KU LEUVEN



Citation	García-Pérez, J, Beuckels, A, Vandamme, D, Depraetere, O, Foubert, I, Parra, R, Muylaert K (2014) Influence of magnesium concentration, biomass concentration and pH on flocculation of <i>Chlorella vulgaris</i> Algal Research, vol 3, 24-29.
Archived version	Author manuscript: the content is identical to the content of the published paper, but without the final typesetting by the publisher
Published version	http://www.sciencedirect.com/science/article/pii/S221192641300115X
Journal homepage	http://www.journals.elsevier.com/algal-research
Author contact	your email <u>dries.vandamme@kuleuven-kulak.be</u> your phone number + 32 (0)56 24 6041
IR	https://lirias.kuleuven.be/handle/123456789/425870

(article begins on next page)



Influence of magnesium concentration, biomass concentration and pH on flocculation of Chlorella vulgaris J. Saúl García-Pérez^a, Annelies Beuckels^b, Dries Vandamme^{b1}, Orily Depraetere^b, Imogen Foubert^b, Roberto Parra^a, Koenraad Muylaert^b ^a Water Center for Latin America and Caribbean, Tecnologico de Monterrey, Av. Eugenio Garza Sada 2501 Sur, 64849 Monterrey, Nuevo Leon, Mexico. ^b Laboratory Aquatic Biology, KU Leuven Kulak, E. Sabbelaan 53, Kortrijk, Belgium ¹Corresponding author: Email: dries.vandamme@kuleuven-kulak.be Tel: +32 56 246041 Fax: +32 56 246999

19	Abstract
20	
21	Autoflocculation is a promising low-cost method for harvesting microalgae for bulk
22	biomass production or wastewater treatment. Autoflocculation can be caused by
23	precipitation of calcium or magnesium at high pH. In this study, we investigated the
24	interactive effects of pH, magnesium concentration and microalgal biomass
25	concentration on flocculation of Chlorella vulgaris by magnesium hydroxide. The
26	minimum pH for inducing flocculation was lower when magnesium concentration in
27	the medium is higher. A higher pH and/or higher magnesium concentration are
28	required for flocculation when microalgal biomass concentration is increased. The
29	sludge volume formed during flocculation is highly variable and is influenced mainly
30	by the amount of magnesium hydroxide that precipitates during flocculation. The
31	sludge volume increases with pH and with magnesium concentration in the medium.
32	There is an optimal pH where flocculation efficiency is maximized (>95%) and
33	sludge volume is minimal (1 - 2% of culture volume). Increasing the pH slightly
34	above this optimum results either in an increase in sludge volume and/or a decrease in
35	flocculation efficiency. We propose that autoflocculation by magnesium hydroxide
36	can be more easily controlled by the dosage of base rather than by targeting a specific
37	pH level.
38	
39	
40	<i>Keywords</i> : flocculation - brucite – magnesium hydroxide - microalgae – harvesting –
41	biomass- coagulation
42	
43	
44	

1. Introduction

4	5
4	6

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

Microalgae are a promising new source of biomass for production of food, feed, fuel and bulk chemicals or for treatment of wastewater [1,2]. Although microalgae are already being produced for high-value products, the cost of production should be reduced by at least an order of magnitude to become a commodity crop [3]. A major challenge in realizing large-scale and low-cost production of microalgae is developing a harvesting technology that can process large volumes of water at a minimal cost [4,5].Flocculation holds a lot of potential as a low-cost method for microalgae harvesting [6,7]. An interesting approach for flocculating microalgae is autoflocculation induced by high pH [5]. Autoflocculation at high pH is the result of precipitation of calcium and magnesium salts. Flocculation can be induced by precipitation of calcium phosphates at pH 8-9, but relatively high phosphate concentrations are required for this process to occur [8,9]. In medium with low phosphate concentrations, flocculation can be induced by magnesium hydroxide precipitation, further referred to as magnesium flocculation. Magnesium hydroxide precipitates at a pH above 10.5 and these precipitates are positively charged up to pH 11.5. These positively charged magnesium hydroxide precipitates can interact with the negatively charged surface of microalgal cells and induce flocculation [10,11]. Because background concentrations of magnesium in most waters are high enough for magnesium flocculation to occur, the only cost involved is the cost of the base used to increase pH [12]. Most studies that explored the potential of magnesium flocculation for harvesting microalgae focused on the flocculation efficiency. A high flocculation efficiency indicates that most algal cells in the medium have flocculated and settled. The flocculation efficiency is thus a measure of the proportion of the biomass that can be harvested by the flocculation method. Studies dealing with magnesium flocculation have documented how the flocculation efficiency is influenced by pH or the magnesium concentration in the medium and how the minimum pH required to

achieve a high flocculation efficiency increases with biomass concentration [11,12].

77

Although flocculation efficiency is an important parameter when evaluating the technical feasibility of magnesium flocculation, other parameters are equally important. When flocculation is used for harvesting microalgae, it is part of a two-stage process in which flocculation is used to remove the bulk of the water, while a mechanical method is used for subsequent dewatering [13,14]. To minimize the cost for mechanical dewatering, it is important that the volume of sludge produced by flocculation is as small as possible. Some studies have reported that the sludge volume may be highly variable in the case of magnesium flocculation. In a study on magnesium flocculation of *Phaeodactylum tricornutum*, it was observed that the sludge volume became very large at high pH [15]. Experiments with autoflocculation of *Chlorella vulgaris* also noted an increase in sludge volume in magnesium flocculation at high pH levels [10]. It is not fully clear, however, what causes these differences in sludge volume.

The goal of this study was to understand how both the flocculation efficiency and sludge volume are influenced by pH and by the magnesium concentration in the medium. This information is important to define optimal conditions for magnesium flocculation of microalgae that maximize the flocculation efficiency and minimize the sludge volume. Based on our results, we propose an approach for implementing magnesium flocculation of microalgae that avoids an excessive accumulation of sludge while ensuring a high flocculation efficiency.

2. Material and methods

2.1 Cultivation of Chlorella vulgaris

The strain *Chlorella vulgaris* 211-11b (SAG, Germany) was used as a model species for studying magnesium flocculation. *Chlorella* has previously been used as a model species in autoflocculation studies (e.g. [10,12]). *Chlorella* was cultured in Wright's Cryptophyte (WC) medium in 30 L plexiglass bubble column photobioreactors (20 cm diameter). The reactors were aerated with 0.2 µm filtered air (5 L min⁻¹) and pH was maintained at 8.5 through pH-controlled addition of carbon dioxide to the air flow. The culture was irradiated from two sides with daylight fluorescent tubes,

giving a light intensity of 60 µEinst m⁻² s⁻¹ at the surface of the reactor. Algal biomass was monitored by measuring absorbance at 750 nm [16]. Absorbance measurements were calibrated against dry weight. Dry weight was determined gravimetrically on pre-weighed GF/F glass fiber filters [16]. Experiments were carried out when the culture was in early stationary phase (1 week old) and when biomass concentrations were 0.2-0.3 g L⁻¹, which is a typical biomass concentration encountered in open pond production systems. Initial biomass concentration was typically 0.01-0.02 g L⁻¹.

2.2 Flocculation experiments

To study the influence of magnesium concentration on flocculation efficiency and sludge volume, *Chlorella* cells were harvested by centrifugation and resuspended in fresh medium. Previous experiments had shown that centrifugation and resuspension in fresh medium has no significant influence on flocculation [9]. This fresh medium was identical to the original Wright's Cryptophyte medium but lacked calcium and magnesium. No calcium was added to avoid precipitation of calcium phosphate or calcium carbonate, which could theoretically also cause flocculation. Magnesium was added to the medium in the desired level by addition of magnesium sulphate. The pH of the medium was adjusted by addition of 0.5 M HCl or 0.5 M NaOH to initiate precipitation of magnesium hydroxide and to induce magnesium flocculation. To study the influence of biomass concentration on magnesium flocculation, *Chlorella* cells were resuspended in fresh medium at different concentrations (0.1, 0.2, 0.4, 0.8 or 1.6 g dry weight L⁻¹). All experiments were carried out in triplicate.

The experiments to study the influence of biomass concentration, pH and magnesium concentration on flocculation efficiency were carried out in jars containing 100 mL of broth and mixing was achieved by a magnetic stirrer. Evaluation of sludge volume was performed in 100 mL measuring cylinders. The experiments to study the influence of pH and magnesium concentration on sludge volume were performed using 1 L broth and mixing was done using an overhead mixer. pH was adjusted during 10 min of intensive mixing at 1000 rpm, followed by 20 min of gentle mixing at 250 rpm. The suspensions were then allowed to settle for 30 minutes. Evaluation of the sludge volume was performed in 1 L Imhoff cones. The flocculation efficiency was estimated from changes in the optical density (measured at 750 nm) prior to pH

146 adjustment (OD_i) and after settling (OD_f) . The flocculation efficiency (η_a) , or the 147 percentage of microalgal biomass removed from suspension, was calculated as: $\eta_a = \frac{OD_i - OD_f}{OD_i}.100$ 148 149 150 2.3 Zeta potential measurements 151 152 To confirm that flocculation was caused by charge neutralisation, we measured 153 changes in the zeta potential of the cells during flocculation. Zeta potential of cell 154 suspensions were measured during stepwise increase of medium pH using a Malvern 155 Zetasizer Nano. Zeta potential measurements were carried out in a control medium 156 lacking magnesium and a medium with magnesium (0.75 mM). 157 158 2.4 Measurements of magnesium concentration 159 160 To quantify the amount of magnesium hydroxide precipitates formed in some 161 experiments, we compared concentrations of magnesium in the medium before and 162 after flocculation. Magnesium concentrations in the medium were measured using 163 ICP-OES (Perkin Elmer, Optima 3300 DV). 164 165 3. Results and discussion 166 167 168 3.1 Influence of magnesium concentration and pH on the flocculation efficiency 169 170 We investigated the effects of free magnesium concentration and pH on the 171 flocculation efficiency in medium with a Chlorella biomass concentration of 0.25 g L⁻¹ (Figure 1). At a very low magnesium concentration (addition of 0.01 and 0.03 172 173 mM magnesium sulphate), no flocculation occurred even when pH was increased to 174 12. At a magnesium concentration of 0.1 mM, flocculation occurred only at the 175 highest pH levels tested (11.5 and 12), and even then the flocculation efficiency 176 remained below 90%. At higher magnesium concentrations (0.3 and 1 mM), 177 flocculation occurred already at a pH of 10.5 and the flocculation efficiency was 178 always higher than 90%. Our results show that the minimum pH to induce

179 flocculation decreases when magnesium concentration in the medium increases. Magnesium flocculation is the result of precipitation of magnesium hydroxide. 180 181 Precipitation of magnesium hydroxide is a function of the product of free magnesium 182 and hydroxide concentrations, the latter increasing in proportion to pH. Thus, when 183 magnesium concentration is high, the amount of hydroxide anions required to initiate 184 precipitation is low and flocculation will occur at a low pH. When magnesium 185 concentration is low, higher hydroxide concentrations and thus a higher pH are 186 required for magnesium hydroxide precipitation and flocculation to occur. 187 Magnesium flocculation is assumed to follow a similar mechanism as flocculation by 188 189 metal salts such as alum or ferric chloride, where flocculation is also caused by metal 190 hydroxide precipitates [10,17]. The minimum metal dose for inducing flocculation 191 with metal salts is in the same order of magnitude as observed in this study: 0.3 to 0.7 192 mM for both ferric chloride flocculation [18,19] and aluminum chloride flocculation 193 [20,21]. This is comparable to observations of magnesium flocculation in a previous 194 study, where 0.25-0.6 mM of magnesium was required to flocculate Chlorella and 195 Scenedesmus [22]. In this study as well as in another study of magnesium flocculation 196 of Chlorella [12], the minimum concentration of magnesium was slightly lower than 197 reported in previous studies 0.15 mM. This lower minimum concentration may be due 198 to the fact that our experiments were carried out with microalgal cells resuspended in 199 fresh medium that did not contain dissolved organic matter excreted by microalgae. 200 This excreted algal organic matter is known to interfere with flocculation and to result 201 in an increased flocculant dose [23,24]. 202 203

204	3.2 Zeta potential measurements
205	
206	To understand the underlying mechanism of magnesium flocculation, we carried out
207	measurements of zeta potential of Chlorella cells in medium with and without
208	magnesium. In the absence of magnesium, the zeta potential of Chlorella cells
209	remained negative and constant over a pH range of 9 to 12 (Figure 2). When the
210	medium contained 0.75 mM magnesium, the zeta potential approached 0 in the pH
211	range 10.75 to 11. Above pH 11.5, the zeta potential decreased again.
212	
213	Magnesium hydroxide precipitates have a point of zero charge of about 11.5 [25].
214	Therefore, the surface charge of magnesium hydroxide is positive below pH 11.5. Our
215	measurements of zeta potential suggest that these positively charged precipitates
216	attach to the negatively charged microalgal cells and neutralize the surface charge.
217	The observed decrease in zeta potential above pH 11.5 can be explained by the fact
218	that the surface charge of magnesium hydroxide precipitates changes from positive to
219	negative at this pH. Our observations are in agreement with Wu et al. [11], who also
220	observed an increase in zeta potential at high pH levels during magnesium
221	flocculation. This neutralization of the surface charge may cause flocculation. Most
222	likely, sweeping flocculation also contributes to the flocculation mechanism,
223	especially at high magnesium concentrations and/or high pH levels, when massive
224	amounts of magnesium hydroxide are formed [17].
225	
226	3.3 Influence of biomass concentration on the minimum pH and magnesium
227	concentration required for flocculation
228	
229	If flocculation is primarily due to charge neutralization, then the amount of
230	magnesium hydroxide required to induce flocculation should increase with increasing
231	biomass concentration. We therefore prepared suspensions of Chlorella cells with
232	different biomass concentrations in medium with either a low (0.5 mM) or a high
233	(1.5 mM) magnesium concentration and adjusted the pH to different levels. When
234	biomass concentration was low (0.1 or 0.2 g L ⁻¹), flocculation could be induced at pH
235	of 10.5 or higher, irrespective of the magnesium concentration (Figure 3). When
236	biomass concentration was 0.4 or 0.8 g L ⁻¹ , flocculation occurred at pH 10.5 when
237	magnesium concentration was high while a higher pH was required when magnesium

concentration was low. At the highest biomass concentration of 1.6 g L⁻¹, no complete flocculation could be induced in the medium with low magnesium concentration while flocculation was still possible at pH 11 and 12 in the medium with higher magnesium concentration. These observations indeed suggest that the amount of magnesium hydroxide required for flocculation increases with increasing biomass concentration. In a previous study on magnesium flocculation, Wu et al. [11] noted that the minimum pH to induce flocculation increased when biomass concentrations were higher. Schlesinger et al. [5], on the contrary, reported that flocculation at high pH was almost independent of biomass concentration. These differences may be due to differences in the magnesium concentration in the medium. When magnesium concentration in the medium is high, massive precipitation of magnesium can occur and this causes flocculation by a sweeping mechanism. When sweeping flocculation occurs, the flocculant dose tends to be independent of biomass concentration [19]. When magnesium concentration is low, charge neutralization is most likely more important than sweeping flocculation and the minimum dose of magnesium will increase with increasing microalgal biomass concentration. 3.4 Influence of magnesium concentration and pH on the sludge volume In previous studies on magnesium flocculation it was noted that the sludge volume is highly variable [10,15]. To better understand what causes variation in sludge volume, we investigated the effects of magnesium concentration and pH on the algal sludge volume (Table 1). Experiments were carried out at a biomass concentration of 0.25 g L⁻¹ Chorella biomass. Magnesium concentrations used in this experiment were relatively high (5, 10 and 15 mM) in order to achieve a high flocculation efficiency at all pH levels tested. Indeed, the flocculation efficiency was always 95% or higher, except in pH 10.5 treatments with 5 and 10 mM magnesium, where the flocculation efficiency was close to 90%. From Table 1, it can be observed that the algal sludge volume increased with both increasing pH and increasing magnesium concentration. When magnesium concentration in the medium was 5 or 10 mM, the algal sludge volume was below 30

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272 mL (on a total volume of 1 L) when pH was only raised to 10.5. But the sludge 273 volume increased to between 100 and 300 mL when pH was raised to 11 or 12. When 274 magnesium concentration in the medium was 15 mM, the algal sludge volume varied 275 from 120 mL at pH 10.5 to more than 400 mL when pH was increased to 11 or 12. 276 Reproducibility was only tested at a single combination of pH and magnesium 277 concentration but was relatively good (standard deviation 2.6% of mean). 278 279 We also measured the magnesium concentration that remained in the medium after 280 flocculation (Table 1) to estimate the amount of magnesium hydroxide that had 281 precipitated during flocculation. When pH was raised to 10.5, less than half of the 282 magnesium disappeared from the medium. When pH was raised further to 11, more 283 than 80% of the magnesium had disappeared from solution. When pH was raised to 284 12, almost no magnesium was left in solution. The amount of magnesium that had 285 disappeared from solution during flocculation was strongly correlated with the algal 286 sludge volume (Pearson correlation coefficient 0.95, p < 0.001, n = 9; Figure 4). This 287 clearly demonstrates that the volume of microalgal sludge formed during magnesium 288 flocculation is highly dependent on the quantity of magnesium hydroxide precipitated. 289 To minimize the sludge volume, it is important that the precipitation of magnesium 290 hydroxide does not exceed the minimum dose required to induce flocculation. 291 Flocculation by magnesium hydroxide precipitation is also used in waste.water 292 treatment in the lime softening treatment [26]. In lime softening, it has also been 293 shown that the sludge volume is a function of the amount of magnesium hydroxide 294 precipitate that is formed [27].

295296

3.5 Optimizing flocculation using magnesium hydroxide

297

298

299

300

301

302

303

304

305

From the results presented above, it is clear that overdosing of base should be avoided to minimize the sludge volume, at least when concentrations of magnesium in the medium are high. Therefore, we carried out an experiment to determine the optimal dose for magnesium flocculation, which is the base dose that maximizes flocculation efficiency and minimizes the sludge volume. We carried out two experiments with very different magnesium concentrations, a low (0.15 mM) and one with high (7.5 mM) magnesium concentration, in order to compare magnesium flocculation in soft waters and waters with a very high magnesium concentration (e.g. very hard water or

306 brackish water). In different beakers, we adjusted the pH to different levels ranging 307 from 9.5 to 12.5, with steps of 0.25 pH units. Chlorella biomass concentration was the same in both experiments (0.25 g L⁻¹). The flocculation efficiency, sludge volume and 308 309 sodium hydroxide dose required to adjust the pH were recorded (Figure 5). 310 311 In the experiment with low magnesium concentration, no flocculation occurred up to 312 pH 10.5. At pH 10.75, flocculation efficiency increased abruptly up to 94%. At this 313 pH level, the sludge volume was 1.5 mL (on a total volume of 100 mL), which 314 corresponds to a concentration factor of about 50. A further increase in pH increased 315 the flocculation efficiency only slightly but resulted in about a doubling of the sludge 316 volume. When pH was increased to 12 or higher, the flocculation efficiency and 317 sludge volume both declined again. This decline can probably be ascribed to a 318 reversal of the surface charge of the magnesium hydroxide precipitates at high pH. In 319 this experiment, the optimal pH was thus 10.75. About 2 mM sodium hydroxide was 320 added at this point in the experiment. Because the medium contained only 0.15 mM 321 magnesium, only 0.3 mM of the 2 mM sodium hydroxide added could have been 322 involved in precipitation of magnesium hydroxide, the remaining part being 323 responsible for increasing free hydroxyl ions (causing the increase in pH) or being 324 absorbed by buffers. 325 326 In the experiment with 7.5 mM magnesium in the medium, flocculation efficiency 327 was 66% at pH 10 and increased to 95% at pH 10.25. At pH 10.25, the sludge volume 328 was 1 mL (on a total volume of 100 mL), corresponding to a concentrating factor of 329 about 100. When pH was increased further, the flocculation efficiency remained more 330 or less constant but the sludge volume increased rapidly to a maximum value of 14 331 mL at pH 11.25, corresponding to a concentration factor of only 7. At pH 12, the 332 flocculation efficiency and sludge volume declined slightly, but not as much as in the 333 experiment with the low magnesium concentration. This may be due to the fact that 334 flocculation was caused more by a sweeping mechanism than by charge 335 neutralization. The optimal pH in this experiment was thus 10.25. Only 0.6 mM of 336 sodium hydroxide was added at this point. This implies that maximum 0.3 of the 7.5 337 mM of magnesium had precipitated at pH 10.25. 338

These results suggests that, at the optimum pH, more or less the same amount of magnesium hydroxide had precipitated in both experiments. This is in agreement with our previous observation that a fixed amount of magnesium hydroxide is required to flocculate *Chlorella* cells at a biomass concentration of 0.2 - 0.3 g L⁻¹. The optimum pH in both experiments, however, was very different. At high magnesium concentration in the medium, flocculation occurs at a lower pH. When pH is increased above the optimum, the sludge volume increases and the concentration factor decreases, particularly when the magnesium concentration in the medium is high. This can relatively easily be avoided by limiting the amount of base that is added, as the dosage of base ultimately determines the amount of magnesium hydroxide that is formed. A recent study of magnesium flocculation of the marine microalgae *Dunaliella* also proposed to control flocculation efficiency and sludge volume by dosage of base rather than by targeting a specific pH value [28].

4. Conclusions

From our experiments, it is clear that the optimum pH for magnesium flocculation is dependent on the magnesium concentration in the medium and the microalgal biomass concentration. In media with a magnesium concentration higher than 5 mM, a small increase in pH above the optimum results in massive precipitation of magnesium hydroxide and a large increase in the sludge volume. It is thus a challenge to optimize magnesium flocculation by targeting a specific pH value. In waters with a high magnesium concentration, magnesium acts as a buffer that absorbs hydroxides. As a result, the sludge volume increases only if a large amount of base is added, much more than the amount to achieve the optimum pH for flocculation. Therefore, rather than targeting a specific pH level, the sludge volume can more easily be controlled by controlling the dosage of base.

370	Acknowledgements
371	
372	This project was financially supported by a bilateral project from KU Leuven and
373	Monterrey TEC (KU Leuven DOC BIL11/13), the Agency for Innovation by Science
374	and Technology in Flanders (IWT strategic research grant O. Depraetere and IWT
375	SBO project "Sunlight") and the Research Foundation Flanders (FWO Ph.D.
376	fellowship A. Beuckels). We thank Sina Salim, Prof. Marjan Vermuë and Prof. Rene
377	Wijffels, Bioprocess Engineering, WUR, Wageningen for the usage of the Malvern
378	Zetasizer during our experiments. The Water Center for Latin America and Caribbean
379	and Tecnologico de Monterrey, Campus Monterrey provided support and assistance
380	during this investigation.
381	
382	
383	

384 385	Refe	rences
386 387	[1]	P.T. Pienkos, A. Darzins, The promise and challenges of microalgal-derived biofuels, Biofuels, Bioprod. Bioref. 3 (2009) 431–440.
388	[2]	N. Savage, Algae: The scum solution, Nature. 474 (2011) 5–6.
389 390	[3]	R.H. Wijffels, M.J. Barbosa, An Outlook on Microalgal Biofuels, Science. 329 (2010) 796–799.
391 392 393	[4]	H.C. Greenwell, L.M.L. Laurens, R.J. Shields, R.W. Lovitt, K.J. Flynn, Placing microalgae on the biofuels priority list: a review of the technological challenges., J. Roy. Soc. Interface. 7 (2010) 703–26.
394 395 396 397	[5]	A. Schlesinger, D. Eisenstadt, A. Bar-Gil, H. Carmely, S. Einbinder, J. Gressel, Inexpensive non-toxic flocculation of microalgae contradicts theories; overcoming a major hurdle to bulk algal production., Biotechnol. Adv. 30 (2012) 1023–30.
398 399 400	[6]	N. Uduman, Y. Qi, M.K. Danquah, G.M. Forde, A. Hoadley, Dewatering of microalgal cultures: A major bottleneck to algae-based fuels, J. Ren. Sust. Energy. 2 (2010) 012701.
401 402 403	[7]	L. Christenson, R. Sims, Production and harvesting of microalgae for wastewater treatment, biofuels, and bioproducts., Biotechnol. Adv. 29 (2011) 686–702.
404 405	[8]	A. Sukenik, G. Shelef, Algal autoflocculation-verification and proposed mechanism, Biotechnol. Bioeng. 26 (1984) 142–147.
406 407 408	[9]	A. Beuckels, O. Depraetere, D. Vandamme, I. Foubert, E. Smolders, K. Muylaert, Influence of organic matter on flocculation of Chlorella vulgaris by calcium phosphate precipitation, Biomass and Bioenergy. 54 (2013) 107–114.
409 410	[10]	B.T. Smith, R.H. Davis, Sedimentation of algae flocculated using naturally-available, magnesium-based flocculants, Algal Research. 1 (2012) 32–39.
411 412 413	[11]	Z. Wu, Y. Zhu, W. Huang, C. Zhang, T. Li, Y. Zhang, et al., Evaluation of flocculation induced by pH increase for harvesting microalgae and reuse of flocculated medium., Bioresour. Technol. 110 (2012) 496–502.
414 415 416	[12]	D. Vandamme, I. Foubert, I. Fraeye, B. Meesschaert, K. Muylaert, Flocculation of Chlorella vulgaris induced by high pH: role of magnesium and calcium and practical implications., Bioresour. Technol. 105 (2012) 114–9.
417 418 419	[13]	L.B. Brentner, M.J. Eckelman, J.B. Zimmerman, Combinatorial life cycle assessment to inform process design of industrial production of algal biodiesel., Environ. Sci. Technol. 45 (2011) 7060–7.

 [15] S. Şirin, R. Trobajo, C. Ibanez, J. Salvadó, Harvesting the microalgae Phaeodactylum tricornutum with polyaluminum chloride, aluminium sulphar chitosan and alkalinity-induced flocculation, J. Appl. Phycol. 24 (2011) 1067 1080. [16] M.J. Griffiths, C. Garcin, R.P. van Hille, S.T.L. Harrison, Interference by pigment in the estimation of microalgal biomass concentration by optical density., J. Microbiol. Methods. 85 (2011) 119–23. [17] J. Bratby, Coagulation and flocculation in water and wastewater treatment,
pigment in the estimation of microalgal biomass concentration by optical density., J. Microbiol. Methods. 85 (2011) 119–23.
430 [17] J. Bratby, Coagulation and flocculation in water and wastewater treatment.
431 IWA Publishing, London, UK, 2006.
L. Chen, P. Li, Z. Liu, Q. Jiao, The released polysaccharide of the cyanobacterium Aphanothece halophytica inhibits flocculation of the alga w ferric chloride, J. Appl. Phycol. 21 (2008) 327–331.
N.B. Wyatt, L.M. Gloe, P. V. Brady, J.C. Hewson, A.M. Grillet, M.G. Hankins, et al., Critical conditions for ferric chloride-induced flocculation of freshwater algae., Biotechnol. Bioeng. 109 (2012) 493–501.
438 [20] A.J. Garzon-Sanabria, R.T. Davis, Z.L. Nikolov, Harvesting Nannochloris oculata by inorganic electrolyte flocculation: effect of initial cell density, ior strength, coagulant dosage, and media pH., Bioresour. Technol. 118 (2012) 418–24.
V.M. Rwehumbiza, R. Harrison, L. Thomsen, Alum-induced flocculation of preconcentrated Nannochloropsis salina: Residual aluminium in the biomass FAMEs and its effects on microalgae growth upon media recycling, Chem. Eng. J. 200-202 (2012) 168–175.
Y. Folkman, A.M. Wachs, Removal of algae from stabilization pond effluen by lime treatment., Water Res. 7 (1973) 419–435.
R.K. Henderson, S.A. Parsons, B. Jefferson, The impact of differing cell and algogenic organic matter (AOM) characteristics on the coagulation and flotation of algae., Water Res. 44 (2010) 3617–24.
D. Vandamme, I. Foubert, I. Fraeye, K. Muylaert, Influence of organic matter generated by Chlorella vulgaris on five different modes of flocculation, Bioresour. Technol. 124 (2012) 508–511.
O. Pokrovsky, Experimental study of brucite dissolution and precipitation in aqueous solutions: surface speciation and chemical affinity control, Geochin Cosmochim. Acta. 68 (2004) 31–45.

457 458	[26]	J. Leentvaar, M. Rebhun, Effect of magnesium and calcium precipitation on coagulation-flocculation with lime, Water Res. 16 (1982) 655–662.
459 460 461	[27]	G.M. Ayoub, F. Merhebi, Characteristics and quantities of sludge produced by coagulating wastewater with seawater bittern, lime and caustic, Adv. Environ. Res. 6 (2002) 277–284.
462 463 464	[28]	A. Besson, P. Guiraud, High-pH-induced flocculation-flotation of the hypersaline microalga Dunaliella salina, Bioresour. Technol. 147 (2013) 464–470.
465		
466		

467 468	Tables and Figures captions
469	
470	Table 1. Flocculation efficiency, sludge volume and residual magnesium
471	concentration in the medium as a function of pH and initial magnesium concentration
472	in the medium. Replicate experiments were only carried out for pH 11 and 10 mM
473	magnesium $(n = 3)$.
474	
475	Fig. 1. The flocculation efficiency of Chlorella vulgaris at different pH levels in
476	media containing different magnesium concentrations.
477	
478	Fig. 2. Zeta potential of Chlorella vulgaris cells as a function of pH in medium
479	without magnesium and with magnesium (0.75 mM). Flocculation occurred between
480	pH 11 and 11.5 in the medium with magnesium while no flocculation occurred in the
481	medium without magnesium.
482	
483	Fig. 3. The flocculation efficiency of Chlorella vulgaris as a function of biomass
484	concentration, at pH 10.5, 11 and 12 in medium with low (0.5 mM) and higher (1.5
485	mM) magnesium concentration.
486	
487	Fig. 4. Relation between the sludge volume and the amount of precipitated
488	magnesium in the flocculation experiments shown in Table 1. Magnesium
489	concentrations varied between 5, 10 and 15 mM while pH varied between 10.5, 11
490	and 12.
491	

492	Fig. 5. Variation in flocculation efficiency, sludge volume and dose of sodium
493	hydroxide added as a function of pH in medium with low (0.15 mM) and high (7.5
494	mM) magnesium concentration.
495	

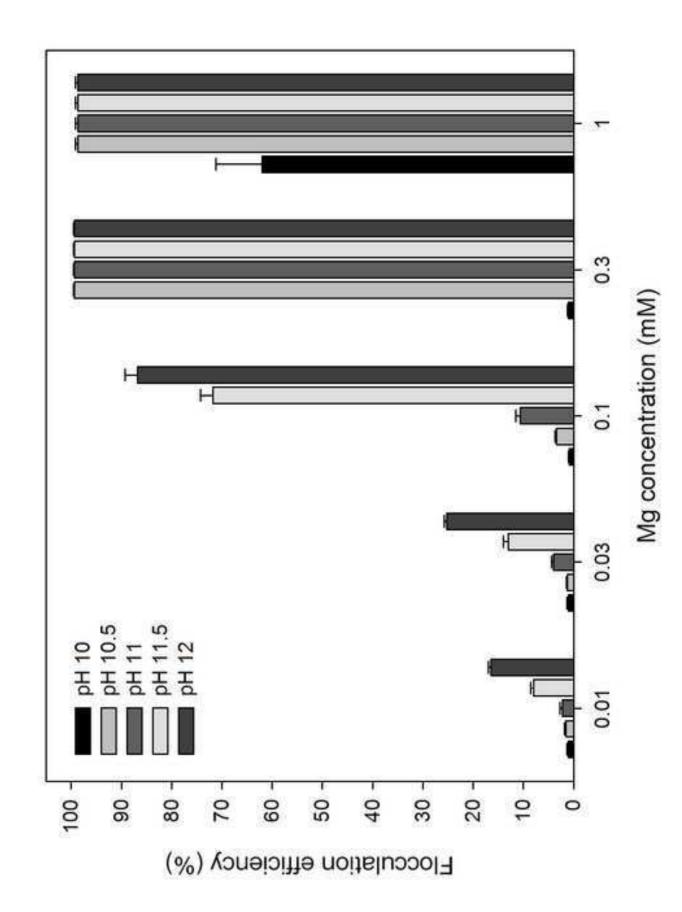
*Acknowledgments

Acknowledgements

This project was financially supported by a bilateral project from KU Leuven and Monterrey TEC (KU Leuven DOC BIL11/13), the Agency for Innovation by Science and Technology in Flanders (IWT strategic research grant O. Depraetere and IWT SBO project "Sunlight") and the Research Foundation Flanders (FWO Ph.D. fellowship A. Beuckels). We thank Sina Salim, Prof. Marjan Vermuë and Prof. Rene Wijffels, Bioprocess Engineering, WUR, Wageningen for the usage of the Malvern Zetasizer during our experiments. The Water Center for Latin America and Caribbean and Tecnologico de Monterrey, Campus Monterrey provided support and assistance during this investigation.

Table 1. Flocculation efficiency, sludge volume and residual magnesium concentration in the medium as a function of pH and initial magnesium concentration in the medium. Replicate experiments were only carried out for pH 11 and 10 mM magnesium (n = 3).

Magnesium	pН	Flocculation	Sludge volume	Residual
		efficiency		magnesium
(mM)		(%)	(mL)	(%)
5	10.5	89.9	20	81
5	11	95.6	127	11
5	12	96.8	192	2
10	10.5	88.9	14	53
10	11	97.4 (±1.1)	283 (±8)	14 (±0.8)
10	12	96.8	286	1
15	10.5	98.2	118	82
15	11	99.4	488	3
15	12	98.7	440	1



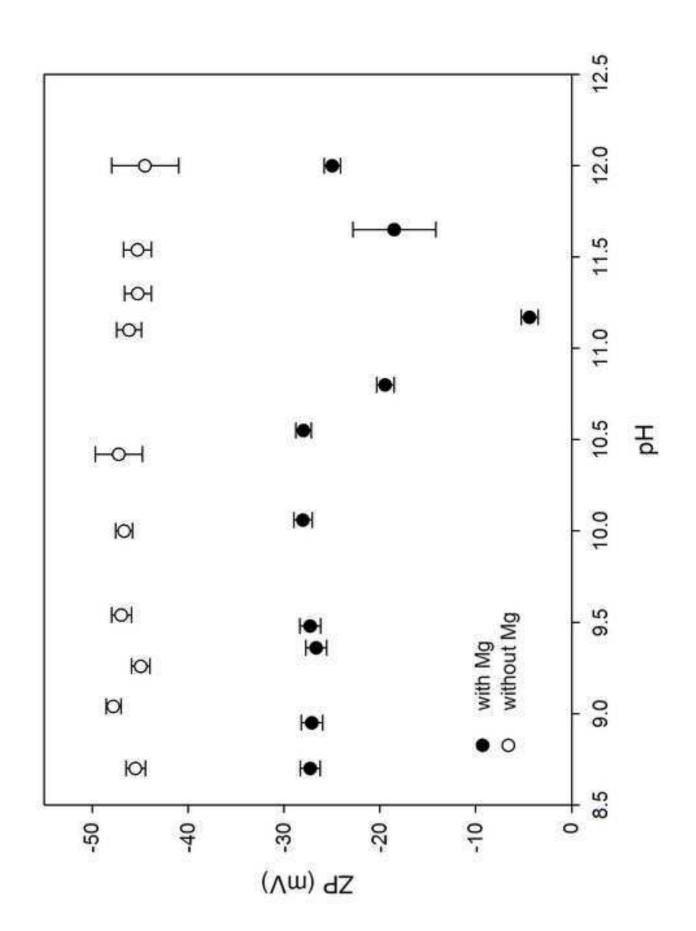


Figure 3 Click here to download high resolution image

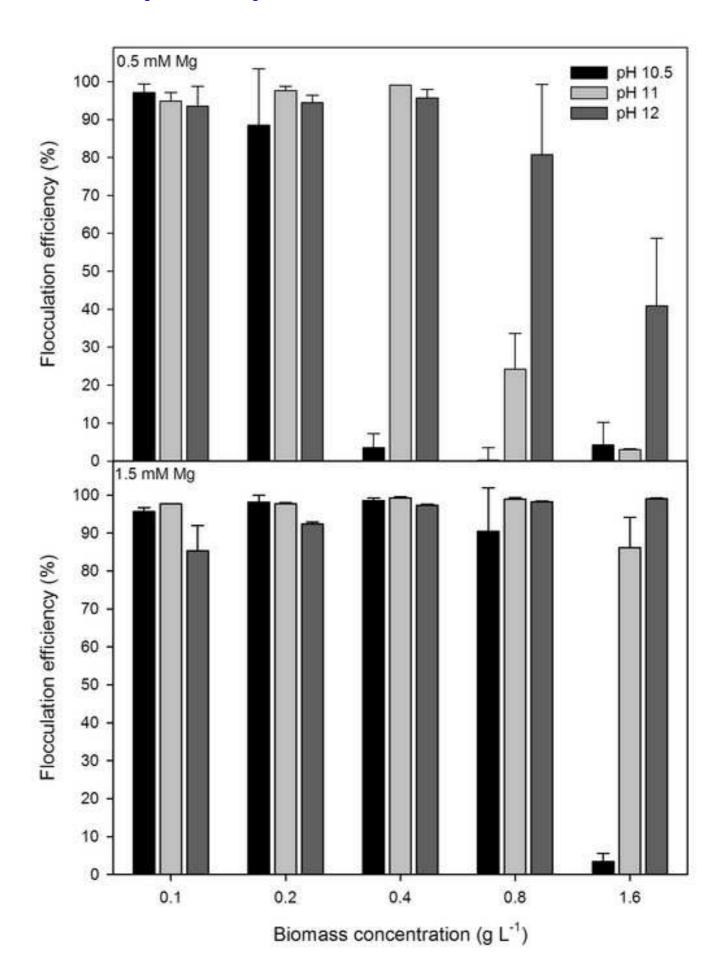


Figure 4 Click here to download high resolution image

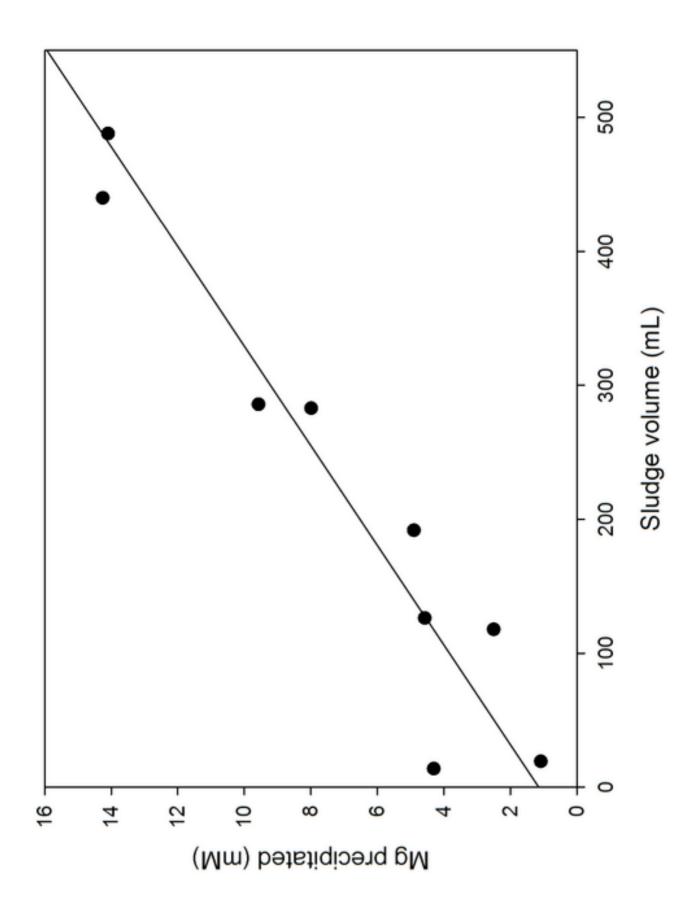


Figure 5 Click here to download high resolution image

