

1 **Nitrogen availability influences phosphorus removal in microalgae-based wastewater**
2 **treatment**

3 Annelies Beuckels^{*a}, Erik Smolders^b and Koenraad Muylaert^a

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5 ^aKU Leuven Kulak, Laboratory Aquatic Biology, E. Sabbelaan 53, 8500 Kortrijk, Belgium

6 ^bKU Leuven, Department of Soil and Water Management, Kasteelpark Arenberg 20, 3001

7 Heverlee, Belgium

8

9 *Corresponding author: Email: Annelies.Beuckels@kuleuven-kortrijk.be

10 Tel: +32 56 246468

11

12 **Abstract**

13 Microalgae offer a promising technology to remove and re-use the nutrients N and P from
14 wastewater. For effective removal of both N and P, it is important that microalgae can adjust
15 the N and P concentration in their biomass to the N and P supply in the wastewater. The aim
16 of this study was to evaluate to what extent microalgae can adjust the N and P concentrations
17 in their biomass to the N and P supply in the wastewater, and to what extent supply of one
18 nutrient influences the removal of the other nutrient. Using *Chlorella* and *Scenedesmus* as
19 model organisms, we quantified growth and biomass composition in medium with different
20 initial N and P concentrations in all possible combinations. Nutrient supply marginally
21 affected biomass yield of both microalgae but had a strong influence on the composition of
22 the biomass. The nutrient concentrations in the biomass ranged 5.0 – 10.1 % for N and 0.5 –
23 1.3 % for P in *Chlorella* and 2.9 – 8.4 % for N and 0.5 – 1.7 % for P in *Scenedesmus*. The
24 concentrations of P in the biomass remained low and were relatively constant (0.6 – 0.8 % P)
25 when the N concentration in the biomass was low. As a result, removal of P from the
26 wastewater was influenced by the concentration of N in the wastewater. When the initial N
27 concentration in the wastewater was above 40 mg L⁻¹ the microalgae could remove up to 6 mg
28 P L⁻¹, but this removal was only 2 mg P L⁻¹ when the initial N concentration was below 20 mg
29 L⁻¹. A lower N supply increased the carbohydrate concentration to about 40% and lipid
30 concentration to about 30% for both species, compared to around 15% and 10% respectively
31 at high N supply. Our results show that sufficiently high N concentrations are needed to
32 ensure effective P removal from wastewater due to the positive effect of N on the
33 accumulation of P.

34

35 *Highlights*

- 36 • N removal was independent of P supply, P removal improved with increased N supply
- 37 • Biomass N and P concentration was adjusted to the nutrient supply
- 38 • Biomass concentration ranged only a factor 2
- 39 • *Chlorella* has a higher N concentration and lower P concentration compared to
- 40 *Scenedesmus*

41

42 *Keywords:* wastewater treatment, microalgae, nutrient removal, biomass composition

43

44 **1. Introduction**

45 Although the idea to use microalgae for wastewater treatment is not new and dates back
46 almost half a century (Oswald et al., 1957), interest in this concept has revived in recent years.
47 Microalgae-based wastewater treatment has several advantages over conventional wastewater
48 treatment using activated sludge. First, microalgae-based wastewater treatment has a lower
49 energy demand because oxygen is supplied through photosynthesis rather than energy-
50 intensive electromechanical blowers (Green et al., 1995). Second, microalgae not only remove
51 nitrogen (N) and phosphorus (P) from wastewater, but also recycle these nutrients in a
52 biomass that can be converted into energy, raw chemicals or other products (Martins et al.,
53 2010). These advantages are very attractive in times when there is a need to increase energy
54 efficiency and to “close the loop” by recycling elements from waste streams. In this respect,
55 the combination of microalgae wastewater treatment with the production of microalgal
56 biofuels has attracted much attention in recent years (Craggs et al., 2013; Pittman et al.,
57 2011).

58 In conventional wastewater treatment, N and P are removed from wastewater in two separate
59 processes. Usually, N is converted to N_2 gas through coupled nitrification-denitrification
60 while P is precipitated with metal salts. Microalgae, on the contrary, remove N and P from
61 wastewater in a single process. Microalgae absorb N and P from wastewater and use these
62 nutrients to produce biomass. Because microalgae require both N and P to produce new
63 biomass, removal of one nutrient depends on the availability of the other: microalgae cannot
64 remove N without the presence of P in the wastewater, or vice versa, because both nutrients
65 are essential for their growth. The concentrations of N and P in wastewater are quite variable.
66 In domestic wastewater, N concentrations vary between 15 and 90 mg L⁻¹ and P
67 concentrations between 4 and 20 mg L⁻¹ (Abdelaziz et al., 2013; Cai et al., 2013; Christenson
68 and Sims, 2011). Because the sources of N and P in wastewater are different, N and P

69 concentrations often vary independently from each other. Human excreta are a source of both
70 P and N while detergents, soaps and personal care products contain P but little N (Smil, 2000;
71 Tjandraatmadja et al., 2010). Because N and P removal are coupled in microalgae, the fact
72 that N and P concentrations in wastewater vary independent from each other poses a
73 challenge when engineering nutrient removal from wastewater using microalgae.

74 Removal of N and P by microalgae depends on the concentrations of these nutrients in the
75 microalgal biomass. The N and P requirements of microalgae have been an active research
76 field in microalgal ecology and physiology for over half a century. In 1958, Redfield studied
77 nutrient concentrations in marine microalgae and found a ratio of about 106:16:1 C:N:P
78 (molar ratio) in the biomass, a ratio that is since then known as the Redfield ratio (Redfield,
79 1958). It soon became apparent, however, that this ratio is not fixed and that nutrient
80 concentrations in microalgal biomass can be quite variable, particularly in freshwater species
81 (Rhee, 1978). Microalgae have the ability to adjust the N and P concentration of their biomass
82 depending on the supply of nutrients in the medium, resulting in a low nutrient concentration
83 in the biomass when the supply of a nutrient is low and a higher concentration when the
84 supply is high (e.g. Geider and La Roche, 2002; Rhee, 1978; Sterner and Elser, 2002). The
85 microalgal P concentration can range from 0.03% to more than 3% of dry biomass and the
86 microalgal N concentration can range from 3% up to 12% (Reynolds, 2006). Because N and P
87 are used by microalgae to produce various biochemical compounds, changes in the
88 concentrations of N and P in the biomass will influence the biochemical composition of the
89 biomass (Klausmeier et al., 2004; Loladze and Elser, 2011). Nitrogen is predominantly used
90 for synthesis of proteins while P is mainly incorporated into ribosomal RNA. When either N
91 or P are limiting, the protein content of the cell is reduced and cell division slows down but C
92 acquisition through photosynthesis continues. Therefore, cells tend to accumulate C-rich
93 metabolites such as carbohydrates or lipids when the supply of N and P is low (González-

94 Fernández and Ballesteros, 2012; Smith et al., 2010). Induction of N or P limitation is
95 commonly used in microalgal production for biofuels to increase the carbohydrate or lipid
96 yield (Craggs et al., 2013).

97 The flexibility of the N and P concentration of microalgal biomass allows microalgae to
98 adjust their intracellular N and P concentration to the supply of N and P in the wastewater.
99 This flexibility is essential to ensure simultaneous removal of N and P from wastewater.
100 Although there have been a large number of studies on N and P removal from wastewater by
101 microalgae, relatively few studies have specifically studied the flexibility of the N and P
102 concentration in microalgal biomass in response to the supply of N or P in the wastewater.
103 Several studies have shown that a lower N or P supply in wastewater results in a lower N or P
104 concentration of the microalgal biomass and this is often associated with an accumulation of
105 carbohydrates and/or lipids (Akerström et al., 2014; Arbib et al., 2013; Aslan and Kapdan,
106 2006; Dickinson et al., 2013; Samorì et al., 2013; Xin et al., 2010). Other studies have
107 investigated to what extent microalgae can accumulate excess nutrients in their biomass,
108 known as luxury uptake, which is interesting for removal of nutrients from high-strength
109 wastewaters. Some species are capable of accumulating P as polyphosphate granules when the
110 P supply is high and can accumulate up to 3% P in the biomass (Eixler et al., 2006; Powell et
111 al., 2009). Other species of microalgae, such as some diatom species, can accumulate nitrate
112 in their central vacuole (Coppens et al., 2014). In most studies, the concentration of only one
113 nutrient is varied while the other is maintained constant (Arbib et al., 2013; Samorì et al.,
114 2013; Xin et al., 2010; Zhang and Hong, 2014) or the concentration of both nutrients is varied
115 simultaneously without changing their ratio (Akerström et al., 2014; Aslan and Kapdan,
116 2006). No studies have investigated to what extent the supply of one nutrient influences the
117 uptake of the other nutrient.

118 The aim of this study was to evaluate to what extent microalgae can adjust the concentration
119 of N and P in their biomass to the supply of these nutrients in the wastewater. Because
120 nutrient availability has a profound influence on the biochemical composition of microalgae,
121 we expect that availability of one nutrient may have an influence the uptake of the other
122 nutrient. Therefore, we used a central composite design in which the supply of N and P to
123 microalgae was varied independently. We tested this for the two microalgae species that are
124 most frequently observed in microalgae-based wastewater treatment systems: *Chlorella* and
125 *Scenedesmus* (Craggs et al., 2013; Pittman et al., 2011). We also evaluated to what extent
126 changes in wastewater N and P supply influenced the biochemical composition of the biomass
127 because this has important implications for the valorisation of the biomass.

128

129 **2. Material and methods**

130

131 *2.1. Microalgae cultivation*

132 *Chlorella vulgaris* (SAG 211-11 B) and *Scenedesmus obliquus* (CCAP 276/3A) stock
133 cultures were maintained in 2 L glass photobioreactors aerated with 0.2 μm filtered air
134 and mixed by stirring. The medium was based on Wright's Cryptophyte (WC) medium
135 and contained 8.7 mg L⁻¹ K₂HPO₄, 85.0 mg L⁻¹ NaNO₃, 36.8 mg L⁻¹ CaCl₂·2H₂O, 37.0 mg
136 L⁻¹ MgSO₄·7H₂O, 12.6 mg L⁻¹ NaHCO₃, trace metals and vitamins (Guillard and
137 Lorenzen, 1972). The pH of the culture was buffered at 8 by addition of Tris buffer. The
138 cultures were irradiated from one side with daylight fluorescent tubes, giving a light
139 intensity of 60 $\mu\text{E m}^{-2} \text{s}^{-1}$ at the surface of the reactors.

140 *2.2. Experimental setup*

141 To evaluate how the concentration of N influences the removal of P and vice versa,
142 *Chlorella* and *Scenedesmus* were cultured in medium with varying concentrations of N
143 and P. The range of N and P concentrations used here represents the range of
144 concentrations observed in weak to medium strength domestic wastewaters (Abdelaziz et
145 al., 2013). The medium had the same composition as the WC medium (see above) but
146 with modified NaNO₃ and K₂HPO₄ concentrations. A total of 25 treatments were prepared
147 with different concentrations of N (10, 20, 30, 40 and 50 mg N L⁻¹, as NaNO₃) and P (2, 4,
148 6, 8 and 10 mg P L⁻¹, as K₂HPO₄) in all possible combinations. The design of the
149 experiment was based on a central composite design: each combination of N and P
150 concentration was tested once while the centre point (30 mg N L⁻¹ and 6 mg P L⁻¹) was
151 replicated three times to provide information about the within-treatment variability. Tris
152 buffer (4.1 mM) was added to the medium to buffer the pH at 8 and to prevent
153 precipitation of calcium phosphates at elevated pH levels. Preliminary experiments had
154 shown that both species were not capable of using Tris as a N source. The treatments were
155 prepared in 1 L round bottom flasks that were bubbled with 0.2 µm filtered and
156 humidified air to mix the culture and to supply CO₂. For each treatment, 1000 mL of
157 medium was inoculated with 50 mL stock culture to obtain an initial optical density (OD)
158 of 0.05 at 750 nm. The contribution of N and P to the medium through the addition of this
159 inoculum was negligible compared to the N and P concentration already present in the
160 treatments (< 3% N and < 2% P). The flasks were continuously irradiated from one side
161 with daylight fluorescent tubes, giving a light intensity of 120 µE m⁻² s⁻¹ at the surface.
162 The temperature of the culture room was kept constant at 20 ±1 °C. All glassware and
163 media were autoclaved before use and the inoculum was added under a sterile hood to
164 ensure cultures were unialgal. The experiment was terminated after 8 days, when all

165 treatments had reached the onset of the stationary phase. All analyses were performed on
166 samples collected on day 8.

167 *2.3. Measurements and biomass analysis*

168
169 The algal biomass was monitored daily by measuring the absorbance at 750 nm (Griffiths
170 et al., 2011). At the end of the experiment, the dry weight biomass concentration in the
171 medium was determined gravimetrically by filtering a known volume of culture on pre-
172 weighed GF/F filters. The N and P remaining in the medium at the end of the experiment
173 were measured as nitrate and phosphate in the supernatant (centrifugation 10 min, 4500
174 rpm) using a microflow segmented flow analysis system (QuAAtro Seal Analytical,
175 Bran+Luebbe, Germany) following the application notes of the manufacturer. At the end
176 of the experiment, the microalgal biomass was harvested by centrifugation (15 min, 9500
177 g) for analysis of the nutrient, lipid and carbohydrate concentration of the biomass. The
178 pellet was freeze-dried and stored at -80°C until further analysis. The freeze-dried
179 biomass was disrupted by tip sonication and the carbohydrates were extracted in 1M
180 sulphuric acid. The extracted carbohydrates were measured using the Dubois method
181 (DuBois et al., 1956). The fatty acid methyl ester (FAME) concentration was analysed
182 using a direct transesterification method (adapted from Cho et al., 2013) (Trace GC Ultra,
183 Thermo Scientific, Interscience, Louvain-la-Neuve, Belgium). The FAME concentration
184 is a good measure for the potential biodiesel yield of the biomass (Laurens et al., 2012).
185 For analysis of the N and P concentration of the biomass, the biomass was digested using
186 an alkaline persulphate digestion (Koroleff, 1983). The N and P concentration in the
187 digestate was measured as nitrate and phosphate as described above.

188

189 *2.4. Statistical analysis*

190 The results were analysed in R using response surface modelling (RSM) based on a
191 central composite design (Lenth, 2009). The RSM design consisted of a full factorial
192 design for the two variables (N and P), with five levels for each variable and three
193 replications for the centre points. The calculated second order model included the first
194 order terms, the two-way interaction term and the quadratic terms of the variables N and
195 P. The obtained quadratic polynomials for the different response variables were visualised
196 in contour plots.

197

198 **3. Results**

199 *3.1. Nutrient uptake by the microalgae*

200 The initial concentrations of N and P in the medium were measured. These concentrations
201 corresponded well (< 6% deviation) to the theoretical concentrations, of 10 to 50 mg L⁻¹ N
202 and 2 to 10 mg L⁻¹ P, used in the response surface model.

203 Mass balances were calculated to verify whether removal of N and P from the medium
204 was matched by accumulation of N and P in the microalgal biomass (total biomass
205 concentrations times N or P concentration of the biomass). On average, the mass balance
206 was 85% ± 10% for N and 100% ± 22% for P in the *Chlorella* experiment and 92% ± 5%
207 for N and 98% ± 8% for P in the *Scenedesmus* experiment.

208 The N and P concentration of the microalgal biomass varied considerably with the
209 supplied N and P concentrations (Figure 1). In *Chlorella*, the N concentration varied
210 between 5.0 and 10.1 % while the P concentration varied between 0.5 and 1.3 %.
211 *Scenedesmus* contained more P (0.5 to 1.7 %) and less N (2.9 to 8.4 %) than *Chlorella*
212 when compared pairwise for each nutrient combination (paired t-test, p= 0.02 for P and p
213 <0.01 for N) (Figure 2). For both species, the biomass N concentration increased

214 quadratically with the supply of N (Table 1). The N concentration of the biomass was
215 unaffected by the supply of P. As for N, the biomass P increased with increasing P supply
216 in the medium, and this increase was linear in *Scenedesmus* and quadratic in *Chlorella*.
217 The P concentration of the biomass was also influenced by the N supply in the medium. In
218 both species, the P concentration of the biomass was higher when the N supply was high.
219 When the P concentration of the biomass is plotted versus the biomass N concentration, it
220 is clear that the P concentration varies over a wide range when biomass N concentration is
221 high, whereas biomass P concentration is confined to a narrow range when biomass N
222 concentration is low (Figure 2).

223 The nutrient concentrations remaining in the medium after 8 days of cultivation were
224 measured (Figure 3). The residual N concentrations ranged from < 1% to 49% of the
225 initial values while the residual P concentrations ranged from <1% to 80%. The residual N
226 and P concentrations increased quadratically with the N and P supply, indicating that high
227 residual N and P concentrations were limited to the treatments with the highest N or P
228 supply (Table 1). For both species, N and P were removed below the current EU emission
229 standards (10 mg L⁻¹ for N and 2 mg L⁻¹ for P, 91/271/CEE and 98/15/EC Urban
230 Wastewater Treatment Directive, (European Commission Directive, 1998) when the N
231 supply was 40 mg L⁻¹ or less or when the P supply was 4 mg L⁻¹ or less. For both species,
232 P removal was more effective when the N supply was high. At high N supply, P was
233 removed to below the emission standard of 2 mg L⁻¹ up to a P supply of 6 mg L⁻¹ for
234 *Chlorella* and up to a P supply of even 8 mg L⁻¹ by *Scenedesmus*. The removal of N by
235 *Scenedesmus* was not influenced by the P supply in the medium. The N removal by
236 *Chlorella* was slightly reduced when the P supply was low, but this effect was relatively
237 small. The removal of N was only influenced by the P supply at the highest N supply

238 level, on average the N removal was 22% lower at the lowest P supply compared to the
239 highest P supply level.

240 3.2. Microalgal biomass concentration and biomass composition

241 The final dry weight microalgal biomass yield was about 10% higher for *Scenedesmus*
242 than for *Chlorella* when compared pairwise for each nutrient combination (paired t-test,
243 $p= 0.03$). The final microalgal biomass yield varied between 245 and 475 mg L⁻¹ for
244 *Chlorella* and between 304 and 564 mg L⁻¹ for *Scenedesmus*. For *Chlorella*, the final
245 biomass yield decreased in a quadratic way with the N supply in the medium and was
246 reduced by about 40 % at the lowest N supply level (Figure 4, Table 1). The final biomass
247 yield was also significantly reduced at lower P supply in the medium, but only by about
248 20 %. The final biomass yield of *Scenedesmus* was unaffected by the medium
249 composition. This is due to the relatively large measurement error on the dry weight
250 biomass measurement for *Scenedesmus* (standard error is 56 mg L⁻¹, i.e. more than 10% of
251 the mean, Table 1). In contrast to *Chlorella* cells, *Scenedesmus* formed small flocs. This
252 complicated subsampling of the cultures and resulted in higher variability between
253 replicate biomass measurements.

254 The supplied nutrient concentrations in the medium had a strong influence on the
255 biochemical composition of the biomass. The carbohydrate concentration ranged from 18
256 to 40 % for *Chlorella* and from 13 to 44 % for *Scenedesmus*, indicating that both species
257 accumulated starch as a result of nutrient starvation (González-Fernández and Ballesteros,
258 2012; Markou et al., 2012) (Figure 4). For both species, the carbohydrate concentration of
259 the biomass decreased linearly with increasing N supply in the medium (Table 1). In
260 *Chlorella*, the carbohydrate concentration also decreased (about 25%) with increasing P
261 supply (> 2 mg P L⁻¹) in the medium, but the response was weaker when compared to that

262 of N. For both species, the FAME concentration responded in a quadratic way to the N
263 supply in the medium (Table 1), being highest when N supply was lowest. The increase in
264 FAME was mainly due to an increase in C16:0 (palmitic acid) and C18:1 (oleic acid).

265

266 **4. Discussion**

267

268 The aim of this study was to evaluate to what extent microalgae can adjust the N and P
269 concentration in their biomass to the supply of these nutrients in the wastewater and to test
270 whether the supply of one nutrient influences the uptake of the other nutrient. In both
271 *Chlorella* and *Scenedesmus*, the N and P concentration in the biomass responded to the
272 supply of these nutrients, being high when the supply in the medium is high and low when
273 the supply in the medium is low. This is in agreement with previous research (Akerström
274 et al., 2014; Arbib et al., 2013; Dickinson et al., 2013; Halfhide et al., 2014). Interestingly,
275 in both species, the uptake of P not only depended on the supply of P but also on the
276 supply of N. At high N supply, the concentration of P in the biomass was a function of the
277 supply of P. At low N supply, the concentration of P in the biomass was low, irrespective
278 of the supply of P in the medium. Conversely, the N concentration in the biomass was
279 independent of the P supply and was only influenced by the N supply in the medium.

280

281 This observation can be explained by the roles of N and P in the cellular metabolism. In
282 microalgae, N is mainly integrated in proteins and a low N supply will thus limit the
283 synthesis of proteins (Geider and La Roche, 2002; Loladze and Elser, 2011). A reduction
284 in protein synthesis will result in a reduction in the number of ribosomes as well as the
285 amount of ribosomal RNA. Because most P in the cell is stored in the ribosomal RNA, a
286 reduction in ribosomes will result in a lower cellular P concentration (Sternier and Elser,

287 2002). If this is indeed the physiological explanation for the influence of N on P uptake,
288 this phenomenon may not only apply to the model species used in this study, but also to
289 other species of microalgae. Indeed, in experiments with other species of microalgae, it
290 has been noted that a reduction in the N supply not only resulted in a reduction of the N
291 concentration but also of the P concentration in the biomass e.g. in *Phaeodactylum* and
292 *Tetraselmis* (Goiris et al., 2015).

293

294 This study confirmed that the N and P concentration in microalgal biomass can deviate
295 strongly from the concentrations predicted by the Redfield ratio. For *Chlorella*, the N:P
296 ratio in the biomass varied between 15 - 42 and for *Scenedesmus* between 7 - 32. Models
297 for nutrient uptake by microalgae based on a fixed Redfield stoichiometry are therefore
298 not reliable for estimating the capacity of microalgae to remove N and P removal from
299 wastewater. More flexible models that take into account a variable N and P concentration
300 in the biomass should be applied, such as the Droop model (Droop, 1973). In these
301 models, the N concentration in the biomass is only dependent on the N supply in the
302 medium. According to our results, these models may be refined in a way that the N
303 concentration in the biomass not only depends on the N supply in the medium, but also on
304 the P supply.

305

306 The amount of N and P that can be removed from wastewater by microalgae depends on
307 the amount of microalgal biomass that is produced as well as on the N and P concentration
308 in that biomass. In our experiments, variability in biomass concentration and variability in
309 nutrient concentrations in the biomass contributed about equally to the nutrient removal
310 from the medium. The final biomass concentration in the treatments varied by about a
311 factor of two between the different treatments, while the N and P concentration in the

312 biomass varied by a factor of two to three. Because the N and P concentration in the
313 biomass influences the growth rate, there is a link between the N and P concentration in
314 the biomass and the biomass productivity. When the N and P supply is high, microalgae
315 accumulate more N and P in their biomass and less C-rich metabolites, and cell division
316 rates and thus biomass productivity are high. When the N and P supply is reduced,
317 microalgae accumulate more C-rich metabolites and less N and P, and cell division rate
318 and biomass productivity are reduced. The accumulation of C-rich metabolites under low
319 nutrient supply was evident from the accumulation of carbohydrates and lipids in the
320 biomass, particularly in treatments with a low N supply. The carbohydrate concentration
321 of the biomass decreased significantly with the N concentration in both *Chlorella*
322 (Pearson correlation -0.681, $p < 0.001$, $n = 27$) and *Scenedesmus* (Pearson correlation -
323 0.936, $p < 0.001$, $n = 27$). Photosynthetic products are redirected from protein synthesis to
324 carbohydrate synthesis when nutrients become limiting (González-Fernández and
325 Ballesteros, 2012). At the lowest N supply treatments (10 mg L^{-1}), lipids were
326 accumulated: the FAME concentration of the biomass increased from about 12 % to 32 –
327 36 % in *Chlorella* and from about 10 % to 25 – 31 % in *Scenedesmus*. P limitation had no
328 influence on the FAME concentration of both species but had some influence on the
329 carbohydrate concentration in *Chlorella*. It is known that microalgae first accumulate
330 carbohydrates and start accumulating lipids only when nutrient stress is more advanced
331 (Procházková et al., 2014; Zhu et al., 2014). The biomass productivity was also
332 significantly affected by the nutrient supply in *Chlorella*. In *Scenedesmus* no significant
333 effect of nutrient supply on biomass productivity was detected, but this can be ascribed to the
334 high variability in biomass analyses for this species, which were due to aggregate
335 formation.

336

337 The dependence of the biomass P concentration on the N supply in the medium had
338 implications for the removal of P from the medium. The removal of P was limited when
339 the N supply was low. P was removed below the EU emission level standard of 2 mg L⁻¹
340 only when the P supply was 4 mg L⁻¹ or less. When the N supply was high, however, P
341 could also be removed below 2 mg L⁻¹ in treatments with a P supply of 6 or even 8 mg L⁻¹.
342 On the contrary, N removal was independent of the P supply. N was removed to below 10
343 mg L⁻¹ (the EU emission level for N) in all treatments where the N supply was 40 mg L⁻¹
344 and even in some of the treatments with a supply of 50 mg N L⁻¹. Because the model
345 species used here are often dominant in HRAP systems used for microalgae-based
346 wastewater treatment, *Chlorella* and *Scenedesmus*, it is likely that the same phenomenon
347 may apply to large-scale systems. Further research is needed to investigate to what extent
348 this link between N and P accumulation in the biomass is influenced by C assimilation,
349 which is in turn influenced by irradiance and CO₂ supply. An increase in C assimilation
350 will most likely increase the C:N and C:P ratio in the biomass (Sturner and Elser, 2002),
351 but it is not known whether this will influence the balance between N and P in the cell.

352

353 There were differences between the two model species that were used with respect to the
354 N and P concentration in the biomass. *Chlorella* was capable of accumulating more N
355 than *Scenedesmus* (5 to 10 % as opposed to 3 to 8%). *Scenedesmus*, on the contrary,
356 accumulated more P than *Chlorella* (0.5 to 1.7 % as opposed to 0.5 to 1.3 %). This makes
357 *Chlorella* a more attractive candidate for treatment of wastewaters that are rich in N, while
358 *Scenedesmus* may be more suitable for waters rich in P. The flexibility in the N and P
359 concentration of the biomass that was observed for these species in this study fell well
360 within the range of concentrations that have been reported in previous studies (Akerström
361 et al., 2014; Arbib et al., 2013; Dickinson et al., 2013). Differences between this and other

362 studies may be due to differences in the N and P supply, as well as differences in time of
363 exposure to nutrient depletion. When nutrient supply is lower, this may result in lower
364 biomass nutrient concentrations than observed in our study. When microalgae are exposed
365 to nutrient stress for a prolonged time, either in batch or in continuous cultures, the
366 nutrient concentration in the biomass may also be lower than observed in this study e.g.
367 (Dickinson et al., 2013).

368

369 **Conclusion**

370

371 *Chlorella* and *Scenedesmus* are capable of adjusting the N and P concentration in their
372 biomass to the N and P supply in the wastewater, which is important to achieve
373 simultaneous removal of both N and P in wastewaters with a variable N and P loading.
374 The degree to which both species adjusted the P concentration in their biomass, however,
375 depended on the N concentration in the biomass. Microalgae can accumulate more P when
376 N concentrations are high but P uptake is reduced when N concentrations are low. This
377 implies that a sufficiently high N concentration in the wastewater is a prerequisite for
378 effective P removal. *Scenedesmus* can accumulate more P in its biomass compared to
379 *Chlorella*, while *Chlorella* can accumulate more N than *Scenedesmus*. Because
380 microalgae adjust the N and P concentration in the biomass, the biomass yield varies
381 relatively little with differences in wastewater nutrient concentration. The biochemical
382 composition of the biomass, on the contrary, is strongly influenced by the wastewater
383 nutrient concentration, with a two- to four-fold increase in lipids or carbohydrates when
384 the wastewater N concentration is low.

385

386

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391

392 **Tables and figures**

393

	N	P	N²	P²	N*P	Standard error
<i>Chlorella</i>						
Residual N (mg L ⁻¹)	< 0.001	0.011	< 0.001	0.370	0.044	1.50
Residual P (mg L ⁻¹)	< 0.001	< 0.001	0.024	0.001	0.018	0.46
Biomass N (%)	< 0.001	0.186	< 0.001	0.602	0.407	0.23
Biomass P (%)	0.008	0.031	0.351	0.007	0.290	0.08
Dry weight (mg L ⁻¹)	< 0.001	0.022	0.001	0.580	0.271	10.14
Carbohydrate (%)	< 0.001	0.018	0.845	0.132	0.986	1.35
Fame (%)	< 0.001	0.929	< 0.001	0.773	0.452	0.96
<i>Scenedesmus</i>						
Residual N (mg L ⁻¹)	< 0.001	0.216	< 0.001	0.917	0.120	0.06
Residual P (mg L ⁻¹)	0.001	< 0.001	0.613	0.481	0.001	0.41
Biomass N (%)	< 0.001	0.318	< 0.001	0.417	0.467	0.68
Biomass P (%)	< 0.001	< 0.001	0.157	0.612	< 0.001	0.03
Dry weight (mg L ⁻¹)	0.970	0.982	0.287	0.654	0.236	56.05
Carbohydrate (%)	< 0.001	0.254	0.428	0.695	0.322	4.49
Fame (%)	< 0.001	0.142	< 0.001	0.792	0.515	0.30

394

395 **Table 1** Significance (p-value) of the regression parameters N, P, N², P² and N*P in the response
 396 surface models and standard error of the 3 centre point replications (30 mg N L⁻¹ and 6 mg P L⁻¹). The
 397 standard error was calculated as the standard deviation, of the response variable for the replicated
 398 centre points, divided by the square of 3 (number of centre points).

399

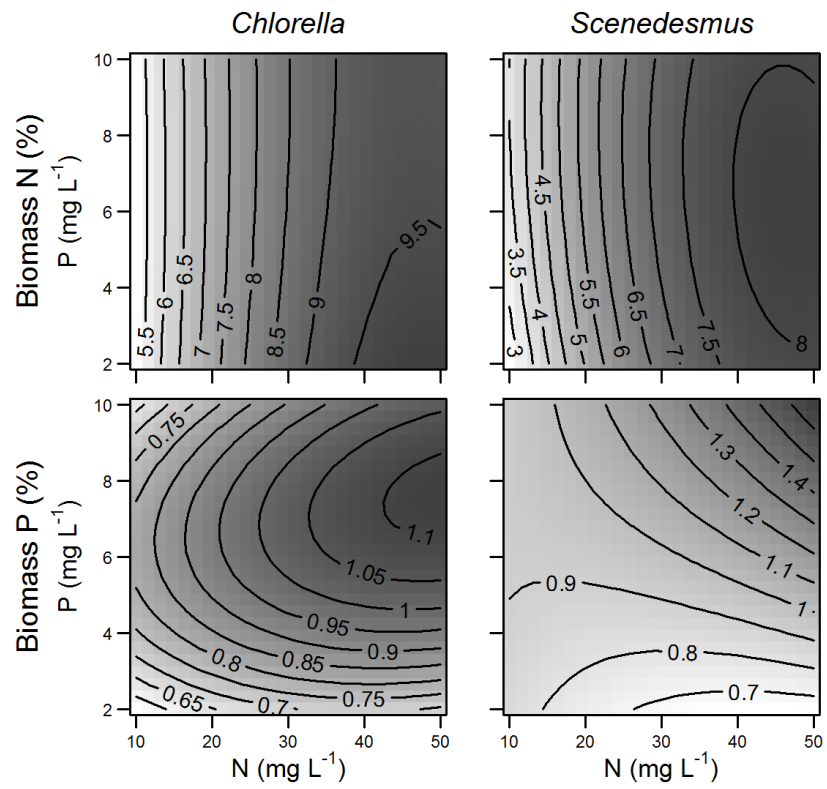


Figure 1. Biomass N and P concentrations (% of dry weight), at day 8 of the batch culture, as a function of the supplied N and P concentrations.

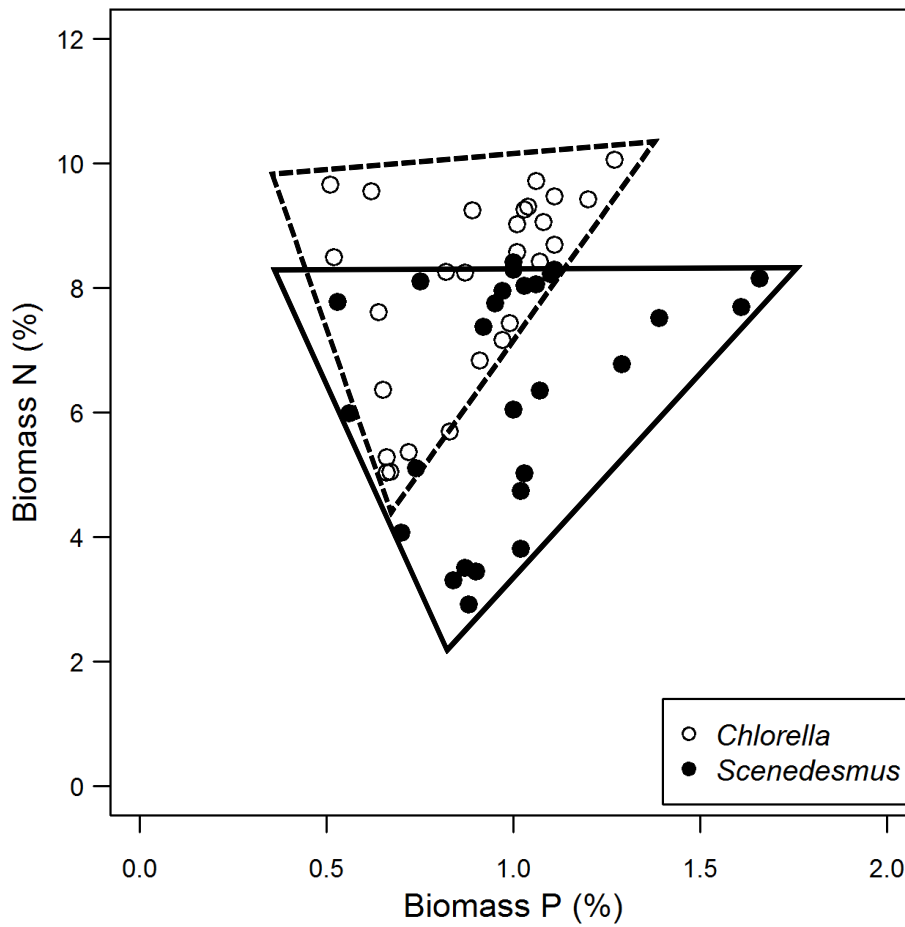


Figure 2: The biomass N concentrations (% of dry weight) as a function of biomass P concentrations (% of dry weight).

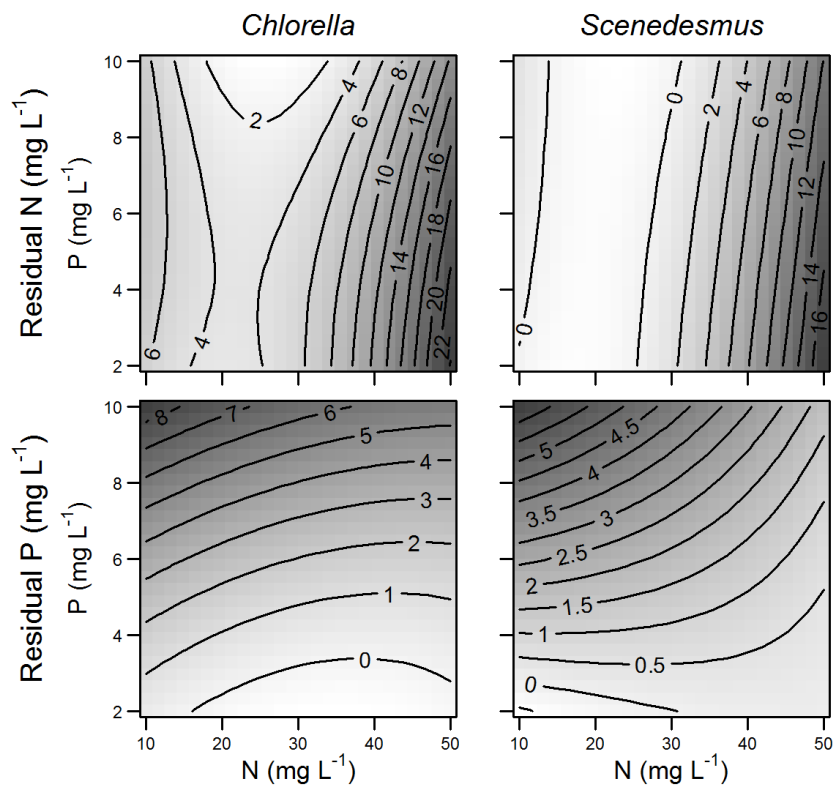


Figure 3. Residual medium N and P concentrations (mg L^{-1}) at day 8 of the batch culture, as a function of the supplied N and P concentrations.

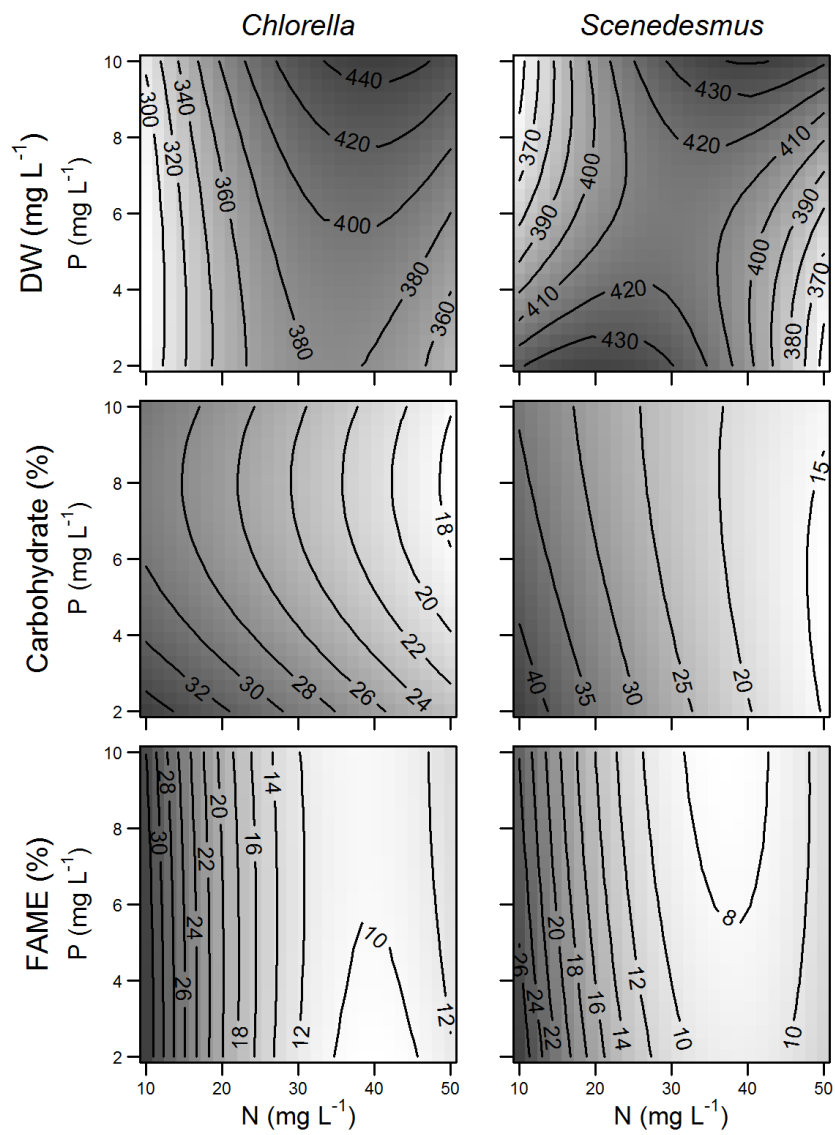


Figure 4. Density measured as dry weight (mg/L) and the biomass carbohydrate and FAME concentrations (% of dry weight), at day 8 of the batch culture, as a function of the supplied N and P concentrations.

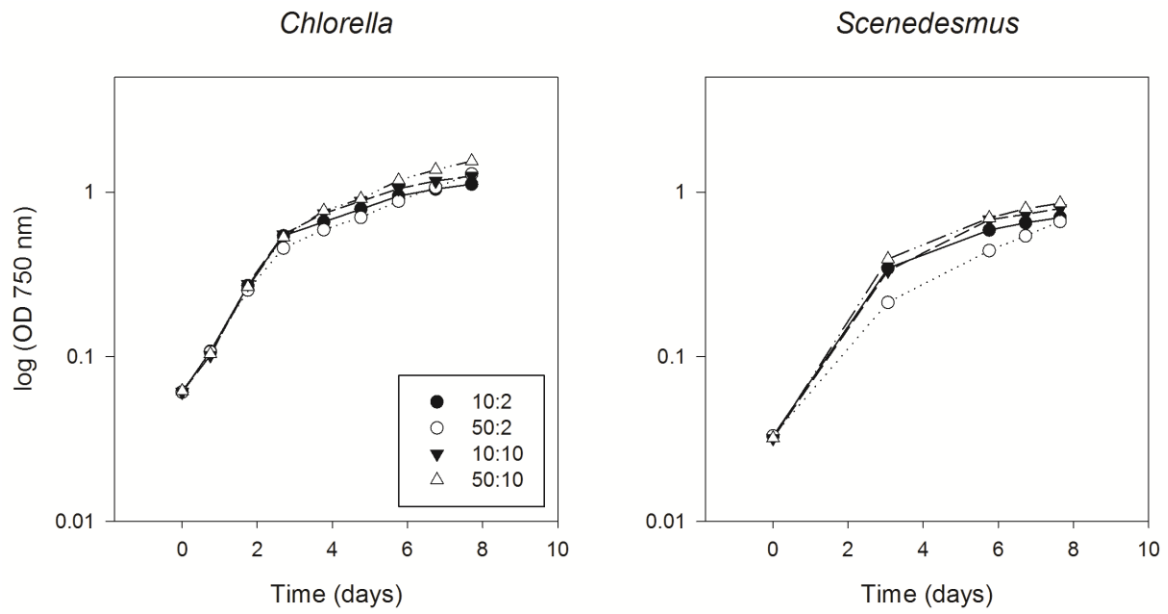


Figure S1. The growth curves represent the logarithmic transformation of the optical density, measured at 750 nm, for *Chlorella* and *Scenedesmus* at a function of time. The treatments with 10 and 50 mg/L N in combination with 2 and 10 mg/L P are shown. The labels show the N:P concentrations eg. label 10:2 is the treatment with 10 mg/L N and 2 mg/L P. After 6 to 8 days the difference between the treatments had stabilised and the algal biomass was harvested at day 8.

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