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Digital surface scanning in flap perfusion

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Abstract. Monitoring vascular perfusion of transferred tissue is essential in reconstructive surgery to recognize early flap failure. The aim of this study was to evaluate the ability of a digital surface scanner to detect vascular perfusion disorders through the monitoring of skin colour changes. A total of 160 surface scans of the forearm skin were performed with a TRIOS 3D scanner. Vascular compromise was simulated at different time-points by intermittent occlusion of the blood supply to the forearm skin (first the arterial blood supply and then the venous blood supply). Skin colour changes were examined according to the hue, saturation, and value colour scale. Colour differences were analysed with a paired t-test. Significant differences were observed between the colour of the normal skin and that of the vascular compromised skin ($P < 0.01$). The surface scanner could distinguish between arterial occlusion and venous congestion ($P < 0.01$). A digital surface scan is an objective, non-invasive tool to detect early vascular perfusion disorders of the skin.

Research Paper Reconstructive Surgery

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Key words: reconstructive surgical procedures; free tissue flap; three-dimensional image; perfusion; ischemia; venous thrombosis; colour.

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Microvascular free flaps are used for the repair of various tissue defects. Flap failure occurs predominantly in the first 24 hours postoperative, due to vascular compromise of the transferred tissue¹[.](#page-4-0) Causes of vascular compromise include both arterial pathologies, such as arterial thrombosis, vasospasm, and anastomotic failure, and venous pathologies, such as extrinsic venous compression and venous thrombosis¹. An [assessment](#page-4-0) of vascular tissue perfusion and early detection of flap failure are essential to achieve the best chance of successful surgical salvage and flap survival². [Monitoring](#page-4-0) of the microvascular flap is paramount to the early recognition of failure. Clinical examination of the colour, capillary refill, tissue turgor, oedema, and bleeding characteristics of the free flap is still considered the gold standard for a majority of microvascular surgeons $^{1,3-5}$. A [limitation](#page-4-0) of this approach is human error in recog-nizing changes in these parameters³[.](#page-4-0) Current technology can be used to optimize the clinical monitoring techniques, in order to analyse and detect early vascular compromise after free flap reconstruction. The ideal monitoring technique needs to be harmless to the patient, rapid, accurate, reliable, and applicable to all flap types, as well as inexpensive, easy to perform, and easily interpretable by the nursing staff $3,6$.

Asthe colour of the skin is considered to be the most predictable clinical indicator of the vascular perfusion, objective quantification of skin colour changes could be useful to examine the flap viability⁷. [The](#page-4-0) aim of this study was to determine whether a digital surface scanner can recognize early skin colour alterations as a result of a vascular compromise.

Materials and methods

A group of 10 healthy Caucasian volunteers (six male, mean age of 29 ± 5 years) were recruited prospectively. Impaired vascular perfusion was simulated at different time-points by intermittent

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occlusion of the blood supply to the forearm skin.

Inclusion criteria were (1) minimum age of 18 years, (2) non-smoker with no medical history of vascular disease, and (3) systolic blood pressure between 100 mmHg and 140 mmHg, and diastolic blood pressure between 60 mmHg and 80 mmHg. Various Fitzpatrick types (type I to IV) were included. Volunteers with a negative Allen manoeuvre, Sudeck atrophy, or who had undergone previous surgical interventions involving the forearm were excluded. The study adhered to the tenets of the Declaration of Helsinki. Institutional review board approval and informed consent were obtained (2016-2690).

Data acquisition

Multiple surface scans of the participants' forearm skin were acquired with a TRIOS 3D scanner (3Shape, Copenhagen, Denmark). One observer performed all of the surface scans according to the scanning protocol provided by the manufacturer⁸. To [simulate](#page-4-0) the radial forearm flap design, a square of 4×5 cm centred over the radial artery was drawn to define the region of interest for scanning.

A baseline scan was obtained at the beginning to document the normal physiological state. Subsequently, the cuff of a standard manual mercury sphygmomanometer was applied to the upper arm to alter the vascular perfusion of the forearm. To simulate arterial occlusion, the participant was asked to hold the arm in 180 anteflexion for 30 seconds to deplete the superficial venous blood system. The cuff was then inflated to 220 mmHg. The forearm skin was scanned after 1 and 10 minutes of arterial occlusion (respectively AO $1'$ and AO $10'$). After this 10-minute scan, the cuff was deflated to restore normal vascular flow, and the forearm skin was scanned 1 minute after deflation to measure revascularization after arterial occlusion (RAO). After a break of approximately 25 minutes, venous congestion was simulated by inflating the cuff to 40 mmHg. As this pressure is higher than central venous pressure and lower than diastolic pressure, the arterial blood supply will not be compromised. Surface scans of the forearm skin were acquired after 1 and 10 minutes of venous congestion (respectively VC $1'$ and VC $10'$). To return venous circulation, the cuff was deflated; 1 minute after deflation, the forearm skin was scanned to measure revascularization after venous congestion (RVC). Finally, 10 minutes after the last

scan, a control surface scan was obtained with the cuff removed and the forearm in neutral position. Identical procedures were performed on the contralateral forearm.

Measurements

To investigate skin colour changes, all surface scans were imported into 3DS Max v2016 software (Autodesk Inc., San Rafael, CA, USA). After cropping the irrelevant areas towards the region of interest, the surface colour was extracted in hue (H), saturation (S), and value (V) format (Fig. 1). The HSV colour scale is a cylindrical geometry whereby 'hue', ranging from 0 to 360° , is formed in a radial slice and can be seen as the basic aspect of a colour. The central axis of neutral colours ranges from black at the bottom to white at the top, representing the 'value' (between 0 and 100%) as the brightness of a colour. The 'saturation' refers to the intensity or purity of a colour (between 0 and $100\%)^9$ $100\%)^9$.

Statistical analysis

All data were analysed using IBM SPSS Statistics version 22 (IBM Corp., Armonk, NY, USA). Mean and standard deviation (SD) values were calculated. ^A paired ^ttest (two-sided) was used to compare the scan results during the different vascular stages and at the different time-points. A *P*-value of ≤ 0.01 was set as the level of statistical significance.

Results

A total of 160 digital surface scans of the forearm skin were obtained. An example of the colour changes during the different stages of vascular compromise are depicted in [Fig.](#page-2-0) 2.

The mean results of the skin colour measurements for each stage and at each time point are reported in [Table](#page-2-0) 1. The temporal evolution of skin colour changes during vascular compromise is shown in [Fig.](#page-3-0) 3. The HSV curves all showed a sinusoidal shape in which the measurements obtained during arterial occlusion displayed an opposite tendency compared to those obtained during the stage of venous congestion. The curve amplitude was most pronounced for H. Transitions of the S curve were more levelled and progressed inversely compared to the V and H curves. The colour measurements at the control stage demonstrated a return towards the baseline for H, S, and V. The skin colour changes differed significantly between the baseline measurement and immediate arterial occlusion and venous congestion for H, S, and V ($P < 0.01$ for baseline vs. AO $1'$ and for baseline vs. VC $1'$). Significant skin colour changes were observed as venous congestion continued $(P < 0.01)$ (H) for VC $1'$ vs. VC $10'$). In contrast, no significant colour changes were seen

Fig. 1. ^A cylindrical colour model defining hue (H), saturation (S), and value (V). 'Hue', ranging from 0 to 360° , can be seen as the basic aspect of a colour. The colour brightness is determined by the height of the cylinder as'value'. 'Saturation' refersto the intensity or purity of a colour and varies from the central axis towards the cylinder periphery.

Fig. 2. Successive digital surface scans of the forearm skin during different stages of simulated vascular compromise. The region of interest was marked on the forearm skin before scanning (interrupted line). Skin colour changes are illustrated during arterial occlusion (AO) and venous congestion (VC) after 1 minute $(1')$ and 10 minutes $(10')$. Immediate recovery of the normal blood flow is shown at the end (revascularization after arterial occlusion (RAO) and after venous congestion (RVC)). Although the subjective colour changes could be very subtle, digital analysis of the colour features was able to detect objective and significant differences.

Table 1. Skin colour measurements at the different stages during vascular compromise; mean \pm standard deviation values.

	Hue $(^\circ)$	Saturation $(\%)$	Value $(\%)$
Baseline	20 ± 3	34 ± 5	71 ± 4
AO ₁	$30 \pm 3^{a,c}$	$32 \pm 5^{a,c}$	$74 + 3^{a,c}$
AO 10'	$30 + 3^a$	$31 + 5^{a,b}$	$73 \pm 3^{a,b}$
RAO	$11 + 3^b$	$41 + 4^{b}$	$72 + 3^b$
VC 1'	$15 \pm 3^{\rm a}$	$35 \pm 4^{\rm a}$	$68 \pm 3^{\rm a}$
VC 10'	$12 \pm 3^{a,b}$	$35 \pm 4^{\circ}$	$65 \pm 3^{a,b}$
RVC	$16 \pm 3^{\rm b}$	35 ± 4	68 ± 3^{6}
Control	$19 \pm 3^{\rm a}$	34 ± 4	70 ± 3

1': 1 minute; 10': 10 minutes; AO, arterial occlusion; RAO, revascularization after arterial occlusion; RVC, revascularization after venous congestion; VC, venous congestion. ${}^{a}P$ < 0.01 compared to the baseline scan.

 $P^{\text{B}}P < 0.01$ compared to the baseline scan.
 $P^{\text{B}}P < 0.01$ compared to the previous stage

 $P < 0.01$ compared to the previous stage.
 $P < 0.01$ compared to venous congestion

 $P < 0.01$ compared to venous congestion after 1 minute. P-values were obtained with a paired two-sided t-test.

during prolonged arterial occlusion $(P = 0.67$ (H) for AO 1' vs. AO 10').
Revascularization after arterial occlusion Revascularization after arterial occlusion and venous congestion illustrated a significant colour shift for both H and V $(P < 0.01$ for RAO vs. AO 10' and for RVC vs. VC 10'). A significant difference in skin colour was seen for arterial occlusion and venous congestion at the same time-point ($P < 0.01$ (H) for AO 1' vs. VC 1'). Finally, the control scan at the end of the experiment and the baseline scan differed significantly for H ($P < 0.01$ for baseline vs. control), but not for S or V.

Discussion

Survival of a freeflap reconstruction depends predominantly on the vascular tissue perfusion. Ischemia due to occlusion of the supplying artery or congestion caused by obstruction of the venous outflow should be recognized as soon as possible to enhance the success rate of salvage surgery¹. [Clinical](#page-4-0) observation remains the gold standard to assess the vascularization of the transferred tissue, but it is crucial to acknowledge that this is a subjective evaluation, highly dependent on the observer's experience.

Various monitoring techniques have been developed to assist flap surveillance in a more objective way. Techniques monitoring vascular flow include hand-held Doppler ultrasonography, laser Doppler flowmetry, colour duplex ultrasonography, fluorescence angiography, implantable Doppler, and implantable flow coupler^{1,4}. Doppler [ultrasonography](#page-4-0) and laser Doppler flowmetry are both excellent non-invasive tools with high positive and negative predictive values. Unfortunately, they are not able to differentiate between arterial and venous compromise. Fluorescent angiography and colour duplex ultrasonography are great adjunctive techniques in preoperative planning and in the intra- and postoperative quantification of perfusion. Nevertheless, they are not recommended as primary flap monitoring tools given the non-continuous nature of use. Implantable Doppler and flow coupler can directly monitor the arterial and venous flow. Disadvantages of these approaches include the invasiveness of attachment of the probe around the vascular anastomosis. Techniques monitoring tissue metabolism and ischemia include near-infrared spectroscopy, which uses dyes such as indocyanine green, and microdialysis to determine the concentrations of glucose, lactate, and pyruvate in

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Fig. 3. Change in colour features during the consecutive stages of compromised vascularity of the skin. The boxplots represent the colour measurements at each stage of impaired vascularity. Evolution of the means (x) is shown for hue (H), saturation (S), and value (V) (dotted line). Statistically significant differences (*) were found between the baseline surface scan and 1 minute of arterial occlusion (baseline vs AO $1'$) and 1 minute of venous congestion (baseline vs VC $1'$), as well as between arterial occlusion and venous congestion (AO $1'$ vs VC $1'$). Measurement differences between the different stages were most obvious for H, which represents the basic feature of a colour.

the flap as surrogate markers of tissue perfusion^{4,5,10}. Both [methods](#page-4-0) show great potential for the early detection of flap failure, however they are invasive, expensive, and require some user experience for an accurate interpretation.

At this time, there is no evidence of any particular technique being superior for the early detection of vascular failure of the reconstructed tissue^{1,4}. [Optimizing](#page-4-0) the current clinical monitoring protocols with advanced technological devices could be beneficial. Since the colour of a free flap is a crucial clinical predictor of the viability of the flap, this study investigated whether the assessment of skin colour, obtained by a digital surface scanner, is able to detect early impairment of vascular tissue perfusion. An intraoral scanner was chosen for three reasons. This handheld device can provide a fast and accurate image of the soft tissue colour¹⁰. [Furthermore,](#page-4-0) the scanning procedure with the TRIOS 3D scanner is non-invasive and easy to perform, even for an inexperienced α bserver^{10–12}. [Previous](#page-4-0) studies have illustrated that scanning of the soft tissues inside the oral cavity is feasible, which makes this device more suitable for colour assessment in difficult to reach areas with changing ambient light $13,1$

The results of this study emphasize that colour assessment by digital scanning of the skin surface can detect impaired vascular tissue perfusion. Simulations of arterial occlusion and of venous congestion provoked alterations of the skin colour that were significantly different to the baseline condition. The change in skin colour due to vascular compromise could already be identified after 1 minute. This indicates that a digital surface scanner is able to discern vascular compromise at an early stage, which could be a great advantage for the rapid detection of a potential free flap failure. Furthermore, the scanner can distinguish arterial ischemia from venous congestion. This could be useful in identifying the source of vascular impairment and determining the optimal treatment.

As seen in this study, hue appears to be the most obvious colour aspect when evaluating vascular changes. In the case of an arterial occlusion, the forearm skin colour will lose its red appearance and become more yellow and pale. This correlates with the study findings, as hue increased from 20° towards 30° , indicating a shift from red–orange towards the yellow spectrum. During venous congestion, expressed as a darker red or purple aspect of the forearm skin colour, the opposite trend is observed; the scanner detected a decrease in hue from 20° to 12° .

Recovery of the vascular perfusion resulted in significant skin colour changes that were most apparent when revascularization occurred after arterial occlusion. Restoration of the circulation after venous congestion is less obvious by clinical observation, but still a significant difference in hue was detected in the present study

measurements, illustrating the benefit of the surface scan to supplement the subjective clinical assessment. While value and saturation had normalized almost entirely at the time of the control scan, hue recovered more slowly and differed significantly when compared to the baseline scan. It is likely that the measurements obtained at the time of the control scan were not completely representative yet, due to the relatively short recovery time (10 minutes); this could have been insufficient to completely reoxygenate the tissues to a normal level.

There are several limitations of the study. Further investigation is required to elucidate how the colour characteristics of the soft tissue will evolve during a surgical procedure. The surgical trauma will inevitably cause vascular and skin colour changes due to tissue reaction. Likewise, the variance in blood pressure or changes in the haemoglobin status of the patient could affect measurement outcomes. The scan results provide an objective measurement suggesting a vascular skin perfusion disorder, but there are no absolute threshold values indicating ischemia or congestion. Asskin colour values at baseline are different in every patient, the evolution of relative changes between measurement stages is most informative, and unusual colour changes should be considered as a warning sign. The study was performed on Caucasian participants and it would be interesting to investigate the skin colour changes in other ethnic groups as well. Only low-grade skin phototypes (grades I to IV) were included, and investigations should be performed to determine whether these findings also apply to the higher Fitzpatrick skin types. Finally, this study was only performed with a simulation of vascular compromise in healthy participants. Thus further research is required to allow clinical implementation of this monitoring technique after tissue transfer in patients with co-morbidities.

This study is novel in attempting to use a digital surface scanner to discern vascular tissue perfusion. Although the results show it is feasible to detect skin colour changes during various stages of impaired tissue perfusion, more studies are needed to elucidate the clinical advantages of this technique during the monitoring of a microvascular free flap.

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This study did not receive any funding.

Competing interests

There are no competing interests to disclose.

Ethical approval

The study adhered to the tenets of the Declaration of Helsinki. Institutional review board approval was obtained (2016-2690).

Patient consent

Not required.

References

- 1. Karinja SJ, Lee BT. [Advances](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0005) in flap monitoring and impact of [enhanced](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0005) recovery [protocols.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0005) J Surg Oncol 2018;118:758–67.
- 2. [Brennan](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0010) PA, Brands MT, Gush R, Alam P. [Laser-speckle](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0010) imaging to measure tissue perfusion in free flaps in oral and [maxillofa](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0010)cial surgery: a [potentially](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0010) exciting and easy to use [monitoring](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0010) method. Br J Oral Maxillofac Surg 2018;56[:556–8.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0010)
- 3. Urken ML, [Weinberg](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0015) H, Vickery C, Buchbinder D, Biller HF. Free flap [design](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0015) in head and neck [reconstruction](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0015) to achieve an external segment for [monitoring.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0015) Arch Otolaryngol Neck Surg 1989;115[:1447–53.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0015)
- 4. Kääriäinen M, Halme E, Laranne J. Modern [postoperative](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0020) monitoring of free flaps. Curr Opin [Otolaryngol](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0020) Head Neck Surg 2018;26: [248–53.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0020)
- 5. Jyranki J, [Suominen](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0025) S, Vuola J, Back L. [Microdialysis](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0025) in clinical practice: monitoring [intraoral](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0025) free flaps. Ann Plast Surg 2006;56[:387–93.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0025)
- 6. Creech B, Miller S. [Evaluation](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0030) of circulation in skin flaps. In: Grabb WC, [Myers](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0030) MB,

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editors. Skin flaps. [Boston:](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0030) Little, Brown and [Company;](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0030) 1975.

- 7. Van [Genechten](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0035) M, Rahmel B, Batstone MD. Red or white? Use of high [colour-rendering](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0035) index, [light-emitting](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0035) diodes in monitoring of free flaps of the head and [neck.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0035) Br J Oral [Maxillofac](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0035) Surg 2015;53:765–6.
- 8. 3Shape instructions. [https://www.3shape.](https://www.3shape.com/en/support-docs) [com/en/support-docs](https://www.3shape.com/en/support-docs) [Accessibility verified [September](https://www.3shape.com/en/support-docs) 6, 2019].
- 9. Li S, Guo G. The application of improved HSV color space model in image processing. Proceedings of the 2010 2nd International Conference on Future Computer and Communication ICFCC 2010. [http://dx.doi.org/](http://dx.doi.org/10.1109/ICFCC.2010.5497299) [10.1109/ICFCC.2010.5497299.](http://dx.doi.org/10.1109/ICFCC.2010.5497299) V2-10– V2-13.
- 10. Deferm JT, Schreurs R, Baan F, [Bruggink](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0050) R, [Merkx](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0050) MAW Xi T, Berge´ SJ, Maal TJJ. Validation of 3D [documentation](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0050) of palatal soft tissue shape, color, and [irregularity](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0050) with intraoral [scanning.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0050) Clin Oral Investig 2018;22[:1303–9.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0050)
- 11. Kim J, Park JM, Kim M, Heo SJ, [Shin](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0055) IH, Kim M. [Comparison](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0055) of experience curves between two [3-dimensional](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0055) intraoral scanners. J Prosthet Dent 2016;116[:221–30.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0055)
- 12. Park HR, Park JM, [Chun](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0060) YS, Lee KN, Kim M. [Changes](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0060) in views on digital intraoral scanners among dental [hygienists](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0060) after training in digital [impression](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0060) taking. BMC Oral [Health](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0060) 2015;15:151.
- 13. Kattadiyil MT, Mursic Z, [AlRumaih](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0065) H, [Goodacre](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0065) CJ. Intraoral scanning of hard and soft tissues for partial [removable](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0065) dental prosthesis [fabrication.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0065) J Prosthet Dent 2014;112[:444–8.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0065)
- 14. Gan N, Xiong Y, Jiao T. [Accuracy](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0070) of intraoral digital [impressions](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0070) for whole upper jaws, including full [dentitions](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0070) and palatal soft tissues. PLoS One 2016;11[:e0158800.](http://refhub.elsevier.com/S0901-5027(20)30202-2/sbref0070)

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