
29 were sufficient to reach half-maximal inhibition of MCF-7 breast cancer cell proliferation and 3-fold lower
30 concentrations were enough to obtain half-maximal differentiation of HL60 promyelocytic cells [14, 15].
31 Evaluation of its calcemic effects revealed that a short-term treatment with 0.2 μ g/kg/d WY 1048 induced
32 a small gain in femoral calcium content without affecting serum and urinary calcium concentrations.
33 Treatment with the same dose of 0.2 μ g/kg/d 1,25(OH)₂D₃ led to overt hypercalcemia and hypercalciuria
34 without increasing femoral calcium content (Supplementary Figure 1). This initial finding led us to
35 evaluate the capacity of WY 1048 to restore bone loss in a preclinical mouse model of estrogen
36 deficiency-induced osteoporosis either alone or in combination with risedronate.



38 **Figure 1. The chemical structure of 1,25(OH)₂D₃ and of WY 1048, a 17-methyl-19-nor-five-membered D-ring**
39 **analog of 1,25(OH)₂D₃. Modified from “Analogues of calcitriol” [15].**

2 Materials and methods

2.1 Substances

WY 1048 (17-methyl-19-nor-five-membered D-ring-1 α ,25-dihydroxyvitamin D₃) was originally synthesized by Yusheng Wu, P. De Clercq and the late M. Vandewalle from the University of Ghent (Belgium). Risedronate and calcitriol were purchased from Sigma Aldrich.

2.2 Animal procedures

For the ovariectomy (OVX) experiment, 12 week old, sham-operated (n=10) and ovariectomized mice (n=100) C3H mice (Charles River Laboratories) were used. Starting from 4 weeks after the surgical procedures, sham-operated mice received arachis oil as vehicle control (group 1). OVX mice were randomized into 10 different treatment groups (10 mice/group) as shown in **Table 1** and received daily doses of vehicle (arachis oil, group 2), risedronate (3 μ g/kg/d, group 3), 1,25(OH)₂D₃ (0.05 μ g/kg/d or 0.125 μ g/kg/d, groups 4 and 5) or a combination of risedronate and calcitriol (groups 6 and 7), WY 1048 (0.2 μ g/kg/d or 0.5 μ g/kg/d, groups 8 and 9) or a combination of risedronate and WY 1048 (groups 10 and 11). All compounds were administered orally via gavage on a daily basis over a period of 8 weeks. All animals were killed by an overdose of pentobarbital anesthesia. To confirm success of ovariectomy and estrogen depletion in OVX mice, the uterus was isolated and checked for atrophy. Mice were housed in a conventional animal facility with 12 h dark/light cycles and constant temperature with food and water supplied ad libitum. All animal experiments were approved by the ethical committee of the KU Leuven.

Table 1. Animal procedures, treatment groups.

	Group (n=10)										
	1	2	3	4	5	6	7	8	9	10	11
Risedronate (μ g/kg/d)			3			3	3			3	3
1,25(OH) ₂ D ₃ (μ g/kg/d)				0.05	0.125	0.05	0.125				
WY 1048 (μ g/kg/d)								0.2	0.5	0.2	0.5

surgical procedure: sham OVX

2.3 Serum, urine, and bone biochemistry

For the OVX experiment, mice were transferred to metabolic cages (5 mice per cage) after 4 and 8 weeks of treatment to obtain 24 h urine collections. Twelve weeks after surgery (i.e. after 8 weeks of treatment) mice were euthanized and blood serum was obtained via cardiac puncture. Calcium concentrations were determined in the individual serum samples and in the collective urine samples by a colorimetric assay (Synchron CX4 system, Beckman Instruments).

2.4 Micro-computed tomography (μ CT)

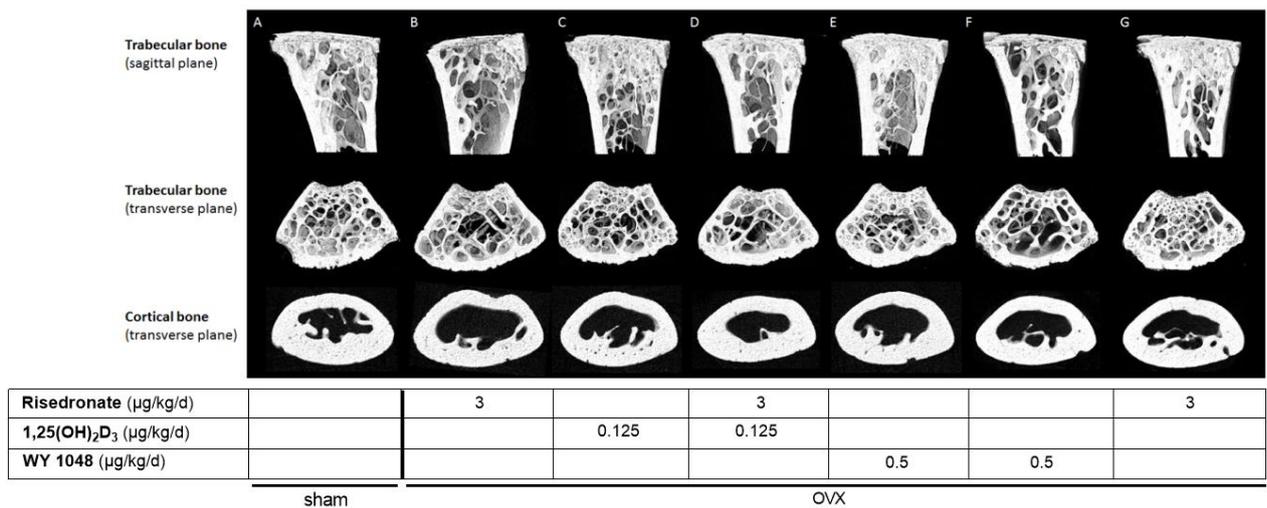
Distal femur μ CT images were taken on the high resolution Skyscan 1172 system (Bruker, Kontich, Belgium). The scanner X-ray source was set at a voltage of 50 kV and a current of 200 μ A and a 0.5 mm aluminum filter was applied. Scans were taken with a pixel size of 5 μ m, the scanning angular rotation was 180° with an angular increment of 0.4° and with a frame averaging of 2. Serial tomographs were reconstructed from raw data with the cone-beam reconstruction software (NRecon software, Skyscan, Bruker) based on the Feldkamp algorithm [16]. 3D morphometric analysis of reconstructed datasets was performed using the CT Analyzer software (CTAn, Bruker). Cortical and trabecular volumes of interest were manually selected [2 – 2.5 mm (cortical) and 0.5 – 1.5 mm (trabecular) under the selected reference point (point where trabecular bone is fully separated into four compartments on a transverse plane)]. Cortical and trabecular regions of interest were selected using the “automated trabecular and cortical bone selection method” (Method note, Bruker). Binary images for analysis were generated using a global thresholding of 80-255 (cortical) or 75-255 (trabecular). 3D models were constructed with the CTvox software (Bruker) using different selections [2.35 – 2.5 mm (cortical) and 0.25 – 2.75 mm (trabecular) under the same reference point].

1 **Figure 2. Combined therapy of risedronate and WY 1048 significantly improves both total body and spine**
 2 **BMD.** BMD was measured by DEXA for total body (A) and spine at the lumbar vertebrae L1-L5 (B) at 12 weeks
 3 post-surgery. Data are represented as mean \pm SD values ($n=6$ /group). * $p<0.05$ vs. SHAM vehicle; ° $p<0.05$ vs. OVX
 4 vehicle; # $p<0.05$ vs. risedronate; § $p<0.05$ vs. dose respective monotherapy.

5 3.2 μ CT trabecular and cortical bone

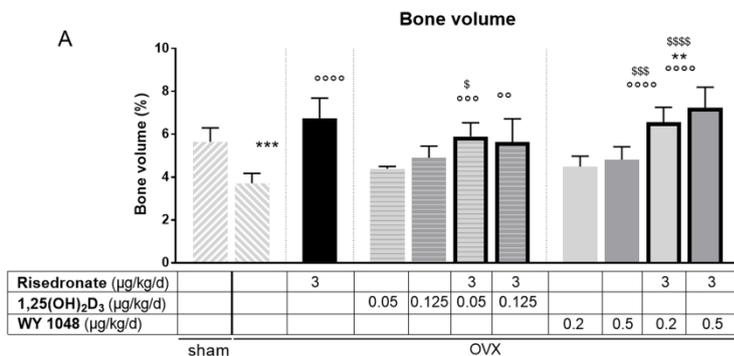
6 μ CT analysis of the distal femur confirmed that ovariectomy induced significant trabecular bone loss,
 7 which was accompanied by a decrease in trabecular number and especially by a reduction of trabecular
 8 thickness (**Figure 3, Figure 4**). Treatment with risedronate efficiently restored OVX-induced bone mass
 9 (**Figure 4A**), mainly by enhancing trabecular number (**Figure 4B**), whereas trabecular thickness was
 10 not affected (**Figure 4C**). Single treatment with either 1,25(OH)₂D₃ or WY 1048 dose-dependently
 11 enhanced trabecular bone mass in ovariectomized mice but was not as effective as risedronate. In
 12 contrast to risedronate, both 1,25(OH)₂D₃ and its analog WY 1048 had little effect on trabecular number
 13 whereas they increased trabecular thickness, and this was especially clear for the higher dose of WY
 14 1048.

15 Addition of risedronate to treatment with 1,25(OH)₂D₃ or WY 1048 led to a significant rise in trabecular
 16 bone mass, but only the combination with the higher dose of WY 1048 (0.5 μ g/kg/d) surpassed the effect
 17 of risedronate alone. Interestingly, this combination treatment resulted in a trabecular bone mass that
 18 even significantly exceeded trabecular bone mass of sham-operated, vehicle-treated mice by increasing
 19 both trabecular number and thickness (**Figure 4**).

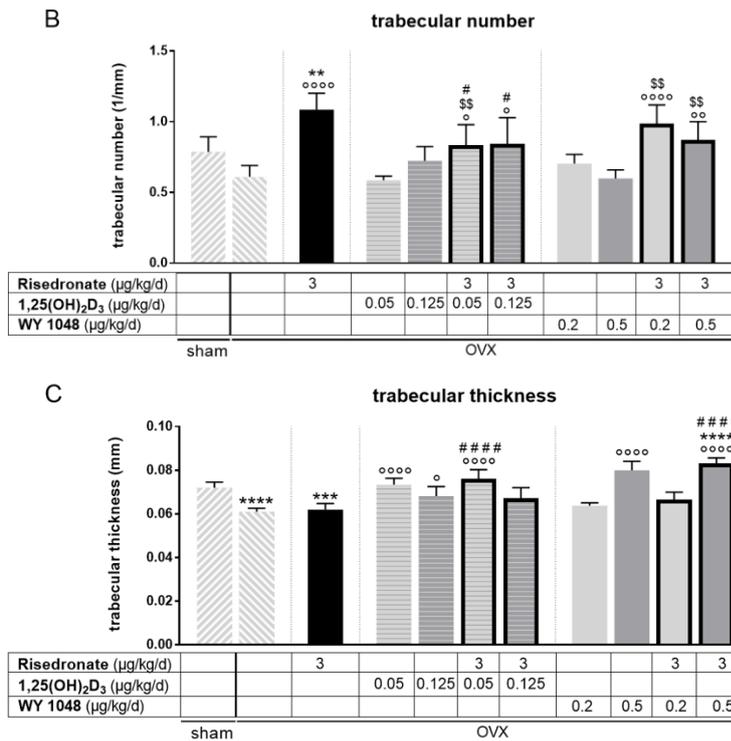


20

21 **Figure 3. Representative 3D models of trabecular bone (sagittal and transverse plane) and cortical bone**
 22 **(transverse plane) for the indicated groups. (A) SHAM vehicle, (B) OVX vehicle, (C) OVX risedronate, (D) OVX**
 23 **1,25(OH)₂D₃, (E) OVX 1,25(OH)₂D₃ + risedronate, (F) OVX WY 1048, (G) OVX WY 1048 + risedronate.**



24

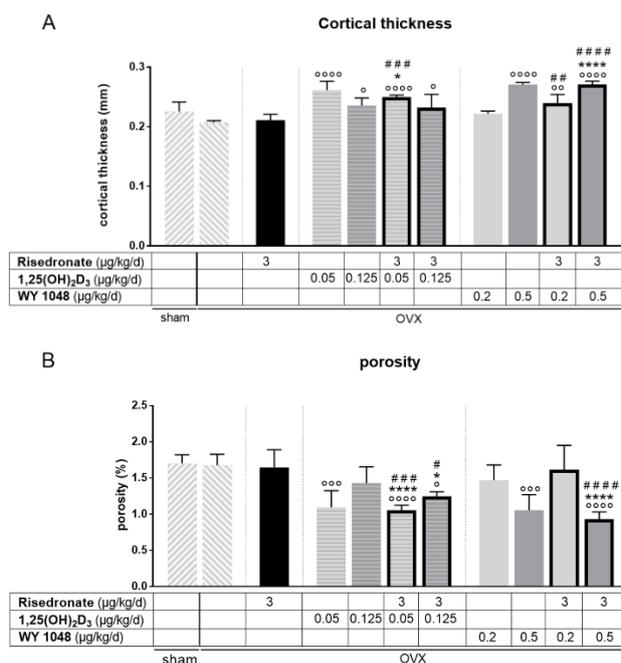


1

2

3 **Figure 4. Combination of risedronate with the highest dose of WY 1048 significantly increases bone volume**
 4 **compared to all other treatments.** Microcomputed tomography data on bone volume (A), trabecular number (B)
 5 and trabecular thickness (C) of the selected trabecular region of the distal femur in sham and OVX mice treated
 6 with the indicated drugs and doses. Values represent mean \pm SD values (n=6/group). *p<0.05 vs. SHAM vehicle;
 7 °p<0.05 vs. OVX vehicle; \$p<0.05 vs. dose respective monotherapy.

8 In this mouse model, ovariectomy did not result in cortical changes of the femur (**Figure 5**). Indeed,
 9 cortical thickness nor cortical porosity was significantly altered in OVX vehicle-treated mice. In addition,
 10 treatment with risedronate did not affect any of the cortical parameters. In contrast, 1,25(OH)₂D₃ at the
 11 lower dose of 0.05 µg/kg/d and WY 1048 at the higher dose of 0.5 µg/kg/d both significantly enhanced
 12 cortical thickness and reduced cortical porosity. Again, the combination of risedronate with the higher
 13 dose of WY 1048 resulted in a cortical thickness that exceeded that of sham-operated, vehicle-treated
 14 mice.



15

16

1 metabolites tends to suppress the calcemic side effects of 1,25(OH)₂D₃, possibly by preventing the drop
2 in PTH levels after treatment with vitamin D₃ metabolites [17, 18]. Although a combination of risedronate
3 with vitamin D₃ metabolites seems to be the ideal treatment strategy for postmenopausal osteoporosis,
4 caution has to be taken because excess dosing of 1,25(OH)₂D₃ results in increased bone resorption and
5 decreased bone mineralization, limiting the therapeutic potential of 1,25(OH)₂D₃ [18–20]. Based on our
6 observation that, for most measured bone parameters, 1,25(OH)₂D₃ at the higher dose (0.125 µg/kg/d)
7 is less effective in restoring bone mass than the lower dose (0.05 µg/kg/d) suggests that the higher dose
8 might exceed the optimal dosing. Hence, new vitamin D₃ analogs with less calcemic side effects and
9 reduced catabolic effects on bone are needed to improve treatment strategies. In this study, we
10 evaluated the capacity of our vitamin D₃ analog WY 1048 to restore bone loss and reduce calcemic side
11 effects in an OVX mouse model of postmenopausal osteoporosis, alone or in combination with
12 risedronate. Because low BMD is one of the most important predictors of a future fracture [23], we first
13 evaluated whether the combination of risedronate with WY 1048 affected BMD. Our DEXA and
14 additional pQCT data (supplementary information) indeed showed that a combined treatment with
15 risedronate and WY 1048 successfully enhanced both trabecular and cortical BMD and BMC. While
16 treatment with risedronate increased bone mass and density by increasing trabecular number, WY 1048
17 increases trabecular thickness possibly by supplying extra calcium to the bone and thereby enlarging
18 the existing mineral matrix. This apparent difference in mechanism of action may explain that the most
19 prominent increase in trabecular bone was observed when risedronate and WY 1048 were combined.
20 Knowing that the anti-fracture efficacy of a treatment is largely related to its ability to decrease cortical
21 porosity, we also evaluated cortical porosity [22, 23]. Risedronate has been shown to slow down or
22 partly reverse cortical deterioration [17]. However, we did not observe an effect on cortical parameters
23 after treatment with risedronate alone. Cortical bone has a low bone matrix volume compared to its
24 surface area as opposed to trabecular bone that has a high bone matrix volume compared to its surface
25 area [26]. Therefore, trabecular bone is more readily infiltrated by risedronate compared to cortical bone
26 and may explain why we observe a protective effect of risedronate on trabecular bone and not on cortical
27 bone. Also other studies reported that changes in cortical bone occur slower than changes in trabecular
28 bone [27]. It is therefore possible that cortical effects would have been observed when treatment with
29 risedronate was pursued for a longer time. Unexpectedly, the combination of 1,25(OH)₂D₃ and
30 risedronate seemed to reverse the calcemic effects of 1,25(OH)₂D₃ whereas this beneficial effect was
31 not observed upon a combined treatment of risedronate with WY 1048. Nevertheless, serum and urinary
32 increases in calcium levels remained minimal. Today, risedronate is often prescribed in combination
33 with vitamin D₃ to reduce the risk of fractures or to improve and speed up recovery of an existing fracture.
34 However the potential of new potent analogs deserve further studying especially because the
35 1,25(OH)₂D₃ analog, ED-71 has been shown to increase bone mass to a greater extent than the vitamin
36 D₃ analog alfacalcidol [13]. In this study, we show that a combination therapy with our vitamin D₃ analog
37 WY 1048 and risedronate represents a promising anti-osteoporotic drug in a preclinical model of
38 ovariectomy-induced bone loss. Future experiments are however needed to elucidate the underlying
39 mechanism of action.

40 **5 Acknowledgements**

41 The authors thank Suzanne Marcelis and Biau Keng Tan for all technical assistance. The authors
42 declare no conflicts of interest. This work was supported by the Flemish fund for scientific research
43 (FWO grants G.0.587.09.N.10 and G0D4217N).

6 References

- [1] K. N. Tu *et al.*, "Osteoporosis: A Review of Treatment Options.," *P T*, vol. 43, no. 2, pp. 92–104, Feb. 2018.
- [2] R. D. Chapurlat and P. D. Delmas, "Drug Insight: bisphosphonates for postmenopausal osteoporosis," *Nat. Clin. Pract. Endocrinol. Metab.*, vol. 2, no. 4, pp. 211–219, Apr. 2006.
- [3] M. J. Rogers *et al.*, "Cellular and molecular mechanisms of action of bisphosphonates.," *Cancer*, vol. 88, no. 12 Suppl, pp. 2961–78, Jun. 2000.
- [4] K. H. Lau and D. J. Baylink, "Vitamin D therapy of osteoporosis: plain vitamin D therapy versus active vitamin D analog (D-hormone) therapy.," *Calcif. Tissue Int.*, vol. 65, no. 4, pp. 295–306, Oct. 1999.
- [5] R. Nicolaysen, N. Eeg-Larsen, and O. J. Malm, "Physiology of Calcium Metabolism," *Physiol. Rev.*, vol. 33, no. 3, pp. 424–444, Jul. 1953.
- [6] K. Weber, M. Goldberg, M. Stangassinger, and R. G. Erben, "1 α -Hydroxyvitamin D₂ Is Less Toxic but Not Bone Selective Relative to 1 α -Hydroxyvitamin D₃ in Ovariectomized Rats," *J. Bone Miner. Res.*, vol. 16, no. 4, pp. 639–651, Apr. 2001.
- [7] A. Shiraishi *et al.*, "Alfacalcidol Inhibits Bone Resorption and Stimulates Formation in an Ovariectomized Rat Model of Osteoporosis: Distinct Actions from Estrogen," *J. Bone Miner. Res.*, vol. 15, no. 4, pp. 770–779, Feb. 2010.
- [8] R. G. Erben, S. Bromm, and M. Stangassinger, "Short-Term Prophylaxis against Estrogen Depletion-Induced Bone Loss with Calcitriol does not Provide Long-Term Beneficial Effects on Cancellous Bone Mass or Structure in Ovariectomized Rats," *Osteoporos. Int.*, vol. 8, no. 1, pp. 82–91, Feb. 1998.
- [9] R. G. Erben, H. Weiser, F. Sinowatz, W. A. Rambeck, and H. Zucker, "Vitamin D metabolites prevent vertebral osteopenia in ovariectomized rats.," *Calcif. Tissue Int.*, vol. 50, no. 3, pp. 228–36, Mar. 1992.
- [10] J. S. Adams and G. Lee, "Gains in bone mineral density with resolution of vitamin D intoxication.," *Ann. Intern. Med.*, vol. 127, no. 3, pp. 203–6, Aug. 1997.
- [11] A. CARLSSON, "Tracer Experiments on the Effect of Vitamin D on the Skeletal Metabolism of Calcium and Phosphorus," *Acta Physiol. Scand.*, vol. 26, no. 2–3, pp. 212–220, Apr. 1952.
- [12] I. Endo and T. Matsumoto, "[Eldecalcitol (ED-71)].," *Clin. Calcium*, vol. 21, no. 1, pp. 53–8, Jan. 2011.
- [13] Y. Uchiyama *et al.*, "ED-71, a vitamin D analog, is a more potent inhibitor of bone resorption than alfacalcidol in an estrogen-deficient rat model of osteoporosis.," *Bone*, vol. 30, no. 4, pp. 582–8, Apr. 2002.
- [14] Y. Wu, P. De Clercq, M. Vandewalle, R. Bouillon, and A. Verstuyf, "Vitamin D₃: synthesis of seco-C-9,11-bisnor-17-methyl-1 α ,25-dihydroxyvitamin D₃ analogues," *Bioorg. Med. Chem. Lett.*, vol. 12, no. 12, pp. 1633–1636, Jun. 2002.
- [15] L. Verlinden, R. Bouillon, P. De Clercq, and A. Verstuyf, "Analogues of Calcitriol," *Vitam. D*, pp. 583–614, Jan. 2018.
- [16] T. Rodet, F. Noo, and M. Defrise, "The cone-beam algorithm of Feldkamp, Davis, and Kress preserves oblique line integrals," *Med. Phys.*, vol. 31, no. 7, pp. 1972–1975, Jun. 2004.
- [17] Y. Bala *et al.*, "Risedronate Slows or Partly Reverses Cortical and Trabecular Microarchitectural Deterioration in Postmenopausal Women," *J. Bone Miner. Res.*, vol. 29, no. 2, pp. 380–388, Feb. 2014.
- [18] R. G. Erben *et al.*, "Prevention of Bone Loss in Ovariectomized Rats by Combined Treatment With Risedronate and 1 α ,25-Dihydroxyvitamin D₃," *J. Bone Miner. Res.*, vol. 17, no. 8, pp.

- 1 1498–1511, Aug. 2002.
- 2 [19] J. A. Fretz, L. A. Zella, S. Kim, N. K. Shevde, and J. W. Pike, "1,25-Dihydroxyvitamin D₃
3 Regulates the Expression of Low-Density Lipoprotein Receptor-Related Protein 5 via
4 Deoxyribonucleic Acid Sequence Elements Located Downstream of the Start Site of
5 Transcription," *Mol. Endocrinol.*, vol. 20, no. 9, pp. 2215–2230, Sep. 2006.
- 6 [20] C. A. Reasner, M. D. Stone, D. J. Hosking, A. Ballah, and G. R. Mundy, "Acute changes in
7 calcium homeostasis during treatment of primary hyperparathyroidism with risedronate.," *J.
8 Clin. Endocrinol. Metab.*, vol. 77, no. 4, pp. 1067–1071, Oct. 1993.
- 9 [21] J. A. Eisman and R. Bouillon, "Vitamin D: direct effects of vitamin D metabolites on bone:
10 lessons from genetically modified mice," *Bonekey Rep.*, vol. 3, Feb. 2014.
- 11 [22] T. J. WRONSKI, B. P. HALLORAN, D. D. BIKLE, R. K. GLOBUS, and E. R. MOREY-HOLTON,
12 "Chronic Administration of 1,25-Dihydroxyvitamin D₃: Increased Bone but Impaired
13 Mineralization*," *Endocrinology*, vol. 119, no. 6, pp. 2580–2585, Dec. 1986.
- 14 [23] S. Fujiwara, F. Kasagi, N. Masunari, K. Naito, G. Suzuki, and M. Fukunaga, "Fracture
15 Prediction From Bone Mineral Density in Japanese Men and Women," 2003.
- 16 [24] B. Borah *et al.*, "Risedronate reduces intracortical porosity in women with osteoporosis," *J.
17 Bone Miner. Res.*, vol. 25, no. 1, pp. 41–47, Jan. 2010.
- 18 [25] J. A. Kanis, O. Johnell, A. Oden, C. De Laet, A. Dawson, and B. Jonsson, "Ten Year
19 Probabilities of Osteoporotic Fractures According to BMD and Diagnostic Thresholds,"
20 *Osteoporos. Int.*, vol. 12, no. 12, pp. 989–995, Dec. 2001.
- 21 [26] A. J. Roelofs *et al.*, "Influence of bone affinity on the skeletal distribution of fluorescently
22 labeled bisphosphonates in vivo," *J. Bone Miner. Res.*, vol. 27, no. 4, pp. 835–847, Apr. 2012.
- 23 [27] Å. Bjørnerem *et al.*, "Remodeling markers are associated with larger intracortical surface area
24 but smaller trabecular surface area: A twin study," *Bone*, vol. 49, no. 6, pp. 1125–1130, Dec.
25 2011.
- 26

1 Supplementary information

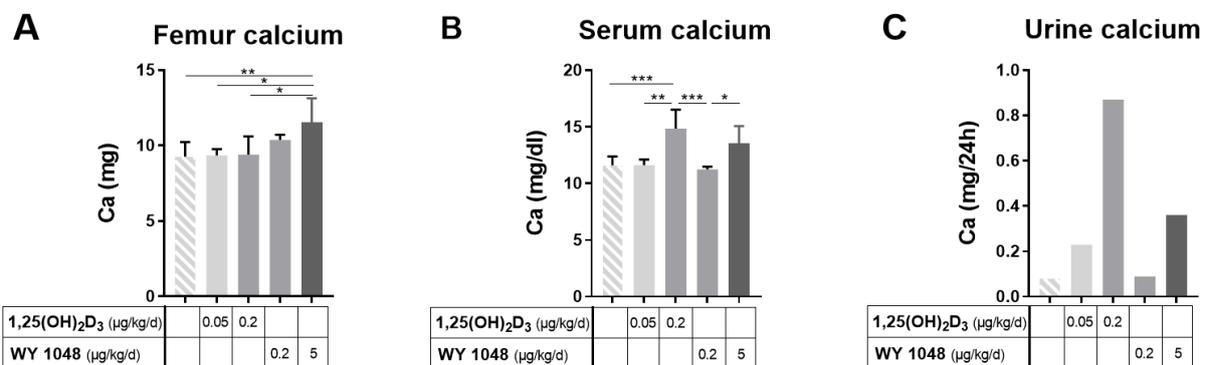
2 Toxicity test

3 Materials and methods

4 Eight week old female NMRI mice (Charles River Laboratories) were intraperitoneally injected for 7
5 consecutive days with either vehicle (arachis oil, n=6), 1,25(OH)₂D₃ (0.05 µg/kg/d or 0.2 µg/kg/d, n=6),
6 or WY 1048 (0.2 µg/kg/d or 5 µg/kg/d, n=5). Mice were then transferred to metabolic cages (5-6 mice
7 per cage, resulting in 1 urine sample per experimental group) to obtain 24 h urine collections. Thereafter,
8 mice were euthanized and blood was collected via cardiac puncture. Femurs were dissected, snap-
9 frozen in liquid N₂ and stored at -80°C for the assessment of calcium content.

10 For the assessment of femoral calcium content, femurs were dried overnight at 100°C and ashed for 8
11 h at 500°C. Femoral ashes were dissolved overnight in 1 ml 1N HCl and diluted 1/50 in milliQ water for
12 the measurement of calcium concentration.

13 figures



14

15 **Supplementary figure 1. Toxicity evaluation of WY1048 revealed that this compound resulted in a gain in**
16 **femoral calcium content with limited effects on serum or urinary calcium levels.** Mice (n=5-6/group) were
17 treated with the indicated drugs and doses during 7 consecutive days. Femoral (A) and serum calcium (B) data are
18 represented as mean ± SD values. Urine calcium (C) data is represented as mean. ***p<0.001, ****p<0.0001
19 between the indicated groups.

20

21 Peripheral quantitative computed tomography (pQCT)

22 Materials and methods

23 *Ex vivo* bone densitometry of the femur was determined by pQCT. Trabecular bone parameters
24 and cortical bone parameters were assessed using the Stratec XCT Research M+
25 densitometer (Norland Medical Systems, Fort Atkinson, WI, USA). Slices of 0.2 mm thickness
26 were scanned using a voxel size of 0.070 mm. Three scans were taken from the distal end of
27 the femur *ex vivo* (at 2.5 ± 0.25 mm), using contmode 1, peelmode 20, and a density threshold
28 of 280 mg/cm³. The trabecular bone region was defined by setting an inner threshold
29 corresponding to 30% of the total cross-sectional area. These three metaphyseal scans were
30 performed to measure the average trabecular volumetric BMD and BMC. A second scan was
31 taken 7 mm from the distal end of the femur using separation mode 1 and a density threshold
32 of 710 mg/cm³. These mid-diaphyseal scans were performed to determine cortical volumetric
33 BMD and BMC and cortical thickness.

