109

110

111

112

113

Higher Drug Exposure During the First 24 Weeks of Ustekinumab Treatment Is Associated With Endoscopic Remission in Crohn's Disease

Q3 ver the past 2 decades, the treatment of Crohn's Odisease (CD) has significantly changed with the introduction of biologicals targeting key inflammatory players. Ustekinumab is a human monoclonal antibody directed against the common p40 subunit of interleukin-12 and interleukin-23, cytokines involved in the immune cascade of chronic immune-mediated inflammatory diseases.¹ As with all biologicals, nonresponse or loss of response to treatment can occur when treating patients with CD with ustekinumab. The UNITI registration trials and subsequent real-world studies reported clinical response rates of around 35%-55% after ustekinumab induction therapy.^{2,3} Inadequate response to treatment can be associated with underexposure to the drug. Several studies have demonstrated the relationship between ustekinumab concentrations and clinical, biologic, or endoscopic response, indicating the potential usefulness of therapeutic drug monitoring to guide clinical decision-making.⁴⁻⁶ Anti-tumor necrosis factor drug concentrations measured during induction are associated with long-term outcomes.^{7,8} For ustekinumab, however, the time point at which the drug concentration is most informative for long-term outcome is unknown. In this study, we evaluated the exposure-response relationship of ustekinumab throughout the first 24 weeks of treatment in patients with CD and investigated the time points at which ustekinumab levels could identify patients achieving endoscopic remission.

1 2

3

4

5

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

A total of 19 patients who started ustekinumab 40 therapy for active CD all received an intravenous infu-41 sion of ~ 6 mg/kg ustekinumab followed by subcutane-42 ous dosing of 90 mg every 8 weeks. The primary 43 outcome was endoscopic remission defined as Simple 44 Endoscopic Score for CD (SES-CD) ≤ 3 after 6 months of 45 therapy. To facilitate multiple sampling, a validated 46 (Supplementary Methods) method of dried blood spot 47 48 (DBS) sampling was applied for the measurement of 49 ustekinumab concentrations. DBS sampling refers to the collection of blood on a protein saver card through a 50 small finger prick, from which the drug can be extracted 51 and measured. Samples were collected at 23 time points 52 over the first 24 weeks of ustekinumab therapy (Week 1, 53 Week 3, Week 4, Week 6, Week 8, Week 8 + 1 day, Week 54 8 + 3 days, Week 8 + 5 days, Week 9, Week 9 + 2 days, 55 Week 10, Week 11, Week 12, Week 16, Week 16 + 1 day, 56 Week 16 + 3 days, Week 16 + 5 days, Week 17, Week 57 17 + 2 days, Week 18, Week 19, Week 20, and Week 24) 58

and ustekinumab concentrations were measured by enzyme-linked immunosorbent assay (CE-labeled Ustekinumab ELISA kit, apDia, Turnhout, Belgium). The local ethics committee approved the study and all patients provided written informed consent (S62619/EudraCTnr: 2019-002038-35).

Spiking various ustekinumab concentrations (0.2–80 μ g/mL) in citrated whole blood and subsequent spotting and extraction showed a consistent ustekinumab recovery and a coefficient of variation <18%. For each spiked concentration, the measured ustekinumab concentration from spotted blood volumes between 15 and 50 μ L was similar. Storing the DBS cards at room temperature for up to 2 weeks and the DBS extracts for up to 2 months at -20° C did not affect the ustekinumab recovery (data not shown).

Of the 19 ustekinumab-treated patients with CD included in this study (Supplementary Table 1), 4 (21%) achieved endoscopic remission. DBS sampling at trough and at various intermediate time points allowed construction of a ustekinumab concentration-time profile, which showed a small peak after each subcutaneous ustekinumab injection that would not be captured when only sampling at trough would be performed (Figure 1A). High variability was observed between the individual concentration-time profiles of the 19 ustekinumabtreated patients with CD. Median concentration-time profiles showed that patients in remission (n = 4) had 97 a significantly higher median area under the curve (AUC) 98 from baseline to Week 24, hence a higher drug exposure, 99 than patients not achieving remission (n = 15) (897 vs 100 479 μ g*day/mL; P < .005; Figure 1B). A similar obser-101 vation could be made for the AUC from baseline to Week 102 8 (517 vs 275 μ g*day/mL; *P* < .01) and from baseline to 103 Week 16 (743 vs 404 μ g*day/mL; *P* < .005) but not for 104 the AUC from baseline to Week 4 (304 vs 209 μ g*day/ 105 mL; P = .0624). Moreover, a negative correlation was 106 observed between the AUC and SES-CD at Week 24 (n =107 19; Spearman r = -0.69; P < .002; data not shown). 108

At multiple time points, the ustekinumab concentration in DBS samples was significantly different between

^aAuthors share co-senior authorship.

 © 2021 by the AGA Institute
 114

 1542-3565/\$36.00
 115

 https://doi.org/10.1016/j.cgh.2021.12.020
 116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

189

190

191

192

193

194

195

196

197

198 199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218



Figure 1. Ustekinumab concentration-time profiles (*A*) and area under the curve (*B*) of patients in endoscopic remission and nonremission. (*A*) Median ustekinumab concentrations (with interquartile range) relative to the number of weeks on ustekinumab therapy in patients in endoscopic remission (*black circles*) compared with patients without endoscopic remission (*white circles*). *Arrows* indicate the time point and dose of administered ustekinumab. (*B*) Median area under the curve (AUC) from baseline to Week 24 of ustekinumab therapy in remitters (n = 4; 897 μ g*day/mL) and nonremitters (n = 15; 479 μ g*day/mL). Mann-Whitney *U* test. ***P* < .005; IV, intravenous; SC, subcutaneous.

patients achieving endoscopic remission and patients not 138 139 achieving remission (Supplementary Table 2). Ustekinu-140 mab concentrations were significantly higher in patients 141 achieving endoscopic remission compared with patients 142 not achieving this outcome, at trough and all evaluated 143 intermediate time points except at Week 1 and, strikingly, 144 also not at the 2 weeks after the subcutaneous dosing at 145 multiple time points (ie, Week 8 + 1 day, Week 9, Week 9 + 2 days, Week 10, Week 16 + 1 day, Week 17, and 146 147 Week 18). When evaluating endoscopic response, defined 148 as a 50% reduction in SES-CD, ustekinumab concentra-149 tions were also significantly different between responders 150 and nonresponders but at less time points than when 151 remission was the evaluated outcome (Supplementary 152 Table 2). A possible explanation for this observation is 153 that the responder's group is a mix of patients with residual inflammation who might relapse and of patients 154 155 that ultimately achieve endoscopic remission.

156 To the best of our knowledge, only 1 study thus far 157 has investigated ustekinumab exposure through the 158 AUC and the relationship with treatment response. In that study by Hanzel et al,9 serum samples from 41 159 ustekinumab-treated patients with CD were collected 160 161 at 1 hour after the intravenous infusion, at Weeks 2, 4, 162 and 8 of treatment. They observed that biochemical, 163 but not endoscopic, remission was associated with 164 ustekinumab exposure over the first 8 weeks of 165 treatment.9

166 The presence of a concentration-response relation-167 ship at trough for ustekinumab in CD is confirmed in our study.^{4–6,9,10} Ustekinumab concentrations at time points 168 between 2 ustekinumab administrations, however, had 169 170 not yet been extensively investigated. Interestingly, 171 ustekinumab levels were not significantly higher in 172 endoscopic remitters compared with nonremitters at 173 time points immediately following subcutaneous dosing. 174 This could suggest that not the absorption but rather the

clearance of ustekinumab may be different between the remitters and nonremitters. Alternatively, significance might not be reached because of the small sample size. Overall, these findings indicate that the first 2 weeks after the subcutaneous ustekinumab injections are not optimal time points to monitor ustekinumab concentrations. Because the concentration-response relationship was observed at multiple time points and no single time point in particular is standing out, monitoring ustekinumab levels can be useful at various time points. Feasibility should also be taken into account when deciding on the time point to monitor ustekinumab concentrations. Larger prospective trials are needed to identify the ustekinumab concentration threshold associated with endoscopic remission at 1 or more of the time points identified in this study.

In conclusion, patients with CD achieving endoscopic remission at Week 24 of ustekinumab therapy have a higher ustekinumab drug exposure than patients not achieving endoscopic remission. Monitoring ustekinumab concentrations at trough or at intermediate time points could help to timely identify patients achieving endoscopic (non)-remission. Q10

	219
NATHALIE VAN DEN BERGHE Q4	220
Laboratory for Therapeutic and Diagnostic	220
Laboratory for morapouto and Diagnootio	221
Antibodies	222
Department of Pharmaceutical and Pharmacological	222
Sciences	223
KILLeuven	224
	225
Leuven, Belgium	226
BRAM VERSTOCKT	227
SÉVERINE VERMEIRE	228
Department of Gastroenterology and Hepatology	229
University Hospitals Leuven	230
Translational Research in Gastrointestinal Disorders	231

Translational Research in Gastrointestinal Disorders 231 Department of Chronic Diseases and Metabolism 232

ARTICLE IN PRESS

KU Leuven 8. Leuven, Belgium 9. 10. Laboratory for Therapeutic and Diagnostic Antibodies Department of Pharmaceutical and Pharmacological Sciences KU Leuven Leuven, Belgium Addre Sciences KU Leuven

Supplementary Material

Note: To access the supplementary material accompanying this article, visit the online version of *Clinical Gastroenterology and Hepatology* at www.cghjournal.org, and at https://doi.org/10.1016/j.cgh.2012.11.00.

References

- 1. Benson JM, et al. mAbs 2011;3:535–545.
- 2. Feagan BG, et al. N Engl J Med 2016;375:1946–1960.
- 3. Macaluso FS, et al. Expert Opin Biol Ther 2020;20:193-203.
- 4. Verstockt B, et al. J Crohns Colitis January 2019;13:864-872.
- 5. Battat R, et al. Clin Gastroenterol Hepatol 2017;15:1427-1434.
- 6. Soufflet N, et al. Clin Gastroenterol Hepatol 2019;17:2610-2612.
- 7. Singh N, et al. Inflamm Bowel Dis 2014;20:1708-1713.

- 8. Verstockt B, et al. Aliment Pharmacol Ther August 2018; 48:731–739.
- 9. Hanžel J, et al. Clin Gastroenterol Hepatol 2021;19:111-118.
- 10. Adedokun OJ, et al. Gastroenterology 2018;154:1660-1671.

Reprint requests

Address requests for reprints to: Debby Thomas, Department of Pharmaceutical and Pharmacological Sciences, KU Leuven, Campus Gasthuisberg O&N2, PB 820, Herestraat 49, B-3000 Leuven, Belgium. e-mail: debby.thomas@ Q5 kuleuven.be.

Acknowledgments

Griet Compernolle and Sophie Tops provided excellent technical assistance with the processing of the DBS samples.

Conflicts of interest

These authors disclose the following: Bram Verstockt reports financial support for research from Pfizer; lecture fees from AbbVie, Biogen, Chiesi, Falk, Ferring, Galapagos, Janssen, MondayNightIBD, MSD, Pfizer, R-Biopharm, Takeda, and Truvion; and consultancy fees from Applied Strategic, Bristol Myers Squibb, Guidepont, Janssen, and Sandoz. Séverine Vermeire reports financial support for research from AbbVie, MSD, Pfizer, J&J, and Takeda; received speaker fees from AbbVie, MSD, Takeda, Ferring, Dr. Falk Pharma, Hospira, Pfizer Inc, and Tillots; and served as a consultant for AbbVie, MSD, Takeda, Ferring, Genentech/Roche, Robarts Clinical Trials, Gilead, Celgene, Prometheus, Avaxia, Prodigest, Shire, Pfizer Inc, Galapagos, Mundipharma, Hospira, Celgene, Second Genome, and Janssen. KU Leuven holds a license agreement with R-Biopharm and apDia. The remaining authors disclose no conflicts.

Funding

Clinical Investigator of the Research Foundation Flanders, Belgium. Bram Verstock is supported by The Leona M. and Harry B. Helmsley Charitable Trust (grant number 2019PG-CD026) and in part by the TBM Grant T003716N of the Research Foundation—Flanders (FWO). Nathalie Van den Berghe is a Strategic Basic Research PhD fellow at FWO. Séverine Vermeire is a Senior Clinical Investigator of the Research Foundation Flanders, Belgium. Bram Verstockt is supported by a Clinical Research Fund (KOOR) from the University Hospitals Leuven, Belgium.

3.e1 Van den Berghe et al

Clinical Gastroenterology and Hepatology Vol. ■, No. ■

Supplementary Methods

For the analytical validation of the DBS method, ustekinumab concentrations (0.2–80 μ g/mL) were spiked in citrated whole blood of healthy donors (Valley Biomedical, Winchester, VA) and spotted (40 μ L) onto a filter paper (Whatman 903 Protein Saver Card, GEHealthcare, Diegem, Belgium). After air-drying the card overnight, a 6-mm diameter disc completely impregnated with blood was punched out. The drug was extracted from the disc by addition of 240 µL Super-BlockT20 buffer (Thermo Fisher Scientific, Rockford, IL) and shaking at 21°C for 1 hour. Ustekinumab concen-trations were subsequently measured using an in-house enzyme-linked immunosorbent assay with antibody MA- UST56A2D11 for capture and MA-UST56C1H12 for detection.

The impact of the spotted volume of blood was investigated by spiking ustekinumab (0.2–80 μ g/mL) to citrated whole blood and spotting different volumes $(15-50 \ \mu L)$ onto filter paper. Subsequently, ustekinumab was measured as previously described.

The stability of ustekinumab in DBS cards kept at room temperature and in DBS extracts at -20° C was investigated by spotting different concentrations of ustekinumab (0.2–80 μ g/mL) on different filter papers. The ustekinumab recovery after storing the DBS card at room temperature for 1 week, 2 weeks, and 4 weeks (relative to overnight drying) or the DBS extracts at -20° C for 1 or 2 months (relative to before freezing), was evaluated.

Supplementary Table	1. Baseline Cha	aracteristics of the	e 19 Included	l Crohn's Disease	Patients
---------------------	-----------------	----------------------	---------------	-------------------	----------

	Remitters	Nonremitters
Number of patients, n (%)	4 (21)	15 (79)
Sex, women, n (%)	3 (75)	6 (40)
Age, median (IQR), y	40.9 (33.5–53.2)	40.8 (32.4–51.3
Disease duration, median (IQR), y	16.3 (12.7–25.6)	16.6 (7.4–26.7)
Simple Endoscopic Score for Crohn's disease, median (IQR)	8.0 (6.5–10.0)	12.0 (7.5–15.0)
Body mass index, <i>kg/m</i> ² , median (IQR)	23.9 (22.6–24.7)	23.3 (22.6–25.8
Previous biologic therapy, n (%) Anti-TNF Vedolizumab	3 (75) 1 (25)	12 (80) 5 (33)
Concomitant steroids, n (%)	0	3 (20)
Disease location, n (%) Ileal disease (L1) Colonic disease (L2) Ileocolonic disease (L3) Upper Gl involvement (L4)	1 (25) 1 (25) 2 (50) 0	5 (33) 1 (7) 9 (60) 2 (13)
Disease behavior, n (%) Inflammatory (B1) Stricturing (B2) Penetrating (B3) Perianal disease (p)	2 (50) 1 (25) 1 (25) 1 (25)	6 (40) 6 (40) 3 (20) 4 (27)
Smoking status, n (%) Active smoking Previously smoking Never smoked	0 0 4 (100)	3 (20) 2 (13) 10 (67)

SSU 5.6.0 DTD ■ YJCGH58234 proof ■ 6 January 2022 ■ 11:49 pm ■ ce CJ

ARTICLE IN PRESS

Supplementary Table 2. Ustekinumab Concentration-Response Analyses

	UST concentration (µg	ration (µg/mL)	nL)		UST concentration (µg/mL)			
Time point	Nonremission ($n = 15$)	Remission $(n = 4)$	P value ^a	AUROC	Nonresponse (n = 12)	$\begin{array}{l} \text{Response} \\ \text{(n}=7) \end{array}$	P value ^a	e ^a AUROC
Week 1	11.5 (9.4–12.6)	15.1 (12.5–17.6)	NS	0.78	11.5 (9.7–12.6)	13.1 (11.3–15.7)	NS	0.70
Week 3	5.7 (4.0–7.8) ^b	9.1 (8.0–10.6)	< .05	0.84	6.0 (4.1–8.2)	7.3 (5.8–9.3) ^b	NS	0.67
Week 4	6.4 (4.7–7.6) ^b	8.9 (8.6–9.5)	< .02	0.89	6.9 (5.7–8.0) ^b	8.5 (4.8–8.9)	NS	0.59
Week 6	2.6 (1.6–3.3)	6.2 (5.6–6.9)	< .005	0.95	2.8 (2.1–3.6)	3.9 (1.7–6.2)	NS	0.62
Week 8 ^c	2.2 (1.2–2.9)	4.4 (3.4–5.3)	< .05	0.85	2.3 (1.3–3.3)	2.2 (2.1–4.4)	NS	0.63
Week 8 $+$ 1 d	2.4 (2.1–3.9) ^d	4.4 (4.2–4.6)	NS	0.82	2.4 (1.9–3.5) ^e	4.3 (4.2–4.5) ^e	NS	0.82
Week 8 $+$ 3 d	2.8 (2.3–3.8) ^f	4.4 (4.2–5.3)	< .05	0.85	2.7 (2.2–3.4) ^b	4.5 (4.3–6.7) ^e	< .01	0.93
Week 8 $+$ 5 d	3.5 (2.1–4.0) ^e	5.9 (5.3–6.6)	< .005	0.96	3.5 (1.9–4.0) ^b	5.0 (4.0–6.2) ^b	< .05	0.83
Week 9	3.3 (2.3–3.9)	4.7 (3.6–5.4)	NS	0.72	3.0 (2.1–4.2)	3.7 (3.0–4.7)	NS	0.64
Week 9 $+$ 2 d	2.9 (2.5–4.3) ^d	5.3 (4.1–6.0)	NS	0.73	3.2 (2.3–4.4) ^e	4.9 (3.0–5.8) ^e	NS	0.68
Week 10	2.8 (2.2–3.4)	3.7 (3.4–4.0)	NS	0.78	2.6 (2.0–3.4)	3.5 (3.1–3.7)	NS	0.75
Week 11	2.3 (1.8–3.2)	4.6 (4.0–5.2)	< .001	1.00	2.1 (1.7–3.2)	3.8 (3.2–4.6)	< .01	0.86
Week 12	1.7 (1.5–2.4)	3.8 (3.8–4.0)	< .001	1.00	1.6 (1.4–2.4)	3.8 (2.2–3.8)	< .02	0.85
Week 16 ^c	1.1 (0.6–1.2)	1.7 (1.4–2.4)	< .02	0.88	1.1 (0.6–1.3)	1.3 (1.0–1.7)	NS	0.70
Week 16 $+$ 1 d	1.9 (1.5–2.4)	2.4 (2.3–2.8)	NS	0.77	1.8 (1.2–2.2)	2.4 (2.3–2.7)	< .01	0.87
Week 16 $+$ 3 d	2.5 (2.3–2.7)	3.4 (3.1–4.0)	< .02	0.88	2.4 (2.0–2.5)	3.4 (3.1–3.7)	< .001	0.94
Week 16 $+$ 5 d	2.9 (2.2–3.1) ^b	4.1 (3.9–4.7)	< .01	0.93	2.9 (1.9–3.0) ^b	4.1 (3.3–4.4)	< .01	0.88
Week 17	3.4 (2.0–3.8)	3.9 (3.5–4.6)	NS	0.77	2.9 (1.6–3.8)	3.5 (3.5–4.0)	NS	0.71
Week 17 + 2 d	3.0 (1.9–3.3) ^e	4.1 (4.0–4.7)	< .005	0.96	3.1 (1.7–3.4) ^b	3.9 (3.4–4.2) ^b	< .05	0.82
Week 18	2.4 (1.7–2.9)	3.2 (2.8–4.2)	NS	0.82	2.3 (1.4–2.6)	3.0 (2.7–3.3)	< .05	0.79
Week 19	1.7 (1.5–2.4)	2.9 (2.6–3.5)	< .02	0.88	1.7 (1.4–2.3)	2.7 (2.0–2.9)	NS	0.76
Week 20	1.5 (1.1–1.7)	2.7 (2.3–3.7) ^b	< .01	0.96	1.4 (0.9–1.7)	2.3 (1.8–2.6) ^b	< .01	0.88
Week 24 ^c	0.6 (0.5–0.9) ^d	1.8 (1.5–2.1)	< .02	0.93	0.6 (0.5–0.7) ^d	1.1 (1.0–1.8)	< .01	0.89

NOTE. No correction for multiple testing was performed.

AUROC, area under the receiving operating characteristics curve; DBS, dried blood spot; NS, not significant; UST, ustekinumab.

^aMann-Whitney U test for comparing ustekinumab concentrations of remitters versus nonremitters and responders versus nonresponders. Ustekinumab concentrations are represented as median (interquartile range).

^b1 datapoint missing.

^cTrough.

^d4 datapoints missing.

^e2 datapoints missing. ^f3 datapoints missing.