

Systematic Review

Regenerative potential of leucocyte- and platelet-rich fibrin. Part A: intra-bony defects, furcation defects and periodontal plastic surgery. A systematic review and meta-analysis

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Castro AB, Meschi N, Temmerman A, Pinto N, Lambrechts P, Teughels W, Quirynen M. Regenerative potential of leucocyte- and platelet-rich fibrin. Part A: intra-bony defects, furcation defects and periodontal plastic surgery. A systematic review and meta-analysis. *J Clin Periodontol* 2017; 44: 67–82. doi: 10.1111/jcpe.12643

Abstract

Aim: To analyse the regenerative potential of leucocyte- and platelet-rich fibrin (L-PRF) during periodontal surgery.

Materials and Methods: An electronic and hand search were conducted in three databases. Only randomized clinical trials were selected and no follow-up limitation was applied. Pocket depth (PD), clinical attachment level (CAL), bone fill, keratinized tissue width (KTW), recession reduction and root coverage (%) were considered as outcome. When possible, meta-analysis was performed.

Results: Twenty-four articles fulfilled the inclusion and exclusion criteria. Three subgroups were created: intra-bony defects (IBDs), furcation defects and periodontal plastic surgery. Meta-analysis was performed in all the subgroups. Significant PD reduction (1.1 ± 0.5 mm, $p < 0.001$), CAL gain (1.2 ± 0.6 mm, $p < 0.001$) and bone fill (1.7 ± 0.7 mm, $p < 0.001$) were found when comparing L-PRF to open flap debridement (OFD) in IBDs. For furcation defects, significant PD reduction (1.9 ± 1.5 mm, $p = 0.01$), CAL gain (1.3 ± 0.4 mm, $p < 0.001$) and bone fill (1.5 ± 0.3 mm, $p < 0.001$) were reported when comparing L-PRF to OFD. When L-PRF was compared to a connective tissue graft, similar outcomes were recorded for PD reduction (0.2 ± 0.3 mm, $p > 0.05$), CAL gain (0.2 ± 0.5 mm, $p > 0.05$), KTW (0.3 ± 0.4 mm, $p > 0.05$) and recession reduction (0.2 ± 0.3 mm, $p > 0.05$).

Conclusions: L-PRF enhances periodontal wound healing.

Key words: bone regeneration; gingival recession; intra-bony defects; leucocyte–platelet-rich fibrin; open flap debridement; platelet-rich fibrin; tissue regeneration

Accepted for publication 22 October 2016

Conflict of interest and source of funding statement

The authors have stated explicitly that there are no conflicts of interest in connection with this article.

The study was self-funded by the authors and their institution.

In the last 20 years, platelet concentrates (PCs) have emerged as a potential regenerative material, used alone or as scaffold for other graft materials. PCs are blood extracts, obtained after processing a whole blood sample, mostly through centrifugation (Dohan

et al. 2014a). In 1970, Matras (1970) published the first article on PCs using fibrin glue to improve skin wound healing. But it was not until Marx's studies (Marx et al. 1998, Marx 2001) that the use of PCs also gained interest in oral and maxillofacial surgery. Since then,

different techniques have been developed and with them, a variety of preparations. The first PCs generation (Fig. 1) include platelet-rich plasma (PRP) and plasma rich in growth factors (PRGF). Their preparation requires anticoagulants at the moment of blood collection to avoid coagulation. Consequently, the fibrin polymerization occurs rapidly, resulting in a weak fibrin network (Dohan et al. 2006a). They are used as liquid solution or in gel form after adding bovine thrombin and calcium chloride. Due to the difficulties in the preparation and the inconsistent outcome of PRP and PRGF formulations, a second PCs generation was introduced in 2001 by Choukroun and co-workers (Choukroun 2001, Dohan et al. 2006a, 2014a). The use of platelet-rich fibrin (PRF) is simple and requires neither anticoagulant, bovine thrombin nor calcium chloride. It is nothing more than centrifuged blood without any additives (Table S1). Whole blood is centrifuged without anticoagulants at high spin so that three layers are obtained: red blood corpuscles (RBCs) at the bottom of the tube, platelet-poor plasma (PPP) on the top and an intermediate layer called “buffy coat” where most leucocytes and platelets are concentrated.

This buffy coat or L-PRF is a bioactive construct that stimulates

the local environment for differentiation and proliferation of stem and progenitor cells (Dohan et al. 2006b). It acts as an immune regulation node with inflammation control abilities, including a slow continuous release of growth factors over a period of 7–14 days (Dohan et al. 2006c). Rich in fibrin, platelets ($\pm 95\%$ of initial blood), leucocytes ($\pm 50\%$ of initial blood), monocytes and stem cells, L-PRF can be further transformed into a membrane, circa 1 mm in thickness, by careful compression (Dohan et al. 2010) (Appendix S1). Its strong fibrin architecture and its superior mechanical properties distinguish it from other kinds of PCs (Khorshidi et al. 2016). PRP, for example, has a thin and non-condensed fibrin network with a low tensile strength so that it is less useful as a space maintainer (Burnouf et al. 2013). The strong fibrin network in L-PRF is explained by the physiological concentrations of thrombin during its preparation. Rowe et al. (2007) concluded that a high thrombin concentration resulted in a high-interconnected fibre mesh with a fine fibre structure. However, as thrombin concentration decreased, fibre size increased as well as the mechanical properties. Apart from the biological and mechanical

properties, antimicrobial effects have also been described (Yang et al. 2015).

The main aim of this systematic review was to study the beneficial effect of L-PRF used as sole filling material and as adjunct to conventional techniques in periodontal surgery.

Materials and Methods

The protocol of this systematic review was based on the guidelines of the Belgian Centre for Evidence-Based Medicine (CEBAM), Belgian Branch of the Dutch Cochrane Centre. It was conducted in accordance with the Transparent Reporting of Systematic Reviews and Meta-analyses (PRISMA statement, Moher et al. 2009).

Focused PICO question

The following statements were used to conduct the systematic search:

- Population (P) = systemically healthy humans (ASA I) with loss of periodontal tissues.
- Intervention (I) = use of L-PRF (protocol 2700 r.p.m./12 min. or 3000 r.p.m./10 min.) as sole biomaterial or in combination to other biomaterials in periodontal surgery.
- Comparison (C) = traditional techniques: open flap debridement with or without grafting, periodontal plastic surgery via coronally advanced flap, with or without connective tissue graft.
- Outcome (O) = alveolar bone and/or periodontal wound healing.

A PICO question was created to define the search strategy: *Does L-PRF promote periodontal wound healing in systemically healthy patients (ASA I) during periodontal surgery compared to traditional techniques?*

Search strategy

An electronic search was performed in three Internet databases: the National Library of Medicine, Washington, DC (MEDLINE-PubMed), EMBASE (Excerpta Medical Database by Elsevier), and Cochrane Central Register of Controlled Trials (CENTRAL). The search terms were defined by combining (Mesh Terms OR Key Words) from “Population” AND (Mesh Terms OR Key Words) from

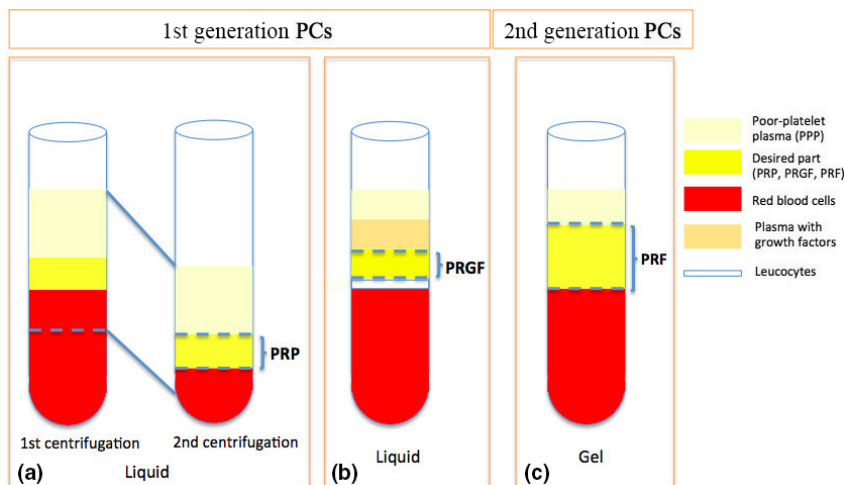


Fig. 1. Differences among PCs preparation. (a) platelet-rich plasma (PRP): after the first centrifugation, the platelet-poor plasma, the “yellow” part called buffy coat and a few red blood cells are carefully collected (pipetting) and centrifuged again in order to obtain the PRP (Dohan et al. 2006a,b,c); (b) PRGF: after centrifugation, the blood is divided in five layers; by pipetting, the undesired parts are discarded; the most concentrated part with growth factors (PRGF) is collected (Anitua, 2001); (c) PRF: after centrifugation, a fibrin clot is obtained in the middle of the tube, which is ready to be used (Dohan et al. 2006a).

“Intervention”, as shown in Table S2.

The search was limited to studies involving humans. No language or time restrictions were applied in the first search. However, only studies in English were included for selection. No follow-up limitations were used. The last electronic search was performed on the 31st of July 2015.

This search was enriched by hand searches, citation screening and expert recommendations. All reference lists of selected papers as well as related reviews were scanned for possible additional studies.

Screening and selection

The titles and abstracts obtained from the first search were screened independently by two reviewers (A.C., N.M.). When publications did not meet the inclusion criteria, they were excluded upon reviewer’s agreement. Any disagreement between the two reviewers was resolved by discussion. All full texts of the eligible articles were obtained and examined by both reviewers. The articles that fulfilled all selection criteria were processed for data extraction. Given some variability in the preparation of L-PRF, two different protocols (2700 r.p.m./12 min. or 3000 r.p.m./10 min.) were included. The inclusion and exclusion criteria are summarized in Table S3.

Assessment of heterogeneity

The heterogeneity of the included studies was judged based on following factors: (1) study design and evaluation period, (2) subject characteristics and smoking habits, and (3) surgical protocol used: (a) centrifugation protocol (2700 r.p.m./12 min. or 3000 r.p.m./10 min.), (b) mL blood used to prepare L-PRF and (c) number of clots/membranes (if used).

Quality assessment

The quality assessment, performed by both reviewers (A.C., N.M.), was based on the Cochrane Collaboration’s tool for assessing risk of bias. Six quality criteria were verified: (1) sequence generation or randomization component, (2) allocation

concealment, (3) blinding of participants, personnel and outcome assessors, (4) incomplete/missing outcome data, (5) selective outcome reporting and (6) other sources of bias. In case of any doubt, the authors were contacted for clarification or to provide missing information. Low risk of bias was indicated if all quality criteria were “present”, moderate risk of bias if one or more key domains were “unclear” and high risk of bias if one or more key domains were “absent”.

Data analysis

The analysed variables were as follows: pocket depth (PD) reduction, clinical attachment level (CAL) gain, bone fill (mm and %), keratinized tissue width (KTW) gain, tissue thickness gain, recession reduction and root coverage (%) at 6 months. For all variables in each group, mean values and standard deviation (SD) were extracted. All data were arranged in groups for the intergroup comparison (L-PRF *versus* control group). When possible, a

meta-analysis was performed. The mean difference was calculated and a 95% confidence interval (CI) was computed. Forest plots were created to display the analysis.

Results

Search and selection

As a result of the electronic and hand search, 205 articles were obtained, of which 23 were duplicate and consequently removed (Fig. 2). A total of 182 articles was included for title and abstract screening. From those, 25 articles were included for full text review. One article was excluded after full text screening, which was conducted independently by two reviewers (A.C., N.M.) (Table S4). Twenty-four randomized control trials (RCTs) fulfilled the inclusion criteria and were included for analysis.

The included articles were classified into three subgroups, depending on the indication for the use of L-PRF (Tables 1–3):

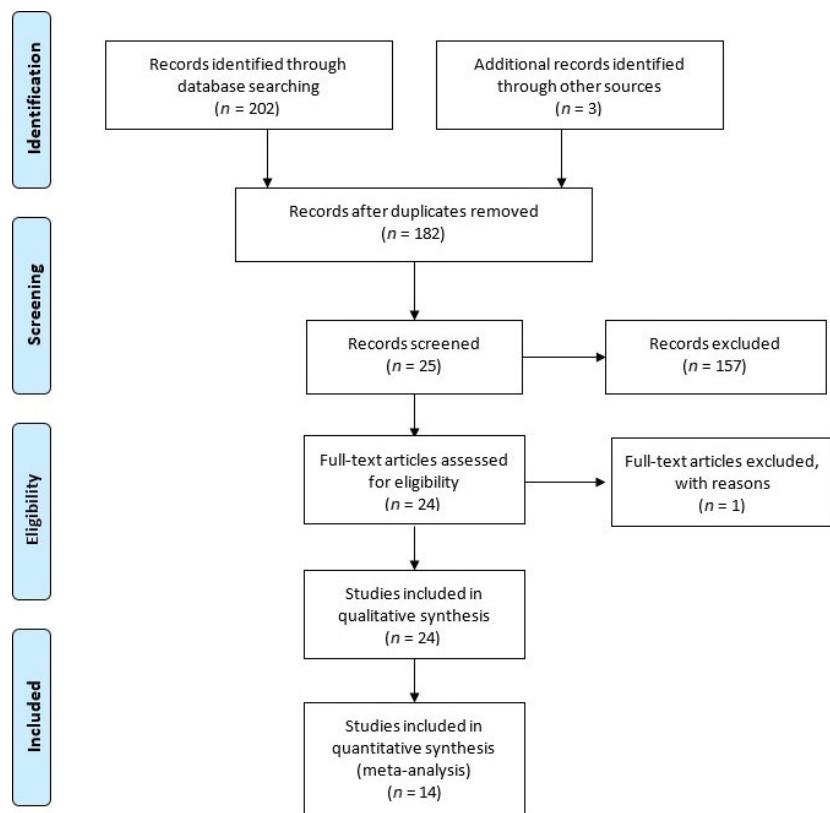


Fig. 2. PRISMA flow diagram.

- Intra-bony defect fill: $n = 13$
 - L-PRF *versus* open flap debridement (OFD): $n = 5$, Sharma & Pradeep (2011b), Thorat et al. (2011), Rosamma et al. (2012), Ajwani et al. (2015), and Pradeep et al. (2015).
 - L-PRF *versus* PRP *versus* OFD: $n = 1$, Pradeep et al. (2012).
 - L-PRF *versus* bovine porous bone mineral (BPBM): $n = 1$, Lekovic et al. (2012).
 - L-PRF *versus* demineralized freeze-dried bone allograft (DFDBA): $n = 3$, Bansal & Bharti (2013), Shah et al. (2015), and Agarwal et al. (2016).
 - L-PRF *versus* Emdogain®: $n = 1$, Gupta et al. (2014).
 - L-PRF *versus* nano-bone®: $n = 1$, Elgandy & Abo Shady (2015).
 - L-PRF *versus* autologous bone graft (ABG): $n = 1$, Mathur et al. (2015).
- Furcation defects: $n = 2$, Sharma & Pradeep (2011a), and Bajaj et al. (2013).
- Periodontal plastic surgery: $n = 9$
 - Coronally advanced flap (CAF) *versus* CAF + L-PRF: $n = 4$, Aroca et al. (2009), Padma et al. (2013), Gupta et al. (2015), and Thamaraiselvan et al. (2015).
 - CAF + L-PRF *versus* CAF + connective tissue graft (CTG): $n = 4$, Jankovic et al. (2012), Eren & Atilla (2014), Keceli et al. (2015), and Tunali et al. (2015).
 - CAF + L-PRF *versus* CAF + Emdogain® (EMD): $n = 1$, Jankovic et al. (2010).

Assessment of heterogeneity

Study design and evaluation period

All studies were RCTs and frequently presented a split-mouth design. The articles with these characteristics are the following: intra-bony defects (IBDs) 7/13 (Lekovic et al. 2012, Rosamma et al. 2012, Bansal & Bharti 2013, Ajwani et al. 2015, Elgandy & Abo Shady 2015, Shah et al. 2015, Agarwal et al. 2016), furcation defects 1/2 (Sharma

& Pradeep 2011a), plastic surgery 7/9 (Aroca et al. 2009, Jankovic et al. 2010, 2012, Padma et al. 2013, Eren & Atilla 2014, Keceli et al. 2015, Tunali et al. 2015). The follow-up ranged slightly (IBDs 6–12 months, furcation defects 9 months and plastic surgery 6–12 months).

Subject characteristics and smoking habits

Healthy subjects with no active periodontal disease were included in all the studies. The studies that did not include smokers are the following: IBDs 9/13 (Sharma & Pradeep 2011a, b, Lekovic et al. 2012, Rosamma et al. 2012, Pradeep et al. 2012, 2015, Gupta et al. 2014, Ajwani et al. 2015, Shah et al. 2015, Agarwal et al. 2016), furcation defects 1/2 (Sharma & Pradeep 2011a), plastic surgery 8/9 (Jankovic et al. 2010, 2012, Padma et al. 2013, Eren & Atilla 2014, Gupta et al. 2015, Keceli et al. 2015, Thamaraiselvan et al. 2015, Tunali et al. 2015).

Surgical protocol

A wide variety of surgical protocols was used. This heterogeneity can be derived from Tables 1–3.

Quality assessment

Appendix S2–S4 shows the quality assessment for the included studies. All articles on furcation defects and periodontal plastic surgery showed a moderate risk of bias. Similarly, 12 articles using L-PRF in IBD had a moderate risk, and one had a low risk of bias.

Quantitative assessment

The extracted data were continuous. The articles with split-mouth design and parallel design were not analysed separately. The control group and test group from the articles with split-mouth design were considered as independent. As shown in the Figs 3 and 4, the studies with split-mouth design do not differ from those with parallel design. Random effects were used due to the heterogeneity of the data.

Intra-bony defects

In the articles on IBDs, benefits in terms of PD reduction, CAL gain and bone fill were shown when L-PRF was used alone or in

combination with other biomaterials (Table 1). Six out of 13 articles (Sharma & Pradeep 2011b, Thorat et al. 2011, Pradeep et al. 2012, 2015, Rosamma et al. 2012, Ajwani et al. 2015) could be used for a meta-analysis since they reported on similar outcome measures comparing OFD to OFD + L-PRF (Fig. 3a–c). The meta-analysis of IBDs showed a statistical significant difference for PD reduction (mean difference: 1.1 mm, $p < 0.001$, CI: 0.6–1.6), CAL gain (mean difference: 1.2 mm, $p < 0.001$, CI: 0.5–1.9), amount of bone fill in mm (mean difference: 1.7 mm, $p < 0.001$, CI: 1.0–2.3) and bone fill when scored as % (mean difference: 46.0%, $p < 0.001$, CI: 33.2–58.7), all in favour of L-PRF.

Furcation defects

Two articles were included for furcation defects (Sharma & Pradeep 2011a, Bajaj et al. 2013). A meta-analysis could be performed for both articles, comparing OFD to OFD + L-PRF (Fig. 3d,e). Statistical significant differences could be found for PD reduction (mean difference: 1.9 mm, $p = 0.01$, CI: 0.4–3.5), CAL gain (mean difference: 1.3 mm, $p < 0.001$, CI: 0.8–1.7), amount of bone fill in mm (mean difference: 1.5 mm, $p < 0.001$, CI: 1.2–1.9), bone fill when scored as % (37.6%, $p < 0.001$, CI: 30.6–44.5), again in favour of L-PRF (Table 2).

Periodontal plastic surgery

In case of a CAF, some studies reported some benefits when L-PRF membranes were added, but others failed to show this advantage (Table 3). When the use of a CTG in a CAF procedure was compared to the use of L-PRF membranes, similar results were obtained. Two meta-analyses could be performed, one comparing a CAF alone *versus* a CAF with L-PRF, and another comparing a CAF with L-PRF *versus* a CAF with a CTG. The following variables were considered: PD reduction, CAL gain, KTW gain, tissue thickness gain, recession reduction and root coverage at 6 months.

For the first comparison (CAF + L-PRF *versus* CAF, Fig. 4a,b), three articles could be included for a meta-analysis (Aroca et al. 2009, Gupta et al. 2015, Thamaraiselvan et al. 2015). The analysis showed no

Table 1. L-PRF for intra-bony defects. Papers have been arranged by subapplications (L-PRF + OFD *versus* OFD, L-PRF *versus* PRP, L-PRF *versus* L-PRF + BPBM, L-PRF + DFDBA *versus* DFDBA, L-PRF *versus* Emdogain[®], L-PRF *versus* nano-bone[®], L-PRF *versus* ABG, L-PRF in furcation lesions: L-PRF + OFD *versus* OFD)

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
L-PRF + OFD <i>versus</i> OFD Thorat et al. (2011)	RCT Parallel Single-blind 9 months	40 – (32) 18 ♀, 22 ♂ Mean age: 31 ± 2 Range: ? Smoking: ?	2 and 3 walls IBDs C: n = 16, OFD T: n = 16, OFD + L-PRF	Hardware: ^a Setting: 400 g/12 min.	1 L-PRF clot 1 L-PRF membrane 10 ml blood/clot	L-PRF + OFD versus OFD SS more PD reduction (4.5 <i>versus</i> 3.5 mm), CAL gain (3.7 <i>versus</i> 2.1 mm) and bone fill (47% <i>versus</i> 29%) in favour of L-PRF group (<i>p</i> < 0.05). L-PRF + OFD versus OFD SS more PD reduction (4.5 <i>versus</i> 3.2 mm) and bone fill (48.2% <i>versus</i> 1.8%) in L-PRF group (<i>p</i> < 0.001). NSS CAL gain between groups (3.1 <i>versus</i> 2.7 mm) (<i>p</i> > 0.05). L-PRF + OFD versus OFD SS PD reduction (4.6 <i>versus</i> 2.4 mm), CAL gain (4.7 <i>versus</i> 1.4 mm) and radiographic intra-bony defect depth (1.9 <i>versus</i> 0.6 mm) in favour of L-PRF sites (<i>p</i> < 0.00).
Sharma & Pradeep (2011b)	RCT Parallel Double-blind 9 months	42 – (35) 18 ♀, 24 ♂ Mean age: 35 ± 6 Range: 30–50 Smoking: No	3 walls IBDs C: n = 17 (28 sites), OFD T: n = 18 (28 sites), L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF clot 2 L-PRF membrane 10 ml blood/clot	
Rosamma et al. (2012)	RCT Split-mouth Not blind 12 months	15 – (15) 9 ♀, 6 ♂ Mean age: 29 ± 7 Range: 17–44 Smoking: No	3 walls IBDs C: n = 15, OFD T: n = 15, OFD + L-PRF	Hardware: ^b Setting: 3000 r.p.m./10 min.	1 L-PRF clot 0 L-PRF membrane 10 ml blood/clot	

Table 1. (continued)

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? , No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
Ajwani et al. (2015)	RCT Split-mouth Single-blind 9 months	20 – (20) 10 ♀, 10 ♂ Mean age: 30.5 Range: ? Smoking: No	2 and 3 walls IBDs C: <i>n</i> = 20, OFD T: <i>n</i> = 20, OFD + L-PRF	Hardware: ^c Setting: 3000 r.p.m./10 min.	1 L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF + OFD versus OFD NSS improvement in PD (1.9 <i>versus</i> 1.6 mm) and CAL (1.8 <i>versus</i> 1.3 mm) (<i>p</i> > 0.05). SS more bone fill (2.6 <i>versus</i> 1.3 mm) in favour of L-PRF group (<i>p</i> < 0.05). L-PRF + OFD versus OFD SS PD reduction (4.0 <i>versus</i> 3.0 mm), CAL gain (4.0 <i>versus</i> 2.9 mm) in favour or T ₁ compared to C (<i>p</i> < 0.05).
Pradeep et al. (2015)	RCT Parallel Triple-blind 9 months	126 – (120) 60 ♀, 60 ♂ Mean age: 41 ± 6 Range: 30–50 Smoking: No	3 walls IBDs C: <i>n</i> = 30, OFD T ₁ : <i>n</i> = 30, OFD + L-PRF T ₂ : <i>n</i> = 30, OFD + 1% MF T ₃ : <i>n</i> = 30, OFD + L-PRF + 1% MF	Hardware: ? Setting: 3000 r.p.m./10 min.	2 L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF + OFD versus OFD SS PD reduction (4.0 <i>versus</i> 3.0 mm), CAL gain (4.0 <i>versus</i> 2.9 mm) in favour or T ₁ compared to C (<i>p</i> < 0.05).
L-PRF <i>versus</i> PRP Pradeep et al. (2012)	RCT Parallel Double-blind 9 months	54 – (50) 27 ♀, 27 ♂ Mean age: 36.8 Range: ? Smoking: No	3 wall IBDs C: <i>n</i> = 17 (30 sites), OFD T ₁ : <i>n</i> = 16 (30 sites), L-PRF T ₂ : <i>n</i> = 17 (30 sites), PRP	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF clot 2 L-PRF membrane 10 ml blood/clot	L-PRF <i>versus</i> PRP SS PD reduction (T ₁ : 3.7 <i>versus</i> T ₂ : 3.7 <i>versus</i> C: 2.7 mm) and bone fill (T ₁ 55% <i>versus</i> C: 2.9 mm) and bone fill (T ₁ 55% <i>versus</i> T ₂ : 56% <i>versus</i> C: 1.5%) in favour of L-PRF and PRP groups (<i>p</i> < 0.05). NSS CAL gain (T ₁ : 3.17 <i>versus</i> T ₂ : 2.9 <i>versus</i> C: 2.9 mm) (<i>p</i> > 0.05).

Table 1. (continued)

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? , No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
L-PRF versus L-PRF + BPBM Lekovic et al. (2012)	RCT Split-mouth Double-blind 6 months	17 – (17) 11 ♀, 6 ♂ Mean age: 44 ± 9 Range: ? Smoking: 12 non smokers/5 smokers	2 and 3 walls IBDs C: <i>n</i> = 17, L-PRF T: <i>n</i> = 17, L-PRF + BPBM	Hardware: ^d Setting: 1000 g/10 min.	1 L-PRF clot 1 L-PRF membrane 10 ml blood/clot	L-PRF versus L-PRF + BPBM SS PD reduction (4.4 versus 3.3 mm), CAL gain (2.4 versus 3.8 mm), and bone fill (2.1 versus 4.6 mm) in favour of L-PRF-BPBM group (<i>p</i> < 0.001).
L-PRF + DFDBA versus DFDBA Bansal & Bharti (2013)	RCT Split-mouth Not blind 6 months	10 – (10) Gender: ? Mean age: ? Range: ? Smoking: ?	Walls IBDs not mentioned C: <i>n</i> = 10, DFDBA T: <i>n</i> = 10, L-PRF + DFDBA	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF + DFDBA versus DFDBA SS PD reduction (4.0 versus 3.1 mm) and CAL gain (3.4 versus 2.3 mm) in favour of L-PRF group (<i>p</i> < 0.05). NNSD for bone fill (2.3 versus 1.9 mm) and alveolar crest resorption (0.02 versus 0.04 mm) (<i>p</i> > 0.01).
Shah et al. (2015)	RCT Split-mouth Not blind 6 months	20 – (20) Gender: ? Mean age: ? Range: 20–55 Smoking: No	2 and 3 walls IBDs C: <i>n</i> = 20, OFD + DFDBA T: <i>n</i> = 20, OFD + L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	? L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF + DFDBA versus DFDBA NSS PD reduction (3.6 versus 3.7 mm), CAL (2.9 versus 2.9 mm) and GML (–0.4 versus –0.3 mm) L-PRF + DFDBA versus DFDBA SS PD reduction (4.2 versus 3.6 mm), CAL gain (3.7 versus 2.6 mm), REC (0.5 versus 1.0 mm), bone fill (3.5 versus 2.5 mm) and defect resolution (3.7 versus 2.7 mm) in favour of DFDBA + L-PRF group (<i>p</i> < 0.05).
Agarwal et al. (2016)	RCT Split-mouth Double-blind 12 months	32 – (30) 14 ♀, 18 ♂ Mean age: 52 ± 7 Range: ? Smoking: No	2 and 3 walls IBDs C: <i>n</i> = 32, DFDBA + saline T: <i>n</i> = 32, DFDBA + L-PRF	Hardware: ? Setting: 400 g/12 min.	1? L-PRF clot >1 L-PRF membrane 10 ml blood/clot	

Table 1. (continued)

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? , No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
L-PRF <i>versus</i> Emdogain® Gupta et al. (2014)	RCT 6 months	30 – (30) 15 ♀, 15 ♂ Mean age: ? Range: 30–65 Smoking: No	3 walls IBDs C: <i>n</i> = 22, OFD + Emdogain® T: <i>n</i> = 22, OFD + L-PRF	Hardware: ^c Setting: 3000 r.p.m./12 min.	? L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF <i>versus</i> Emdogain® NSS PD reduction (1.8 <i>versus</i> 1.8 mm) and CAL gain (2.0 <i>versus</i> 1.8 mm) (<i>p</i> > 0.05). SS more defect resolution in Emdogain® group (43% <i>versus</i> 32%) (<i>p</i> < 0.05).
L-PRF <i>versus</i> nano-bone® Elgendy & Abo Shady (2015)	RCT Split-mouth Not blind 6 months	20 – (20) Gender: ? Mean age: C: 40 ± 6, T: 44 ± 8 Range: ? Smoking: No or light smokers (<10 cig/day)	Walls IBDs not mentioned C: <i>n</i> = 20, OFD + nano-bone® T: <i>n</i> = 20, L-PRF + nano-bone®	Hardware: ? Setting: 3000 r.p.m./ 10 min.	? L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF <i>versus</i> nano-bone® SS PD reduction (7.1 <i>versus</i> 6.7 mm) and CAL gain (7.4 <i>versus</i> 7.1 mm) in favour of L-PRF group (<i>p</i> < 0.01).
L-PRF <i>versus</i> ABG Mathur et al. (2015)	RCT Parallel Not blind 6 months	25 – (25) 11 ♀, 14 ♂ Mean age: 40 ± 5 Range: ? Smoking: ?	3 walls IBDs C: <i>n</i> = 19, OFD + ABG T: <i>n</i> = 19, OFD + L-PRF	Hardware: ^c Setting: 3000 r.p.m./ 10 min.	? L-PRF clot 0 L-PRF membrane 10 ml blood/clot	L-PRF <i>versus</i> ABG NSS PD reduction (2.6 <i>versus</i> 2.4 mm), and CAL gain (2.5 <i>versus</i> 2.6 mm) (<i>p</i> > 0.05).

ABG, autologous bone graft; BPBM, Bovine porous bone mineral; C, control group; CAL, clinical attachment level; DFDBA, demineralized freeze-dried bone allograft; IBDs, intra-bony defects; NSS, no statistically significant; OFD, open flap debridement; PD, pocket depth; PRP, platelet-rich plasma; REC, gingival recession; SS, statistically significant; T, test group.

^aProcess protocol, Nice, France.

^bKW-70, AlmicroTM Instruments, Ambala Cantt., Haryana, India.

^cR-4C, REMI, Mumbai, India.

^dLabofuge 300, Kendro Laboratory Products GmbH, Osterrode, Germany.

Table 2. L-PRF for furcation defects. Papers have been arranged by subapplications (L-RF + OFD versus OFD)

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? , No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
L-PRF + OFD versus OFD						
Sharma & Pradeep (2011a)	RCT Split-mouth Double-blind 9 months	18 – (18) 8 ♀, 10 ♂ Mean age: 34.2 Range: ? Smoking: No	Furcation degree II C: n = 18, OFD T: n = 18, L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF clot 2 L-PRF membrane 10 ml blood/clot	L-PRF + OFD versus OFD SS PD reduction (4.1 versus 2.9 mm), CAL gain (2.3 versus 1.2 mm) and bone fill (50% versus 16.7%) in favour of L-PRF group ($p < 0.001$).
Bajaj et al. (2013)	RCT Parallel Double-blind 9 months	42 – (37) 20 ♀, 22 ♂ Mean age: 39.4 Range: ? Smoking: ?	Furcation degree II C: n = 12 (23), OFD T ₁ : n = 12 (24), L-PRF T ₂ : n = 13 (25), PRP	Hardware: ^a Setting: 400 g/10 min.	1 L-PRF clot 2 L-PRF membrane 10 ml blood/clot	L-PRF + OFD versus OFD SS PD reduction (T ₁ : 4.2 versus T ₂ : 3.9 versus C: 1.5 mm), CAL gain (T ₁ : 2.8 versus T ₂ : 2.7 versus C: 1.3 mm) and bone fill (T ₁ : 44% versus T ₂ : 42% versus C: 2.8%) ($p < 0.001$).

C, control group; CAL, clinical attachment level; OFD, open flap debridement; PD, pocket depth; PRP, platelet-rich plasma; SS, statistically significant; T, test group.
^aR-4C, REMI, Mumbai, India.

statistical significant difference in PD reduction (mean difference: 0.2 mm, $p = 0.2$, CI: -0.08 to 0.4), CAL gain (mean difference: 0.4 mm, $p = 0.09$, CI: -0.06 to 0.8), KTW gain (mean difference: 0.3 mm, $p = 0.1$, CI: -0.06 to 0.6), tissue thickness (mean difference: 0.2 mm, $p = 0.09$, CI: -0.03 to 0.4) and root coverage at 6 months (mean difference: 9.6%, $p = 0.6$, CI: -23.2 to 42.4), although the results showed a trend that L-PRF was superior for all of these variables. However, statistically significant difference could be found for recession depth reduction (mean difference: 0.6 mm, $p < 0.01$, CI: 0.2–1.1), in favour of the for the L-PRF treatment.

For the second comparison (CAF + L-PRF versus CAF + CTG, Fig. 4c) also three articles could be used for a meta-analysis (Jankovic et al. 2012, Eren & Atilla 2014, Tunali et al. 2015). No statistical significant differences could be found for all of the variables: PD reduction (mean difference: 0.2 mm, $p = 0.4$, CI: -0.5 to 0.2), CAL gain (mean difference: 0.2 mm, $p = 0.3$, CI: -0.3 to 0.7), KTW gain (mean difference: 0.3 mm, $p = 0.2$, CI: -0.7 to 0.2) and recession reduction (mean difference: 0.2 mm, $p = 0.2$, CI: -0.4 to 0.1). Root coverage could not be included in this meta-analysis since only one article (Jankovic et al. 2012) fully analysed this variable; Eren & Atilla (2014), and Tunali et al. (2015) did not include the standard deviations.

The adverse events were only registered in some articles within the group of periodontal plastic surgery (Aroca et al. 2009, Jankovic et al. 2010, 2012, Eren & Atilla 2014, Gupta et al. 2015). Each article analysed the adverse events with a different scale, so no meta-analysis could be performed. Five out of the nine articles on periodontal plastic surgery reported on pain, swelling and hypersensitivity. All of them observed less side effects in L-PRF sites.

Discussion

L-PRF has often shown a positive effect when applied during periodontal surgery. Although it has been classified as a platelet concentrate (Dohan et al. 2014a), it can also be considered as a living tissue graft due to its cellular content and its constant

Table 3. L-PRF for periodontal plastic surgery. Papers have been arranged by subapplications (CAF + L-PRF versus CAF, CAF + L-PRF versus CAF + CTG, L-PRF versus EMD).

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? , No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
CAF + L-PRF versus CAF Aroca et al. (2009)	RCT Split-mouth Not blind 6 months	20 – (20) 15 ♀, 5 ♂ Mean age: 31.7 Range: 22–47 Smoking: No or ≤20 cig/day	C: n = 21, CAF T: n = 21, CAF + L-PRF	Hardware: ^a Setting: 3000 r.p.m./10 min.	4? L-PRF membrane Modified CAF 10 ml blood/clot	CAF + L-PRF versus CAF SS more root coverage at 3 months (91.5% versus 80%) and 6 months (88% versus 81%) in favour of control group ($p < 0.01$), NSSD for PD reduction in both groups. more CAL gain (2.6 versus 2.5 mm) and GTH (0.0 versus 0.3 mm) in favour of control group ($p > 0.05$). CAF + L-PRF versus CAF SS more root coverage (100% versus 68%) in favour of L-PRF group ($p < 0.05$). SS more WKG (2.4 versus 2.2 mm) in favour of L-PRF group ($p < 0.05$).
Padma et al. (2013)	RCT Split-mouth Not blind 6 months	15 – (15) Gender: ? Mean age: ? Range: 18–35 Smoking: No	C: n = 15, CAF T: n = 15, CAF + L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF membrane CAF 10 ml blood/clot	CAF + L-PRF versus CAF SS more root coverage (100% versus 68%) in favour of L-PRF group ($p < 0.05$). SS more WKG (2.4 versus 2.2 mm) in favour of L-PRF group ($p < 0.05$).
Gupta et al. (2015)	RCT Parallel Not blind 6 months	26 – (26) 10 ♀, 16 ♂ Mean age: 37 ± 9 Range: ? Smoking: No	C: n = 15, CAF T: n = 15, CAF + L-PRF	Hardware: ^b Setting: 2700 r.p.m./12 min.	1 L-PRF membrane Modified CAF 10 ml blood/clot	CAF + L-PRF versus CAF NSSD for outcomes in both groups for any parameter ($p > 0.05$).
Thamaraiselvan et al. (2015)	RCT Parallel Single-blind 6 months	20 – (20) 2 ♀, 18 ♂ Mean age: ? Range: 21–47 Smoking: No	C: n = 10, CAF T: n = 10, CAF + L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF membrane + surgical site rinsed with L-PRF exudate CAF 10 ml blood/clot	CAF + L-PRF versus CAF NSSD for outcomes in both groups for any parameter ($p > 0.05$).
CAF + L-PRF versus CAF + CTG Jankovic et al. (2012)	RCT Split-mouth Single-blind 6 months	15 – (15) 10 ♀, 5 ♂ Mean age: ? Range: 19–47 Smoking: No	C: n = 15, CAF + CTG T: n = 15, CAF + L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF membrane CAF 10 ml blood/clot	CAF + L-PRF versus CAF + CTG NSSD for PD, CAL and root coverage for L-PRF and CTG group ($p > 0.05$). SS more gain of keratinized tissue width (0.8 versus 1.4 mm) for CTG group ($p < 0.05$). SS enhanced healing in L-PRF group ($p < 0.05$).

Table 3. (continued)

Authors (year)	Study design, Duration	No. of participants baseline (end), gender, age (mean/range), Smoking (? , No, Yes)	Groups C: control T: test	L-PRF preparation	Surgical protocol	Results
Eren & Atila (2014)	RCT Split-mouth Single-blind 6 months	27 – (22) 13 ♀, 9 ♂ Mean age: 34 ± 13 Range 18.5 Smoking: No	C: n = 22, CAF + SCTG T: n = 22, CAF + L-PRF	Hardware: ^c Setting: 400 g/12 min	1 L-PRF membrane CAF 10 ml blood/clot	CAF + L-PRF versus CAF + CTG NSSD for root coverage in L-PRF group (92.7%) and control group (94.2%) ($p > 0.05$). NSSD for complete root coverage in L-PRF group (72.7%) and control group (77.3%) ($p > 0.05$). CAF + L-PRF versus CAF + CTG NSSD for outcomes in both groups for any parameter ($p > 0.05$). CAF + L-PRF versus CAF + CTG Similar outcomes in both groups for any parameter.
Keceli et al. (2015)	RCT Split-mouth Single-blind 6 months	40 – (40) 27 ♀, 13 ♂ Mean age: 40 ± 7 Range: ? Smoking: No	C: n = 20, CAF + CTG T: n = 20, CAF + CTG + L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF membrane CAF 10 ml blood/clot	CAF + L-PRF versus CAF + CTG NSSD for outcomes in both groups for any parameter ($p > 0.05$). CAF + L-PRF versus CAF + CTG Similar outcomes in both groups for any parameter.
Tunali et al. (2015)	RCT Split-mouth Single-blind 12 months	10 – (10) 6 ♀, 4 ♂ Mean age: 34.2 Range: 25–52 Smoking: No	C: n = 10, CAF + CTG T: n = 10, CAF + L-PRF	Hardware: ^a Setting: 2700 r.p.m./12 min.	1 L-PRF membrane CAF 10 ml blood/clot	CAF + L-PRF versus CAF + CTG Similar outcomes in both groups for any parameter.
L-PRF versus EMD Jankovic et al. (2010)	RCT Split-mouth Not blind 12 months	20 – (20) 12 ♀, 8 ♂ Mean age: ? Range: 21–48 Smoking: No	C: n = 20, CAF + EMD T: n = 20, CAF + L-PRF	Hardware: ? Setting: 3000 r.p.m./10 min.	1 L-PRF membrane Modified CAF 10 ml blood/clot	L-PRF versus EMD More complete root coverage (65% versus 60%) in L-PRF group. Similar WKG between groups.

C, control group; CAF, coronally advanced flap; CAL, clinical attachment level; CTG, connective tissue graft; EMD, Emdogain[®]; GTH, gingival thickness; PD, pocket depth; SCTG, subepithelial connective tissue graft; T, test group; WKG, width of keratinized gingiva.

^aEBA 20, Hettich GmbH & Co KG, Tuttingen, Germany.

^bRC-4, REMI, Mumbai, India.

^cNive Laboratory Equipments, NF200, Ankara, Turkey.

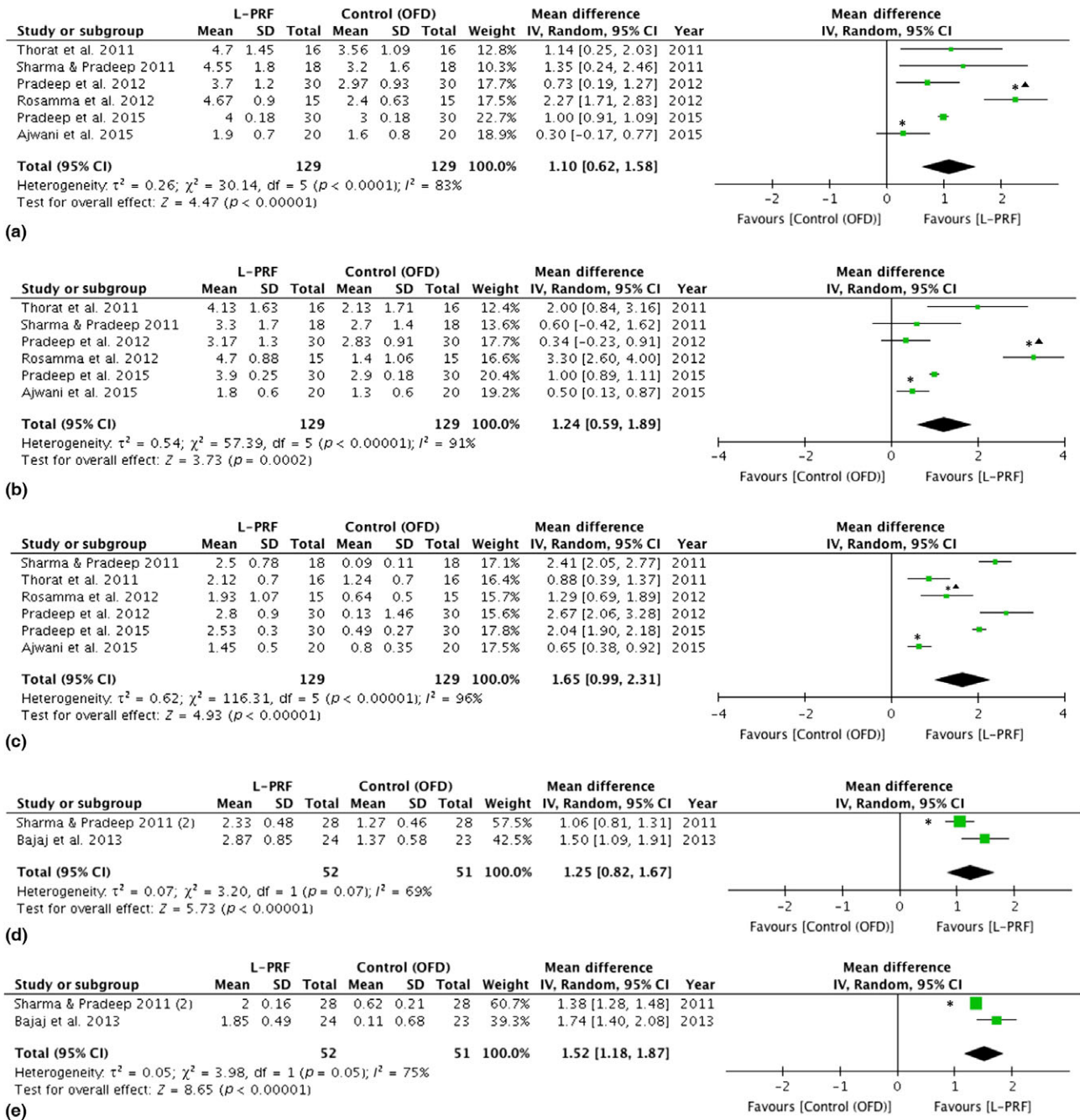


Fig. 3. Forest plot comparing OFD versus OFD + L-PRF in the treatment of intra-bony defects (IBDs) and furcation defects.: ▲ different follow-up from the rest of the studies included. *: study with split-mouth design. (a) Forest plot comparing OFD versus OFD + L-PRF in the treatment of IBDs, PD reduction (mm). (b) Forest plot comparing OFD versus OFD + L-PRF in the treatment of IBDs, CAL gain (mm). (c) Forest plot comparing OFD versus OFD + L-PRF in the treatment of furcation defects, CAL gain (mm). (d) Forest plot comparing OFD versus OFD + L-PRF in the treatment of furcation defects, bone fill (mm). (e) Forest plot comparing OFD versus OFD + L-PRF in the treatment of furcation defects, bone fill (mm). CAL, clinical attachment level; OFD, open flap debridement; PD, Pocket depth.

release of growth factors for more than 7 days (Dohan et al. 2006b).

This review demonstrates that L-PRF has many applications but there is no clear standard protocol per surgical procedure. For example,

the number of clots used varies enormously, as well as the amount of blood drawn to prepare L-PRF. The type of centrifuge and setting also differed from one study to another. More standardized protocols are

necessary in order to better compare and standardize outcomes.

The effectiveness of L-PRF in the treatment of intra-bony defects has been studied by different research groups (Rock 2013, Shah et al. 2014).

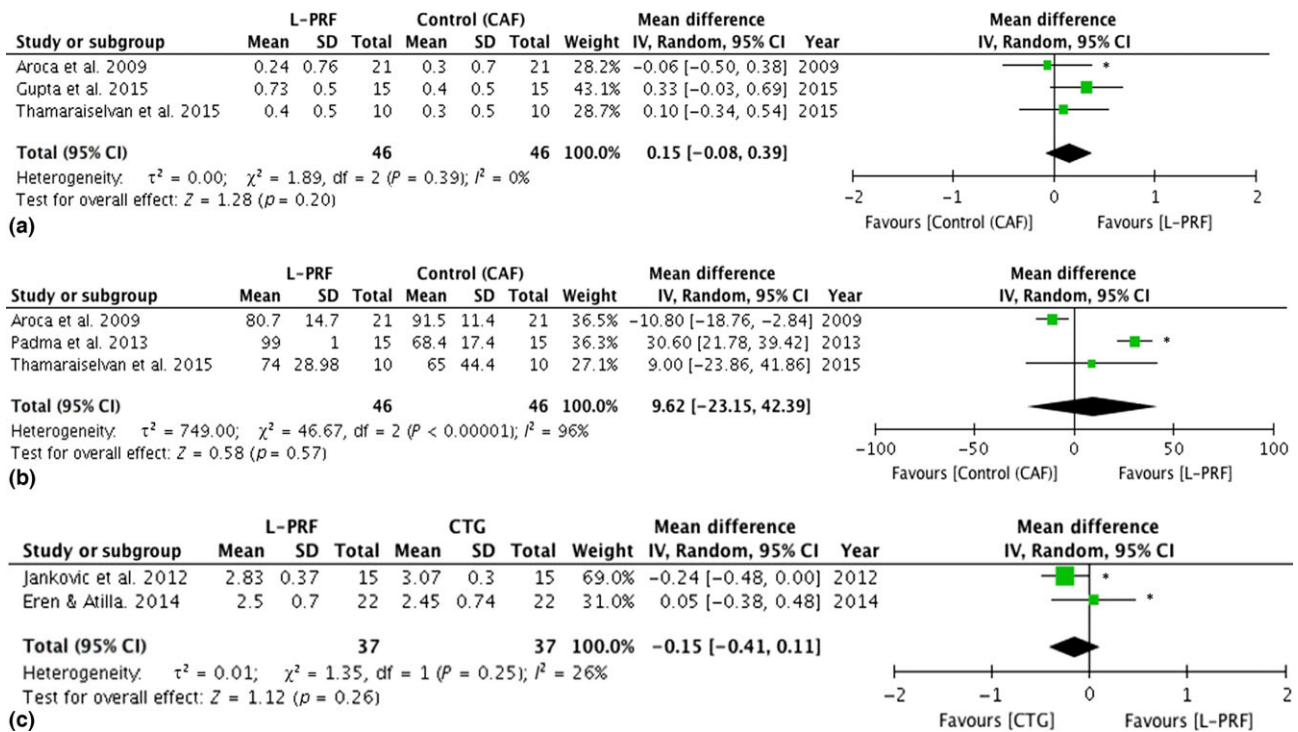


Fig. 4. Forest plot comparing CAF + L-PRF versus CAF and CAF + L-PRF versus CAF + CTG in periodontal plastic surgery. *: study with split-mouth design. (a) Forest plot comparing CAF versus CAF + L-PRF in periodontal plastic surgery, recession reduction (mm). (b) Forest plot comparing CAF versus CAF + L-PRF in periodontal plastic surgery, root coverage 6 months (%). (c) Forest plot comparing CAF + CTG versus CAF + L-PRF in periodontal plastic surgery, recession reduction (mm). CAF, Corionally advanced flap; CTG, connective tissue graft.

In these studies, L-PRF was placed in the defect and L-PRF membranes were used to cover the defect similar to a guided tissue regeneration (GTR) membrane. Clinical and radiographic evaluations showed statistically significant greater PD reduction, CAL gain and radiographic intra-bony defect fill in the L-PRF group (Table 3). Different graft materials were also compared to L-PRF during GTR. The outcomes showed a favourable effect of L-PRF in all clinical parameters measured, or an improvement of the outcomes in studies where L-PRF was combined with other biomaterials (Table 3). Although very limited data exist, the use of L-PRF in furcation defects has also shown favourable results.

For periodontal plastic surgery, the comparison of CAF + L-PRF versus CAF led to controversial results. Although most articles did not show statistically significant differences, L-PRF was superior for all of the parameters recorded. Comparing CAF + L-PRF versus CAF + CTG, L-PRF might be an

alternative to a connective tissue graft. The latter is supported by some case reports (Anilkumar et al. 2009, Agarwal et al. 2013, Singh & Bharti 2013). In this systematic review, a mean root coverage of 86.5% at 6 months has been recorded for CAF + L-PRF treatment. For CAF + EMD and CAF + CTG, a mean root coverage of 91.2% and 90.3% was, respectively, reported in a recent systematic review at 6 months (Cairo et al. 2008).

Some limitations have to be taken into consideration while processing this systematic review. Most of the included articles showed a moderate risk of bias. In those articles, the power analysis was often performed after the recruitment of the participants, where for a RCT it should be done prior to the recruitment in order to determine the sample size. Working in the opposite way, a selective outcome reporting bias can be introduced. Additionally, the allocation concealment and blinding methods were frequently

not applied which increased the risk of bias.

Meta-analysis could be performed in the three indications. However, also here the results of certain studies have to be considered very cautiously. For instance, for the IBDs subgroup, Ajwani et al. (2015) obtained the worst results compared to the rest of the selected articles. The reason could be that two- and three-wall IBDs were included but not analysed separately. Moreover, only one L-PRF clot without membrane was used, so the stability of one L-PRF clot in a two-wall defect without the use of a membrane might not have been ideal. Given the importance of stability in GTR, the use of L-PRF clots in two- or one-wall defects should be accompanied by a L-PRF membrane. In periodontal plastic surgery, Aroca et al. (2009) published the only article that reported better outcome for the control group (CAF). However, smokers (<20 cig/day) were also included, though smoking negatively influences the healing process and affects

complete root coverage (Chambrone et al. 2009, De Sanctis & Clementini 2014). Tobacco smoke might directly affect the peripheral blood cells within the L-PRF (Armilli et al. 2012), yielding to uncertain outcomes.

Regardless the limitations of the included studies, it is worth pointing out some strengths of this systematic review. A total number of 722 participants was enrolled in the selected studies (479 in intra-bony defects, 55 in furcation defects, 188 in periodontal plastic surgery). Taking into consideration the rather short history of L-PRF, this review comprehends a quite large sample of patients. Moreover, the follow-up varied slightly in the articles included for meta-analysis. The duration in the follow-up ranged from 9 to 12 months in the studies selected for quantitative assessment for the IBDs group. Considering furcation defects and periodontal plastic surgery, all of them had a follow-up of 9 and 6 months, respectively. Moreover, only the two most accepted protocols of centrifugation (3000 r.p.m./10 min. or 2,700 r.p.m./12 min.) were included. All other protocols that were not explained in detail or with a non-standardized procedure were excluded. A correct handling of L-PRF is of the utmost importance. It should be clearly distinguished what L-PRF is and what not. For example, L-PRF and PRP contain different cell concentrations, release different amount of growth factors, and have different mechanical properties although both come from a blood sample (Dohan et al. 2006a).

Conclusion

Favourable effects on hard and soft tissue healing and postoperative discomfort reduction were often reported when L-PRF was used. Nevertheless, standardization of the protocol is needed to obtain an optimal effect of L-PRF in regenerative procedures. Correct handling of L-PRF as well as the use of enough clots/membranes per surgical site might be crucial to obtain benefits from this technique. This biomaterial can be taken into consideration due to its reported good biological effects, low costs and ease of preparation.

Acknowledgements

We acknowledge GC Europe N.V. for the chair in bio-regeneration and Intra Lock International Inc. for the chair in optimized osseointegration. The authors also acknowledge Wim Coucke for his support in the statistical analysis and Mrs. Trudy Bekkering from the Belgian Centre for Evidence-Based Medicine (CEBAM).

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Supporting Information

Additional Supporting Information may be found in the online version of this article:

Table S1. Description of platelet concentrates (PCs) characteristics. Although PRP and PRF can be prepared with or without leucocytes (Dohan et al. 2010, Dohan et al. 2014a), this table presents the most common formulations.

Table S2. Search terms used for PUBMED, EMBASE and CENTRAL.

Table S3. Inclusion and exclusion criteria.

Table S4. Excluded articles and reason for exclusion.

Appendix S1. L-PRF preparation. A. Blood is withdrawn from the patient. B. Tubes are centrifuged within 60 s after blood collection without any additives. C. After 12 min. of centrifugation, a clear separation between the platelet-poor plasma, the buffy coat and the red blood cells is obtained. D. L-PRF is presented in the middle of the tube. E. Different L-PRF forms can be produced: liquid, clots or membranes.

Appendix S2. Quality assessment for IBDs. Cochrane tool's for assessment of risk of bias for RCTs.

Appendix S3. Quality assessment for furcation defects. Cochrane tool's for assessment of risk of bias for RCTs.

Appendix S4. Quality assessment for IBDs for periodontal plastic surgery. Cochrane tool's for assessment of risk of bias for RCTs.

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Clinical Relevance

Scientific rationale for the study: The aim of this systematic review and meta-analysis is to extensively analyse the additional regenerative potential of L-PRF during periodontal surgery.

Principal findings: The meta-analysis showed significant clinical benefits of L-PRF for the treatment of IBDs and for furcation defects, and similar outcomes when a connective tissue graft (CTG) was replaced by L-PRF membranes during periodontal plastic surgery.

Practical implications: These results indicate that L-PRF has favourable effects periodontal wound healing, and postoperative discomfort reduction. Nevertheless, standardization of the protocol is needed to obtain an optimal effect.