



Featured Article

Prevalence of the apolipoprotein E ε4 allele in amyloid β positive subjects across the spectrum of Alzheimer's disease

Niklas Mattsson^{a,*}, Colin Groot^b, Willemijn J. Jansen^c, Susan Landau^d, Victor Villemagne^e, Sebastiaan Engelborghs^f, Mark Mintun^g, Alberto Lleó^h, José Luis Molinuevoⁱ, William Jagust^d, Giovanni B. Frisoni^{j,k}, Adrian Ivanoiu^l, Gaël Chételat^m, Catarina Resende de Oliveiraⁿ, Karen M. Rodrigue^o, Johannes Kornhuber^p, Anders Wallin^q, Aleksandra Klimkiewicz-Mrowiec^r, Ramesh Kandimella^s, Julius Popp^t, Pauline P. Aalten^c, Dag Aarsland^u, Daniel Alcolea^h, Ina S. Almdahl^v, Inês Baldeirasⁿ, Mark A. van Buchem^w, Enrica Cavado^{k,x}, Kewei Chen^y, Ann D. Cohen^z, Stefan Förster^{aa}, Juan Fortea^h, Kristian S. Frederiksen^{bb}, Yvonne Freund-Levi^{cc}, Kiran Dip Gill^s, Olymbia Gkatzima^{dd}, Timo Grimmer^{ee}, Harald Hampel^{x,ff}, Sanna-Kaisa Herukka^{gg}, Peter Johannsen^{hh}, Koen van Laereⁱⁱ, Mony de Leon^{jj}, Wolfgang Maier^{kk}, Jan Marcusson^{ll}, Olga Meulenbroek^{mmm}, Hanne M. Møllergård^v, John C. Morrisⁿⁿ, Barbara Mroczko^{oo}, Arto Nordlund^q, Sudesh Prabhakar^{pp}, Oliver Peters^{qq}, Lorena Ramiⁱ, Eloy Rodríguez-Rodríguez^{rr}, Catherine M. Roeⁿⁿ, Eckart Rüther^{ss}, Isabel Santanaⁿ, Johannes Schröder^{tt}, Sang W. Seo^{uu}, Hilka Soininen^{gg}, Luiza Spuru^{vv}, Erik Stomrud^a, Hanne Struyfs^f, Charlotte E. Teunissen^{ww}, Frans R. J. Verhey^c, Stephanie J. B. Vos^c, Linda J. C. van Waalwijk van Doorn^{xx,yy}, Gunhild Waldemar^{bb}, Åsa K. Wallin^a, Jens Wiltfang^{ss,zz}, Rik Vandenberghe^{aaa}, David J. Brooks^{bbb}, Tormod Fladby^v, Christopher C. Rowe^e, Alexander Drzezga^{ccc}, Marcel M. Verbeek^{xx,yy}, Marie Sarazin^{ddd}, David A. Wolk^{eee}, Adam S. Fleisher^{y,fff,ggg}, William E. Klunk^z, Duk L. Na^{uu}, Pascual Sánchez-Juan^{rr}, Dong Young Lee^{hhh}, Agneta Nordbergⁱⁱⁱ, Magda Tsolaki^{dd}, Vincent Camus^{jjj}, Juha O. Rinne^{kkk}, Anne M. Faganⁿⁿ, Henrik Zetterberg^{lll,mmm,nnn,ooo}, Kaj Blennow^{nnn,ooo}, Gil D. Rabinovici^{ppp}, Oskar Hansson^a, Bart N. M. van Berckel^{qqq}, Wiesje M. van der Flier^{b,rrr}, Philip Scheltens^b, Pieter Jelle Visser^{b,c}, Rik Ossenkoppele^{a,b,qqq,**}

^aClinical Memory Research Unit, Clinical Sciences Malmö, Lund University, Lund, Sweden

^bDepartment of Neurology and Alzheimer Center, VU University Medical Center, Neuroscience Campus Amsterdam, Amsterdam, the Netherlands

^cDepartment of Psychiatry and Neuropsychology, School for Mental Health and Neuroscience, Alzheimer Center Limburg, Maastricht University, Maastricht, the Netherlands

^dHelen Wills Neuroscience Institute, University of California, Berkeley, CA, USA

^eDepartment of Nuclear Medicine and Centre for PET, Austin Health, Melbourne, Australia

^fReference Center for Biological Markers of Dementia (BIODEM), University of Antwerp, Antwerp, Belgium

^gAvid Radiopharmaceuticals, Philadelphia, PA, USA

^hNeurology Department, Hospital de Sant Pau, Barcelona, Spain

ⁱAlzheimer's Disease and Other Cognitive Disorders Unit, IDIBAPS, Clinic University Hospital, Barcelona, Spain

^jMemory Clinic and LANVIE- Laboratory of Neuroimaging of Aging, University Hospitals, and University of Geneva, Geneva, Switzerland

^kLaboratory of Alzheimer's Neuroimaging and Epidemiology, IRCCS Centro San Giovanni di Dio Fatebenefratelli, Brescia, Italy

*Corresponding author. Tel.: ■■■■; Fax: ■■■■.

E-mail address: niklas.mattsson@med.lu.se (N.M.), r.ossenkoppele@

**Corresponding author. Tel.: ■■■■; Fax: ■■■■.

vumc.nl (R.O.)

<https://doi.org/10.1016/j.jalz.2018.02.009>

1552-5260/© 2018 the Alzheimer's Association. Published by Elsevier Inc. All rights reserved.

- ^lMemory Clinic and Neurochemistry Laboratory, Saint Luc University Hospital, Institute of Neuroscience, Université catholique de Louvain, Brussels, Belgium 177
- ^mInserm, Inserm UMR-S U1237, Université de Caen-Normandie, GIP Cyceron, Caen, France 178
- ⁿCenter for Neuroscience and Cell Biology, Faculty of Medicine, Centro Hospitalar e Universitário de Coimbra, Portugal 179
- ^oCenter for Vital Longevity, School of Behavioral and Brain Sciences, The University of Texas at Dallas, Dallas, TX, USA 180
- ^pDepartment of Psychiatry and Psychotherapy, Friedrich-Alexander University of Erlangen- Nuremberg, Erlangen, Germany 181
- ^qInstitute of Neuroscience and Physiology, Sahlgrenska Academy at University of Gothenburg, Mölndal, Sweden 182
- ^rJagiellonian University College of Medicine, Krakow, Poland 183
- ^sPostgraduate Institute of Medical Education and Research (PGIMER), Department of Biochemistry, Research Block-A, Chandigarh, India 184
- ^tDepartment of Psychiatry, Service of Old Age Psychiatry, University Hospital of Lausanne, Lausanne, Switzerland 185
- ^uCenter for Age-Related Medicine, Stavanger University Hospital, Stavanger, Norway 186
- ^vDepartment of Neurology, Akershus University Hospital, Lørenskog, Norway 187
- ^wDepartment of Radiology, Leiden University Medical Center, Leiden, the Netherlands 188
- ^xAXA Research Fund & UPMC Chair, Sorbonne Universités, Université Pierre et Marie Curie (UPMC) Paris 06, Inserm, CNRS, Institut du Cerveau et de la Moelle Épinière (ICM), Département de Neurologie, Institut de la Mémoire et de la Maladie d'Alzheimer (IM2A), Hôpital Pitié-Salpêtrière, Paris, France 189
- ^yBanner Alzheimer's Institute, Phoenix, AZ, USA 190
- ^zUniversity of Pittsburgh School of Medicine, Department of Psychiatry, Pittsburgh, PA, USA 192
- ^{aa}Department of Nuclear Medicine, Technische Universitaet München, Munich, Germany 193
- ^{bb}Danish Dementia Research Center, Department of Neurology, Rigshospitalet, University of Copenhagen, Copenhagen, Denmark 194
- ^{cc}Department of Geriatrics, Karolinska University Hospital Huddinge, Section of Clinical Geriatrics, Institution of NVS, Karolinska Institutet, Stockholm, Sweden 195
- ^{dd}Third Department of Neurology, Aristotle University of Thessaloniki, Thessaloniki, Greece 196
- ^{ee}Department of Psychiatry and Psychotherapy, Klinikum rechts der Isar der Technischen Universitaet München, Munich, Germany 197
- ^{ff}Department of Psychiatry, Alzheimer Memorial Center and Geriatric Psychiatry Branch, Ludwig-Maximilian University, Munich, Germany 199
- ^{gg}Department of Neurology, University of Eastern Finland and Kuopio University Hospital, Kuopio, Finland 200
- ^{hh}Memory Clinic, Danish Dementia Research Center, Rigshospitalet, Copenhagen, Denmark 201
- ⁱⁱDepartment of Imaging and Pathology, Catholic University Leuven, Leuven, Belgium 202
- ^{jj}School of Medicine, Center for Brain Health, New York University, New York, NY, USA 203
- ^{kk}Department of Psychiatry and Psychotherapy, University of Bonn, German Center for Neurodegenerative Diseases (DZNE), Bonn, Germany 204
- ^{ll}Geriatric Medicine, Department of Clinical and Experimental Medicine, University of Linköping, Linköping, Sweden 205
- ^{mm}Department of Geriatric Medicine, Radboud Alzheimer Center, Radboud University Medical Center, Nijmegen, the Netherlands 206
- ⁿⁿKnight Alzheimer's Disease Research Center, Department of Neurology, Washington University School of Medicine, St Louis, MO, USA 207
- ^{oo}Department of Neurodegeneration Diagnostics, Leading National Research Centre in Białystok (KNOW), Medical University of Białystok, Białystok, Poland 208
- ^{pp}Department of Neurology, Research Block-A, Chandigarh, India 209
- ^{qq}Department of Psychiatry and Psychotherapy, Charité Berlin, German Center for Neurodegenerative Diseases (DZNE), Berlin, Germany 210
- ^{rr}Neurology Service, University Hospital Marqués de Valdecilla, CIBERNED, IDIVAL, Santander, Spain 211
- ^{ss}Department of Psychiatry and Psychotherapy, University Medical Center, Georg-August University, Göttingen, Germany 212
- ^{tt}Sektion Gerontopsychiatrie, Universität Heidelberg, Heidelberg, Germany 213
- ^{uu}Department of Neurology, Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, South Korea 214
- ^{vv}Department of Geriatrics-Gerontology-Gerontopsychiatry, Carol Davila University of Medicine and Pharmacy, Bucharest, Romania 215
- ^{ww}Neurochemistry Laboratory and Biobank, Department of Clinical Chemistry, Amsterdam Neuroscience, VU University Medical Center, Amsterdam, the Netherlands 216
- ^{xx}Department of Neurology, Donders Institute for Brain, Cognition and Behaviour, Radboud Alzheimer Center, Radboud University Medical Center, Nijmegen, the Netherlands 217
- ^{yy}Department of Laboratory Medicine, Donders Institute for Brain, Cognition and Behaviour, Radboud Alzheimer Center, Radboud University Medical Center, Nijmegen, the Netherlands 218
- ^{zz}Department of Psychiatry and Psychotherapy, University Medical Center, Georg-August University, Göttingen, Germany 219
- ^{aaa}Laboratory for Cognitive Neurology and Alzheimer Research Centre KU Leuven, Catholic University Leuven, Leuven, Belgium 220
- ^{bbb}Division of Neuroscience, Medical Research Council Clinical Sciences Centre, Imperial College London, London, UK 221
- ^{ccc}Department of Nuclear Medicine, University of Cologne, Cologne, Germany 222
- ^{ddd}Neurologie de la Mémoire et du Langage, Centre Hospitalier Sainte-Anne, Université Paris Descartes, Sorbonne Paris Cité, Paris, France 223
- ^{eee}Department of Neurology, University of Pennsylvania, Philadelphia, PA, USA 224
- ^{fff}Eli Lilly, Indianapolis, IN, USA 225
- ^{ggg}Department of Neurosciences, University of California, San Diego, CA, USA 226
- ^{hhh}Department of Neuropsychiatry, Seoul National University, College of Medicine, Seoul, South Korea 227
- ⁱⁱⁱDepartment NVS, Center for Alzheimer Research, Translational Alzheimer Neurobiology, Karolinska Institutet and Geriatric Medicine, Karolinska University Hospital, Stockholm, Sweden 228
- ^{jjj}CHRU de Tours, CIC INSERM 1415, INSERM U930, Université François Rabelais de Tours, Tours, France 229
- ^{kkk}Turku PET Centre and Division of Clinical Neurosciences Turku, University of Turku and Turku University Hospital, Turku, Finland 230
- ^{lll}Department of Molecular Neuroscience, UCL Institute of Neurology, Queen Square, London, UK 231
- ^{mmm}UK Dementia Research Institute, London, UK 232
- ⁿⁿⁿInstitute of Neuroscience and Physiology, Department of Psychiatry and Neurochemistry, The Sahlgrenska Academy at University of Gothenburg, Mölndal, Sweden 233
- ^{ooo}Sweden and Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden 234
- ^{ppp}Department of Neurology, Memory and Aging Center, University of California, San Francisco, CA, USA 235
- ^{qqq}Department of Radiology and Nuclear Medicine, VU University Medical Center, Neuroscience Campus Amsterdam, Amsterdam, the Netherlands 236
- ^{rrr}Department of Epidemiology and Biostatistics, VU University Medical Center, Amsterdam, the Netherlands 237

Q3 Abstract

Introduction: Apolipoprotein E (*APOE*) $\epsilon 4$ is the major genetic risk factor for Alzheimer's disease (AD), but its prevalence is unclear because earlier studies did not require biomarker evidence of amyloid β ($A\beta$) pathology.

Methods: We included 3451 $A\beta+$ subjects (853 AD-type dementia, 1810 mild cognitive impairment, and 788 cognitively normal). Generalized estimating equation models were used to assess *APOE* $\epsilon 4$ prevalence in relation to age, sex, education, and geographical location.

Results: The *APOE* $\epsilon 4$ prevalence was 66% in AD-type dementia, 64% in mild cognitive impairment, and 51% in cognitively normal, and it decreased with advancing age in $A\beta+$ cognitively normal and $A\beta+$ mild cognitive impairment ($P < .05$) but not in $A\beta+$ AD dementia ($P = .66$). The prevalence was highest in Northern Europe but did not vary by sex or education.

Discussion: The *APOE* $\epsilon 4$ prevalence in AD was higher than that in previous studies, which did not require presence of $A\beta$ pathology. Furthermore, our results highlight disease heterogeneity related to age and geographical location.

© 2018 the Alzheimer's Association. Published by Elsevier Inc. All rights reserved.

Keywords:

APOE; Prevalence; Amyloid; PET; CSF; Alzheimer's disease; Mild cognitive impairment; Subjective cognitive decline; Age; Sex; Education; Geographical location

1. Introduction

Alzheimer's disease (AD) is the most common type of dementia and a major cause of morbidity and mortality worldwide [1]. Pathological metabolism and accumulation of amyloid β ($A\beta$) peptides are thought to be an initiating event in AD, leading to downstream spread of tau pathology, synaptic loss, neurodegeneration, and cognitive decline [2–4]. The main risk factors for the development of AD are increasing age and the $\epsilon 4$ allele of the apolipoprotein E (*APOE*) gene [5–7], the strongest genetic risk factor for sporadic AD [8,9]. *APOE* encodes for apolipoprotein E, which is a major lipid transporting protein in the brain [10]. In humans, the gene exists in three allele variants called $\epsilon 2$, $\epsilon 3$, and $\epsilon 4$. Compared with *APOE* $\epsilon 3/\epsilon 3$ (the most common genotype), *APOE* $\epsilon 4$ heterozygosity increases the risk for developing clinical AD by about 3–4 times and *APOE* $\epsilon 4$ homozygosity by about 10–15 times [8,11]. The overall prevalence of *APOE* $\epsilon 4$ positivity has been reported to be approximately 15%–20% in the normal population [11,12] and 50%–60% in patients with AD dementia [8,9,13]. These numbers, however, vary widely and may depend on different characteristics of the study population, including ethnicity [14] and geographical location [13]. In addition, most previous studies included clinically diagnosed AD patients without neuropathological confirmation and/or supportive pathophysiological AD biomarkers. Studies applying cerebrospinal fluid (CSF) and positron emission tomography (PET) have revealed that a substantial proportion of patients with a clinical diagnosis of AD dementia have no evidence of $A\beta$ pathology [15–18], which makes the underlying AD pathology highly unlikely. This mismatch between the clinical diagnosis and $A\beta$ biomarkers seems especially prevalent in *APOE* $\epsilon 4$ noncarriers, as illustrated by a clinical trial in which 36% of *APOE* $\epsilon 4$ -negative patients with a diagnosis of “AD dementia” lacked $A\beta$ pathology as determined by PET [19]. Earlier studies emphasize

the importance of the matter, as *APOE* $\epsilon 4$ was found to be more strongly associated with biomarker evidence of $A\beta$ pathology (irrespective of clinical status) than a clinical diagnosis of AD [20]. Similarly, the effect size of *APOE* $\epsilon 4$ increased if the presence or absence of $A\beta$ pathology was neuropathologically confirmed [21].

Another critical point of previous studies is the focus on the dementia stage of AD. AD is believed to follow a long trajectory in which $A\beta$ pathology is present, and clinical symptoms gradually develop before the threshold for dementia is reached [22–24]. Few studies have investigated *APOE* $\epsilon 4$ positivity in prodromal AD [25], that is, mild cognitive impairment (MCI) due to AD ($A\beta$ biomarker positive), but prevalence rates around 25%–55% have been reported. Similarly, not many studies reported the proportion of *APOE* $\epsilon 4$ carriers among people with preclinical AD, that is, presence of $A\beta$ pathology without clinical symptoms [26–29].

In the present study, we aimed to investigate the prevalence of *APOE* $\epsilon 4$ positivity across the clinical and preclinical spectrum of AD in a large sample of $A\beta$ biomarker-positive individuals, including cognitively normal (CN) controls, MCI, and AD dementia. We also tested whether the prevalence of *APOE* $\epsilon 4$ positivity varied by age, sex, and geographical location. For comparison, we included a group of $A\beta$ -negative participants.

2. Methods

2.1. Participants

We used data from the Amyloid Biomarker Study Group, which is a worldwide collaborative project on $A\beta$ PET and CSF biomarkers in conjunction with demographic, clinical, and genetic variables [5,30,31]. From all contributing sites, we received individual participant-level data on 9480 individuals (3903 CN, 4189 MCI, 1359 probable AD dementia,

and 538 non-AD dementia). Because we aimed to investigate the prevalence of *APOE* $\epsilon 4$ across the spectrum of AD, we applied the following selection procedure for this study: (1) we excluded patients with a clinical diagnosis of non-AD dementia; (2) among CN, MCI, or AD dementia participants, we selected $A\beta$ -positive ($A\beta+$) individuals as determined by PET and/or CSF and their $A\beta$ -negative ($A\beta-$) counterparts for comparison; and (3) we excluded individuals who lacked information on *APOE* $\epsilon 4$ status.

Normal cognition was defined as normal scores on cognitive tests, the absence of cognitive complaints (for which medical help was sought), or both [5,31]. Some of the CN participants had subjective cognitive decline (SCD, $n = 533$ [102 $A\beta+$ and 431 $A\beta-$]), defined as the presence of a cognitive complaint but normal cognition on neuropsychological tests [32]. We combined the SCD subjects with the other CN participants [24,33], except for one subanalysis (Section 3.7). MCI and probable AD dementia were defined according to established diagnostic criteria [22,23,34]. $A\beta-$ “AD dementia” cases most likely do not have AD as the underlying cause of their cognitive impairment, although it should be noted that $A\beta$ biomarkers could misclassify subjects, especially when biomarker signals are close to the cutoffs [35,36].

2.2. PET or CSF procedures

Individual PET scans were dichotomized ($A\beta+$ or $A\beta-$) using quantitative thresholds or visual reads according to the method used at the study site [5,30]. CSF biomarkers were dichotomized as negative (normal) or positive (abnormal) using study-specific cutoffs [5]. For AD dementia patients, we only had PET data available [30]. For CN and MCI patients, we selected the first available biomarker in time if a participant had both PET and CSF data [5]. Detailed PET or CSF procedures for each site are presented in [Supplementary Table 1](#).

2.3. APOE genotyping

By design, all participants in this study had data on *APOE* $\epsilon 4$ status. For 2955/3114 (95.5%) CN and 3054/3335 (91.6%) MCI subjects, we had specific genotypes (e.g., $\epsilon 3/\epsilon 4$, in addition to *APOE* $\epsilon 4$ status), which allowed breakdown into *APOE* $\epsilon 4$ noncarriers, heterozygotes, and homozygotes. Specific genotypes were not available for AD dementia patients, as they were only collected for CN and MCI participants in our previous studies [5,30].

2.4. Age, sex, education, and geographical location

Information on age at time of clinical assessment was available for all participants. There were missing data for sex (130/7,419, 1.8%) and years of education (1137/7,419, 15.3%). We used a previously published classification system for geographical location [13] to divide the participants into Southern Europe ($n = 653$ [215 $A\beta+$, 438 $A\beta-$]), Central

Europe ($n = 832$ [343 $A\beta+$, 489 $A\beta-$]), Northern Europe ($n = 1667$ [792 $A\beta+$, 875 $A\beta-$]), Australia ($n = 395$ [190 $A\beta+$, 205 $A\beta-$]), North America ($n = 3359$ [1292 $A\beta+$, 2067 $A\beta-$]), or Asia ($n = 315$ [114 $A\beta+$, 201 $A\beta-$]). Some participants ($n = 637$ [303 $A\beta+$, 334 $A\beta-$], 8.1%) could not be classified, as they were included in a multicenter study that covered multiple geographical locations.

2.5. Statistical analyses

Baseline differences were assessed using analysis of variance (with post hoc Bonferroni correction) and χ^2 tests. The prevalence of *APOE* $\epsilon 4$ positivity was defined by calculating the percentage of *APOE* $\epsilon 4$ -positive individuals of the total number of participants in each diagnostic group. Generalized estimating equations were used to estimate the effects of age, sex, education, and geographical location on the prevalence of *APOE* $\epsilon 4$ positivity. Generalized estimating equations were the method of choice for the study as it allows analysis of binary-correlated data, such that participant-level data from all cohorts can be modeled while simultaneously accounting for participants within studies. A logit link function for binary outcomes with an exchangeable correlation structure was assumed to account for within-study correlation. Analyses were conducted using the total study population, unless specified otherwise. Age was entered as a continuous measure centered at the mean. We tested two- and three-way interactions between variables, and these terms were retained in the model if they appeared significant by the Wald statistical test. The generalized estimating equations derived unstandardized β coefficients, and standard errors of the main effect were reported. Significance was set at $P < .05$ (two-sided). SPSS software (IBM, version 23.0) was used for statistics.

3. Results

3.1. Participants

Demographic and clinical information for each diagnostic group is provided in [Table 1](#). We included 7419 subjects, among which 970 with a clinical diagnosis of AD dementia (853 $A\beta+$ and 117 $A\beta-$), 3335 with MCI (1810 $A\beta+$ and 1525 $A\beta-$), and 3114 CN subjects (788 $A\beta+$ and 2326 $A\beta-$). Demographic differences among the diagnostic groups included fewer males in the CN group ($P < .05$) and less education in the MCI group compared with the other groups ($P < .001$). Furthermore, in the dementia group, $A\beta$ status was only determined using PET, whereas in the MCI group, the proportion of subjects with CSF data (78%) was greater than that in the CN group (64.9%). In $A\beta+$ individuals, comparisons within diagnostic groups between *APOE* $\epsilon 4$ positive and negative groups showed that the mean age was lower in *APOE* $\epsilon 4$ -positive than that in *APOE* $\epsilon 4$ -negative CN and MCI patients ($P < .01$) ([Supplementary Table 2](#)). [Supplementary Table 3](#) shows the demographic and clinical characteristics

Table 1
Participant characteristics

	CN			MCI			AD dementia		
	Total	Aβ ⁻	Aβ ⁺	Total	Aβ ⁻	Aβ ⁺	Total	Aβ ⁻	Aβ ⁺
N	3552	2764	788	3335	1525	1810	970	117	853
Age*, mean	67.3 ± 11.8	65.8 ± 12.0	72.6 ± 9.4	70.2 ± 8.6	68.4 ± 8.9	71.8 ± 8.0	69.4 ± 9.4	71.6 ± 9.6	69.1 ± 9.3
Age, range	18–109	18–93	32–109	36–97	36–91	44–97	37–95	48–90	37–95
Sex† (% male)	43.9	42.9	47.2	53.6	54.8	52.7	56.4	64.1	55.3
MMSE‡, mean	29.0 ± 1.2	29.0 ± 1.2	28.8 ± 1.3	26.9 ± 2.5	26.7 ± 2.6	26.5 ± 2.6	21.8 ± 4.8	22.9 ± 4.0	21.6 ± 4.9
Education§, yrs	14.3 ± 3.7	14.3 ± 3.7	14.3 ± 3.8	12.4 ± 4.4	11.9 ± 4.3	12.9 ± 4.4	13.8 ± 3.6	13.6 ± 3.6	13.9 ± 3.6
Modality for Aβ positivity (% PET vs. % CSF)	41.6/58.4	42.9/57.1	36.1/63.9	22.0/78.0	21.0/79.0	22.8/77.2	100/0	100/0	100/0
APOE ε4 positivity¶ (%)	30.5	24.6	50.9	47.2	27.9	63.5	61.1	24.8	66.1
Region									
North America, n	1469	1044	425	1077	412	665	375	50	325
% APOE ε4 positive	432 (29.4)	238 (22.8)	194 (45.6)	522 (48.5)	96 (23.3)	426 (64.1)	227 (60.5)	7 (14)	220 (67.7)
Australia, n	200	140	60	76	26	50	118	4	114
% APOE ε4 positive	76 (38)	38 (27.1)	38 (63.3)	42 (55.3)	4 (15.4)	38 (76.0)	72 (61.0)	-	72 (63.2)
Northern Europe, n	712	568	144	714	365	349	241	38	203
% APOE ε4 positive	251 (35.3)	164 (28.9)	87 (60.4)	375 (52.5)	125 (34.2)	250 (71.6)	166 (68.9)	16 (42.1)	150 (73.9)
Central Europe, n	195	154	41	536	304	232	101	12	89
% APOE ε4 positive	60 (30.8)	36 (23.4)	24 (58.5)	223 (41.6)	92 (30.3)	131 (56.5)	60 (59.4)	2 (16.7)	58 (65.2)
Southern Europe, n	269	221	48	343	163	180	41	1	40
% APOE ε4 positive	61 (22.7)	43 (19.5)	18 (37.5)	135 (39.4)	37 (22.7)	98 (54.4)	19 (46.3)	0 (0)	19 (47.5)
Asia, n	80	71	9	141	76	65	94	12	82
% APOE ε4 positive	18 (22.5)	14 (19.7)	4 (44.4)	47 (33.3)	10 (13.2)	37 (56.9)	49 (52.1)	4 (33.3)	45 (54.9)

Abbreviations: Aβ, amyloid β; CN, cognitively normal; MCI, mild cognitive impairment; AD, Alzheimer's disease; MMSE, Mini-Mental State Examination; PET, positron emission tomography; CSF, cerebrospinal fluid; APOE, apolipoprotein E.

NOTE. Data are presented as mean ± SD unless indicated otherwise. Differences between diagnostic groups (assessed separately for Aβ-positive and Aβ-negative groups) were assessed using analysis of variance (age, education, and MMSE) and χ^2 tests (sex, modality, and APOE ε4 status) with post hoc Bonferroni tests.

*Aβ⁻ CN < MCI/AD, $P < .001$; MCI < AD, $P < .01$; Aβ⁺ CN/MCI > AD dementia, $P < .001$.

†Aβ⁻ CN < MCI/AD, $P < .05$; Aβ⁺ CN > MCI/AD dementia, $P < .05$.

‡Aβ⁻ CN < MCI/AD, $P < .001$; MCI < AD, $P < .05$; Aβ⁺ AD dementia < CN/MCI, $P < .001$; MCI < CN, $P < .001$.

§Aβ⁻ MCI < CN/AD, $P < .001$; Aβ⁺ MCI < CN/AD dementia, $P < .001$.

||Aβ⁻ AD > MCI/CN, CN > MCI, $P < .001$; Aβ⁺ AD dementia > CN/MCI, $P < .001$; CN > MCI, $P < .001$.

¶Aβ⁺ AD dementia/MCI > CN, $P < .001$.

of individuals tested versus not tested for APOE in the complete Amyloid Biomarker Study Group data set [5,30,31].

3.2. Prevalence of APOE ε4 positivity

In Aβ⁺ subjects, the prevalence of APOE ε4 positivity was 50.9% in CN, 63.5% in MCI, and 66.1% in AD dementia (Table 1). The prevalence of APOE ε4 positivity was higher in Aβ⁺ MCI and Aβ⁺ AD dementia than that in Aβ⁺ CN ($P < .001$), but there was no difference between Aβ⁺ MCI and Aβ⁺ AD dementia ($P = .19$). For comparison, the APOE ε4 prevalence in Aβ⁻ subjects was 24.5% in CN, 27.9% in MCI, and 24.8% in AD dementia, which was significantly lower than that in Aβ⁺ counterparts (all $P < .001$).

3.3. Prevalence of APOE ε4 positivity by age, sex, education, and modality

The prevalence of APOE ε4 positivity was lower at older age in Aβ⁺ CN (β for change in prevalence per year ± standard error: -0.02 ± 0.01 , $P < .05$, Fig. 1) and Aβ⁺ MCI

($\beta = -0.03 \pm 0.01$, $P < .01$). For example, at age 50, the prevalence of APOE ε4 positivity was 61% in Aβ⁺ CN and 75% in Aβ⁺ MCI, compared with 42% and 47% at age 90, respectively (Supplementary Fig. S1). There was no age effect on AD dementia ($\beta = 0.01 \pm 0.01$, $P = .66$). There was also no effect of age in AD dementia when excluding patients ($n = 91$) with a known atypical presentation, who are typically associated with lower prevalence of APOE ε4 ($\beta = 0.00 \pm 0.01$, $P = .99$, Supplementary Fig. S2). In Aβ⁻ subjects, the prevalence of APOE ε4 also decreased with age in CN ($\beta = -0.03 \pm 0.01$, $P < .001$; difference with Aβ⁺: $P = .62$) and MCI ($\beta = -0.03 \pm 0.01$, $P < .001$; difference with Aβ⁻: $P = .82$) but not in AD dementia ($\beta = -0.01 \pm 0.02$, $P = .55$; difference with Aβ⁺: $P = .19$). All effects described previously were similar when adjusting for sex and education.

In Aβ⁺ subjects, sex and education had no direct effects on APOE ε4 positivity, either across or within diagnostic groups (all $P > .05$). Furthermore, in Aβ⁺ subjects, there was an interaction between age and sex ($P < .05$), whereby prevalence decreased with age for women but not for men. Examining the three-way interaction with diagnosis

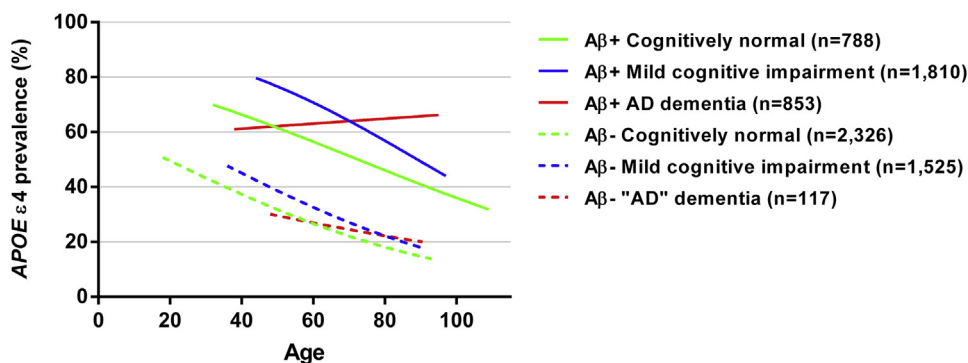


Fig. 1. Prevalence of *APOE* $\epsilon 4$ positivity by age, diagnosis, and $A\beta$ status. Curves were plotted using the point estimates generated by generalized estimating equations and are within the age limits of the diagnostic groups. The models were adjusted for study (site) effect. The 95% confidence intervals are presented in [Supplementary Fig. S1](#). Abbreviations: $A\beta$, amyloid β ; AD, Alzheimer's disease; *APOE*, apolipoprotein E.

revealed that the interaction between age and sex was present in MCI ($P < .01$), and at trend level in AD dementia ($P = .053$), but not in CN subjects ($P = .26$). In $A\beta^-$ MCI subjects, there was a trend toward higher prevalence of *APOE* $\epsilon 4$ positivity in women (β : 0.19 ± 0.10 , $P = .06$). There were no direct or interaction effects for education and no interaction effects (all $P > .05$). The prevalence of *APOE* $\epsilon 4$ positivity was higher for CSF than for PET only in $A\beta^-$ MCI subjects ($\chi^2 = 6.68$, $P = .01$; [Supplementary Table 4](#)). See [Supplementary Table 5](#) for an overview of all main and interaction effects.

3.4. Prevalence of specific *APOE* genotypes in CN and MCI

Next, we stratified CN ($n = 2955$ [751 $A\beta^+$ and 2204 $A\beta^-$]) and MCI ($n = 3054$ [1638 $A\beta^+$ and 1416 $A\beta^-$]) subjects with *APOE* genotype information available into groups of *APOE* $\epsilon 4$ noncarriers, *APOE* $\epsilon 4$ heterozygotes, and *APOE* $\epsilon 4$ homozygotes, and divided them into quartiles according to age. Both in CN and MCI subjects, the proportion of *APOE* $\epsilon 4$ heterozygotes and *APOE* $\epsilon 4$ homozygotes decreased with advancing age ([Fig. 2](#)). Prevalence of the specific genotypes (i.e., *APOE* $\epsilon 2/\epsilon 2$, $\epsilon 2/\epsilon 3$, $\epsilon 2/\epsilon 4$, $\epsilon 3/\epsilon 3$, $\epsilon 3/\epsilon 4$, and $\epsilon 4/\epsilon 4$) is provided in [Table 2](#).

3.5. Prevalence of *APOE* $\epsilon 4$ positivity by geographical location

Next, we assessed the effect of geographical location on prevalence of *APOE* $\epsilon 4$ positivity. Within $A\beta^+$ subjects, we found that the prevalence of *APOE* $\epsilon 4$ positivity across diagnostic groups was higher in Northern Europe than that in all other geographical locations except Australia (all $P < .001$, Bonferroni corrected; [Fig. 3A](#)). In addition, the prevalence of *APOE* $\epsilon 4$ positivity was lower in Southern Europe than that in North America, Central Europe ($P < .05$, uncorrected), and Australia ($P < .001$, Bonferroni-corrected), and higher in Australia than that in Asia ($P < .05$, uncorrected). Within $A\beta^-$ subjects, the prevalence

of *APOE* $\epsilon 4$ positivity was higher in Northern Europe ($P < .001$, Bonferroni-corrected) and Central Europe ($P < .05$, uncorrected) than that in all other geographical locations ([Fig. 3B](#)). These findings were similar when assessing each diagnostic group separately ([Supplementary Fig. S3](#), [Supplementary Table 5](#)).

3.6. Predictive effect of *APOE* $\epsilon 4$ status on disease stage

Finally, to assess whether the *APOE* allele is predictive of AD dementia or MCI beyond its effect on $A\beta$, we performed binary logistic regression models, including age, sex, education, $A\beta$ status (positive or negative), and *APOE* $\epsilon 4$ status (positive or negative) for CN versus MCI and CN versus AD. We found that *APOE* $\epsilon 4$ status predicted both CN versus MCI (odds ratio: 1.629, 95% confidence interval: 1.348–1.968, $P < .001$) and CN versus AD (odds ratio: 1.811, 95% confidence interval: 1.457–2.251, $P < .001$).

3.7. Prevalence of *APOE* $\epsilon 4$ positivity by SCD

The prevalence of *APOE* $\epsilon 4$ was higher in participants with SCD than those without, both among $A\beta^+$ (64.7% vs. 48.8%, $P < .05$) and $A\beta^-$ (33.6% vs. 22.4%, $P < .05$) subjects ([Supplementary Table 6](#)). The relationship between age and *APOE* prevalence was not affected by the presence or absence of SCD (all $P < .05$).

4. Discussion

We found that the prevalence of *APOE* $\epsilon 4$ positivity was 51% in preclinical AD ($A\beta^+$ CN), 64% in prodromal AD ($A\beta^+$ MCI), and 66% in $A\beta^+$ AD dementia. Among $A\beta^-$ subjects, the prevalence of *APOE* $\epsilon 4$ positivity was 25% in CN, 28% in MCI, and 25% in AD dementia. Our estimates of *APOE* $\epsilon 4$ prevalence in $A\beta$ biomarker-verified AD-type dementia are higher than reported in previous studies that defined AD-type dementia based on clinical criteria. This resonates well with studies examining the effect size of *APOE* $\epsilon 4$ in pathology- or biomarker-confirmed cases [[20,21](#)] and suggests that the prevalence

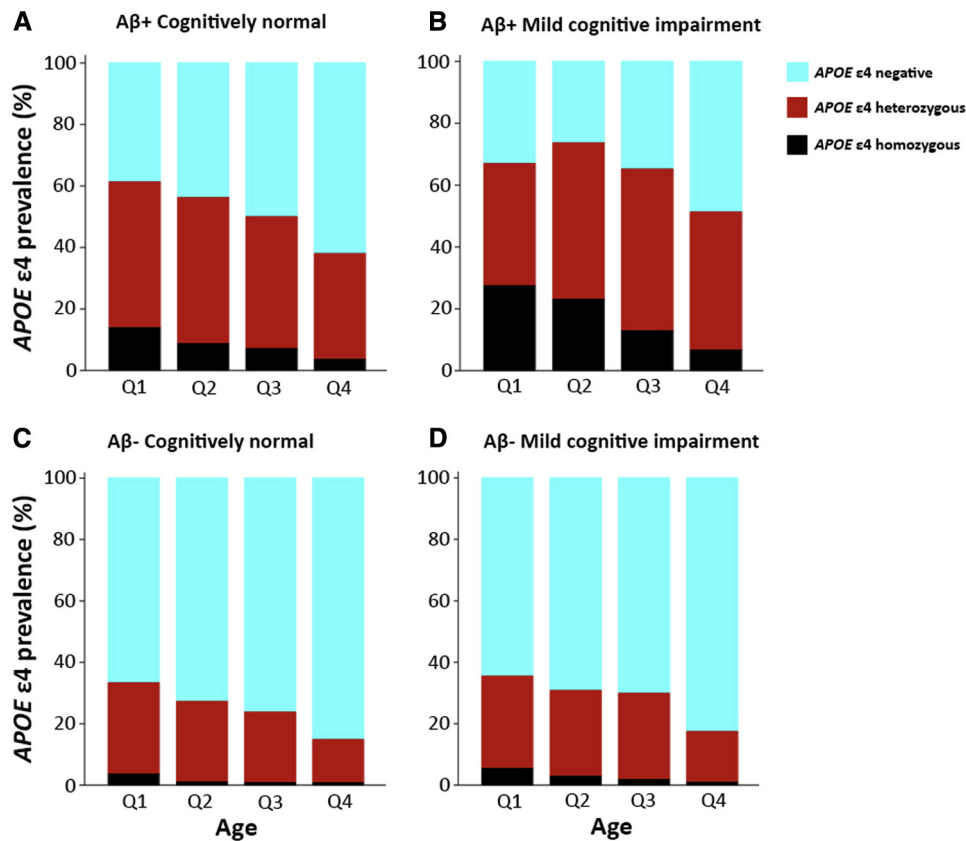


Fig. 2. Distribution of *APOE* ε4 negative, *APOE* ε4 heterozygous, and *APOE* ε4 homozygous subjects across different age quartiles ([A]; Q1 = <67 years, Q2 = 67–73.2, Q3 = 73.21–78.76, Q4 = >78.77 years; [B]; Q1 = <66.67 years, Q2 = 66.68–72.28, Q3 = 72.29–77.19, Q4 = >77.2; [C]; Q1 = <59.5 years, Q2 = 59.5–67.1, Q3 = 67.11–75.65, Q4 = >73.66 years; [D]; Q1 = <62 years, Q2 = 62.01–68.41, Q3 = 68.42–75.0, Q4 = >75.01 years). Abbreviations: Aβ, amyloid β; *APOE*, apolipoprotein E; Q, quartile.

of *APOE* ε4 in AD-type dementia (66%) may have been underestimated in previous studies (50%–60% [8,9,13]).

Another main finding of this study was that the prevalence of *APOE* ε4 decreased with age in preclinical and prodromal AD. There are several possible explanations. First, the additive effects of *APOE* ε4 and Aβ may have resulted in greater conversion from the CN and MCI groups to AD dementia [37]. Higher conversion rates could also be due to earlier and more pronounced accumulation of Aβ load

in *APOE* ε4 carriers [38], but the binary nature (Aβ positive or negative) of our data set does not allow testing of this hypothesis. Second, supposedly due to the increased risk for cardiovascular diseases in ε4 carriers, *APOE* ε4 has been linked to increased mortality rates [39–41]. This observation fits our finding that *APOE* ε4 carriership also decreased with age in Aβ– CN and MCI subjects, although the reduction of *APOE* ε4 in Aβ– subjects can also be caused by individuals transitioning from Aβ– to

Table 2
Prevalence of *APOE* genotype in CN and MCI subjects according to Aβ status

Group	<i>APOE</i> ε2/ε2	<i>APOE</i> ε2/ε3	<i>APOE</i> ε2/ε4	<i>APOE</i> ε3/ε3	<i>APOE</i> ε3/ε4	<i>APOE</i> ε4/ε4	<i>APOE</i> ε2 carrier	<i>APOE</i> ε3 carrier	<i>APOE</i> ε4 carrier	Missing
Aβ+/- CN and MCI, n (%)	22 (0.4)	566 (9.4)	126 (2.1)	3028 (50.4)	1845 (30.7)	422 (7.0)	714 (11.9)	5565 (92.6)	6009 (37.7)	440 (6.8)
Aβ+ CN and MCI, n (%)	2 (0.1)	88 (3.7)	61 (2.6)	861 (36.0)	1027 (43.0)	350 (14.7)	151 (6.3)	2037 (85.3)	1377 (57.6)	209 (8.0)
Aβ- CN and MCI, n (%)	20 (0.6)	478 (13.2)	65 (1.8)	2167 (59.9)	818 (22.6)	72 (2.0)	563 (15.6)	3528 (97.5)	890 (24.6)	231 (6.0)
Aβ+ CN, n (%)	1 (0.1)	28 (3.7)	19 (2.5)	336 (44.7)	304 (40.5)	63 (8.4)	48 (6.4)	687 (91.5)	367 (48.9)	37 (4.7)
Aβ+ MCI, n (%)	1 (0.1)	60 (3.7)	42 (2.6)	525 (32.1)	723 (44.1)	287 (17.5)	103 (6.3)	1350 (82.4)	1010 (61.7)	172 (9.5)
Aβ- CN, n (%)	15 (0.7)	311 (14.1)	38 (1.7)	1331 (60.4)	478 (21.7)	31 (1.4)	364 (16.5)	2158 (97.9)	509 (23.1)	122 (5.2)
Aβ- MCI, n (%)	5 (0.4)	167 (11.8)	27 (1.9)	836 (59.0)	340 (24.0)	41 (2.9)	199 (14.1)	1370 (96.8)	381 (26.9)	109 (7.1)

Abbreviations: Aβ, amyloid β; CN, cognitively normal; MCI, mild cognitive impairment; *APOE*, apolipoprotein E.

NOTE. Information on *APOE* genotype was available in 93.2% of subjects with normal cognition and mild cognitive impairment. For subjects with AD dementia, only information on *APOE* status (+ or -) was provided.

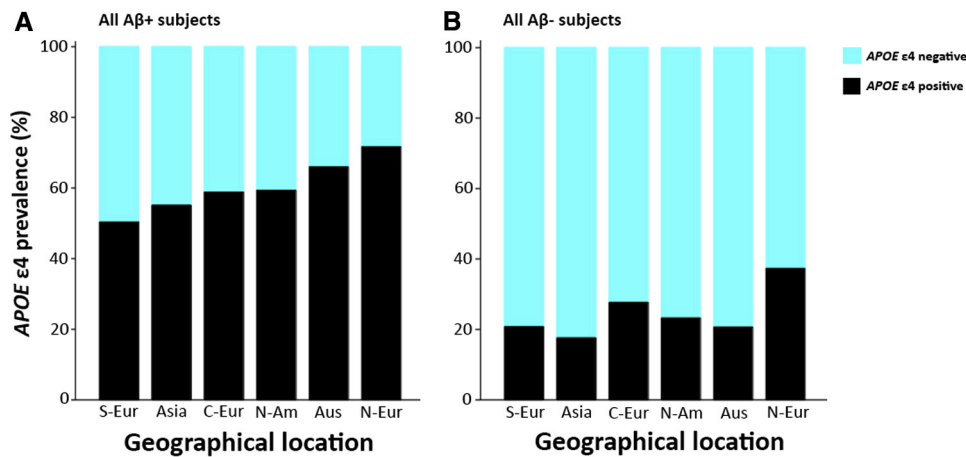


Fig. 3. Distribution of *APOE* $\epsilon 4$ negative and *APOE* $\epsilon 4$ positive subjects by geographical location for all $A\beta+$ (A) and $A\beta-$ (B) participants across diagnostic groups. A further breakdown into diagnostic groups is provided in [Supplementary Fig. S2](#); 8.1% of participants ($n = 637$ [303 $A\beta+$, 334 $A\beta-$]) could not be classified, as they were included in a multicenter study that covered multiple geographical locations. Abbreviations: $A\beta$, amyloid β ; *APOE*, apolipoprotein E.

$A\beta+$ with advancing age. Finally, as *APOE* $\epsilon 4$ accelerates the onset of amyloid aggregation by approximately 15 years [5,26], the prevalence of $\epsilon 4$ carriers in $A\beta+$ subjects will be higher at younger age ranges. Remarkably, the prevalence of *APOE* $\epsilon 4$ did not change with age in AD-type dementia. It may be hypothesized that the higher mortality in *APOE* $\epsilon 4$ carriers is counterbalanced at the dementia stage by individuals transitioning from preclinical and prodromal AD into AD dementia. We also tested whether this lack of an age effect was caused by the inclusion of atypical variants of AD dementia as this group is characterized by lower prevalence of *APOE* $\epsilon 4$ [42,43], but this was not the case ([Supplementary Fig. S2](#)). The pathogenesis of early onset AD is complex because this group includes a mix of *APOE* $\epsilon 4$ carriers who develop the disease at younger age and of *APOE* $\epsilon 4$ noncarriers with rapidly progressive AD [44,45]. This may confound relationships between *APOE* $\epsilon 4$ and age, especially in young patients with AD-type dementia. Furthermore, it has been shown that the mortality effect of *APOE* $\epsilon 4$ is less pronounced at older age [46], which may explain the lack of an age effect in AD dementia patients. It is not clear why $A\beta+$ women had decreasing prevalence of *APOE* $\epsilon 4$ with age. However, a recent large meta-analysis also found an interaction between *APOE* $\epsilon 4$, sex, and age, so that *APOE* $\epsilon 4$ conferred a greater risk for AD in women than in men at younger ages but not in older [47]. It is possible that physiological changes around menopause may interact with *APOE* $\epsilon 4$ in women and increase the risk for $A\beta$ pathology in younger ages [48]. If this leads to an earlier onset of the disease, and earlier death, the *APOE* $\epsilon 4$ prevalence may appear to decrease with age in $A\beta+$ women.

Another main finding was the lower prevalence of *APOE* $\epsilon 4$ in both $A\beta+$ and $A\beta-$ CN subjects compared with the MCI and dementia stages. This may be explained by a selection bias, as the vast majority of the MCI and AD dementia

subjects visited a memory clinic, while many CN subjects were recruited as research volunteers. Also, *APOE* $\epsilon 4$ + MCI patients may be more likely to seek medical help, and *APOE* $\epsilon 4$ carriers with dementia may be more willing to participate in research due to a positive family history. Another possible reason is that *APOE* $\epsilon 4$ may accelerate the transition from preclinical to clinical AD. For example, *APOE* $\epsilon 4$ may have an effect on brain structure and function through non- $A\beta$ pathways [49–53], which may act synergistically with $A\beta$ pathology to shorten the time between the start of $A\beta$ deposition and cognitive decline. Thus, because *APOE* $\epsilon 4$ carriers will develop symptoms earlier, the prevalence of *APOE* $\epsilon 4$ positivity in CN is lower than that in MCI and dementia cases at the same age range. Finally, *APOE* $\epsilon 4$ noncarriers (which would include *APOE* $\epsilon 2$ carriers) may have mechanisms of resilience (i.e., cognitive reserve) that are less present in $\epsilon 4$ carriers [54].

We also found geographical differences in *APOE* $\epsilon 4$ prevalence, with higher prevalence in AD patients from Northern Europe, Central Europe, and Australia and lower prevalence in patients from Southern Europe and Asia. This is consistent with previous epidemiological studies in clinically diagnosed AD dementia and MCI patients [13,55] and with lower prevalence of *APOE* $\epsilon 4$ in the general population in Southern Europe and Asia compared with Northern Europe [14,55–57]. The novelty of this study is that we confirm these geographical differences in $A\beta$ biomarker-defined AD and throughout the continuum from preclinical to prodromal and dementia stages. The different geographical prevalence of *APOE* $\epsilon 4$ may be important for recruitment of participants in clinical trials and for the use of *APOE* $\epsilon 4$ in algorithms to predict $A\beta$ positivity [58].

Strengths of this study include the large number of $A\beta$ -positive subjects across the spectrum from preclinical to prodromal and dementia stages of AD. Limitations include

relatively few participants who came from Asia ($n = 315$) and Australia ($n = 394$), and there were no participants from Africa and South America. There were no data on ethnicity of the participants, which may confound the results because ethnicity has been related to both *APOE* $\epsilon 4$ and AD [14,59]. Also, this study is based on an assembly of different study cohorts that may not be representative for typical memory clinic populations or the general population. Finally, $A\beta$ positivity was determined using different modalities (i.e., PET or CSF) and methods (e.g., visual read vs. quantitative threshold for PET and different assays for CSF). There was an unexpected effect of CSF assay (Innotest vs. Luminex), which could be interpreted as a cohort effect as the majority of subjects with CSF analyzed using the Luminex assay are ADNI participants (Supplementary Table 5). We found no effects of modality (PET vs. CSF) on *APOE* $\epsilon 4$ prevalence, and in previous studies using these data, we found only little evidence for heterogeneity related to modality and methodology [5,30].

With about 2/3 of prodromal AD and AD dementia patients being *APOE* $\epsilon 4$ carriers, our results further emphasize the importance of *APOE* $\epsilon 4$ for the development of AD [8,9]. This may be useful for the development of disease-modifying treatments, which may be focused on attenuating the detrimental effects of *APOE* $\epsilon 4$ and for understanding the molecular pathogenesis of AD [60]. Furthermore, the finding that the prevalence of *APOE* $\epsilon 4$ decreases with age in CN and MCI subjects has potential implications for clinical trials in prodementia populations, as screening based on *APOE* status to enrich for $A\beta$ positivity may be less effective with advancing age. Finally, it may be of importance to evaluate other proposed AD susceptibility genes [61] in cohorts with known $A\beta$ status, as to date, this has only been assessed in cohorts of clinically diagnosed AD patients and CN elderly.

5. Conclusions

We have quantified the prevalence of *APOE* $\epsilon 4$ in $A\beta$ biomarker-defined preclinical AD, prodromal AD, and AD dementia. The results emphasize the prominent role of *APOE* $\epsilon 4$ in AD, but also point to disease heterogeneity, because *APOE* $\epsilon 4$ positivity is markedly less common in elderly subjects in prodementia stages of AD and in people from specific geographical locations, including Southern Europe and Asia. Further studies on phenotypic differences between *APOE* $\epsilon 4$ -negative and *APOE* $\epsilon 4$ -positive AD patients may be important to understand different pathways that may lead to AD and ultimately to tailor disease-modifying treatments to specific patient subgroups.

Acknowledgments

H.H. is supported by the AXA Research Fund, the “Fondation Université Pierre et Marie Curie”, and the “Fondation

pour la Recherche sur Alzheimer”, Paris, France. Ce travail a bénéficié d’une aide de l’Etat “Investissements d’avenir” ANR-10-IAIHU-06. The research leading to these results has received funding from the program “Investissements d’avenir” ANR-10-IAIHU-06 (Agence Nationale de la Recherche-10-IA Agence Institut Hospitalo-Universitaire-6). W.E.K. is supported by the National Institutes of Health grants: P50 AG005133, RF1 AG025516, and PO1 AG025204.

R.O. is supported by Marie Curie FP7 International Outgoing Fellowship [628812] and the donors of [Alzheimer’s Disease Research], a program of the BrightFocus Foundation.

P.S.-J. received grants from Instituto de Salud Carlos III (Fondo de Investigación Sanitario, PI08/0139, PI12/02288, PI16/01652, and the CIBERNED program).

Disclosures: D.A. reported having received research support or honoraria from Astra-Zeneca, H. Lundbeck, Novartis Pharmaceuticals, and GE Health. A.W. reported having received speakers’ bureau fees from Esai and Triolab and serving on the advisory board for Nutricia and Esai. K.B. reported having received personal fees (advisory boards or consulting) from Roche Diagnostics, IBL International, Novartis, Fujirebio Europe, and Eli Lilly and is a co-founder of Brain Biomarker Solutions in Gothenburg AB, a GU Venture-based platform company at the University of Gothenburg. K.C. reported having received grants from the National Institutes of Health (NIH). A.D. reported having received speaker honoraria and consulting fees from GE Healthcare, AVID/Lilly, and Piramal. A.M.F. reported having received grants from the NIH, Fred Simmons and Olga Mohan, and Charles and Joanne Knight Alzheimer’s Research Initiative of the Washington University Knight Alzheimer’s Disease Research Center; having received personal fees (advisory boards or consulting) from IBL International, Roche Diagnostic, Diami R, and AbbVie. T.F. reported having a patent “Methods and compositions for monitoring phagocytic activity,” PCT/US2011/062233, pending. A.S.F. reported having been a full-time employee of the Banner Alzheimer’s Institute at the time of data collection; currently being a full-time employee of Eli Lilly. S.F. reported having received personal fees (consultancy) from Piramal, Bayer, and GE. G.B.F. reported having received grants and/or personal fees from Lilly, Bristol-Myers Squibb, Bayer, Lundbeck, Elan, AstraZeneca, Pfizer, Taurx, Wyeth, GE, Baxter, Avid, Roche, Piramal, and the Alzheimer’s Association. K.D.G. reported having received grants from the Indian Council of Medical Research, New Delhi, India. T.G. reported having received consulting fees from Actelion, Eli Lilly, MSD, Novartis, Quintiles, and Roche Pharma; lecture fees from Biogen, Lilly, Parexel, and Roche Pharma; and grants for his institution from Actelion and PreDemTech. H.H. declares no conflict of interest with the content of the present manuscript. He serves as a Senior Associate Editor for the Journal *Alzheimer’s & Dementia*; he has been a scientific consultant and/or speaker and/or attended scientific advisory boards of Axovant, Anavex, Eli

Lilly and company, GE Healthcare, Cytos Ltd, Jung Diagnostics GmbH, Roche, Biogen Idec, Takeda-Zinfandel, Oryzon Genomics, and Qynapse; and he receives research support from the Association for Alzheimer Research (Paris), Pierre and Marie Curie University (Paris), and Pfizer & Avid (paid to institution); and he has patents but receives no royalties. O.H. has received research support (to the institute) from GE Healthcare, AVID radiopharmaceuticals, and Hoffmann-La Roche. W.J. reported having received personal fees from Banner Alzheimer Institute/Genentech, Synarc, Biogen, and Novartis. A.I. reported having served on an advisory board for Eli Lilly and Nutricia, having received compensation as a speaker and consultant for GE Healthcare and Nutricia, having received clinical trial agreements with GEHC, Merck, and Eli Lilly, having received grants from the Fonds de la Recherche Scientifique (F.R.S.—FNRS), Belgium and nonfinancial support from GEHC. W.J.J. reported having received research support from Biogen. W.E.K. reported being a co-inventor of the amyloid imaging tracer PiB and, as such, having a financial interest in the license agreement. (PiB intellectual property is owned by the University of Pittsburgh, and GE Healthcare holds a license agreement with the University of Pittsburgh based on the PiB technology described in this article and receives “inventors share” payments from the University of Pittsburgh based on income from that license). J.K. reported having received grants from the German Federal Ministry of Education and Research (BMBF): Kompetenznetz Demenzen (01GI0420) and the German Federal Ministry of Education and Research (BMBF): The Frontotemporal Lobar Degeneration Consortium (FTDL-C), 01GI1007 A and having a patent, PCT/EP2004/003963, “Diagnosis of Alzheimer’s disease,” issued; a patent, EP 1811304 A1, “Large A β -peptide binding particles (LAPS) in diagnosis and therapy of Alzheimer’s dementia,” issued; a patent, WO2007/082750 A1, “Immunoglobulin-bound Ab-peptides and immunoglobulins-binding Ab-peptides in diagnosis and therapy of Alzheimer’s dementia,” issued; a patent, EP 2437067A2, “Methods of differentially diagnosing dementias,” issued; and a patent, “New formulations for diagnosis of Alzheimer’s disease,” pending. S.L. reported having received grants from NIH and personal fees from Biogen Idec, Genentech, and Synarc. A.L. reported having received grants from Instituto de Salud Carlos III (Fondo de Investigación Sanitario, PI10/01,878; PI13/01,532; PI11/2425; PI11/3035 and the CIBERNED program). M.M. reported being an employee of Avid Radiopharmaceuticals, a wholly owned subsidiary of Eli Lilly. J.C.M. reported having received grants from NIH (P50AG005681, P01AG003991, P01AG026276, and U19AG032438). B.M. reported having received grants and personal fees from the Leading National Research Centre (KNOW), Medical University of Bialystok, Poland; and consultation and/or lecture honoraria from Roche, Cormay, and Biameditek. O.P. reported having received grants and/or personal fees from Lilly, Roche, Genentech, Lundbeck,

Affris, Piramal, Novartis, and Trx-Pharmaceuticals. J.P. reported having received grants from the Swiss National Science Foundation (SNF 320030L_141179), Fujirebio Europe, and from the Nestlé Institute of Health Sciences. G.D.R. reported having received grants from Avid Radiopharmaceuticals and personal fees from GE Healthcare and Piramal. J.O.R. reported having received grants from Sigrid Juselius Foundation and Turku University Hospital clinical grants. C.C.R. reported having received grants from Avid Radiopharmaceuticals, Piramal Imaging, AstraZeneca, GE Healthcare, Avid/Lilly, Navidea, CSIRO, NHMRC, Alzheimer’s Association, and an anonymous foundation and having had a patent licensed for PET image processing. M.S. reported having received personal fees from Eisai, Janssen, Novartis (lecture), and Allianz (lecture) and research grants from the French Health Ministry, Institute Roche de Recherche et Médecine Translationnelle (paid to the institution). P.S. reported having received grants from GE Healthcare, Piramal, and Merck, paid to his institution. H.S. reported having received grants from the Academy of Finland, European Union 7ThFP 601055 VPH-DARE, Kuopio University Hospital VTR, and University of Eastern Finland. C.E.T. reported being a member of the international advisory board at Innogenetics and Roche; and having research contracts at Probiobio, Boehringer, Roche, EIP Pharma, Brainsonline, Axon Neurosciences, and PeopleBio. W.M.v.d.F. reported having received grants from Boehringer Ingelheim, Piramal Imaging, and Roche. K.V.L. reported having received grants through KU Leuven from Merck, Janssen Pharmaceuticals, UCB, Novartis, Pfizer, and GE Healthcare. R.V. reported having received clinical trial agreements with GEHC, Merck, Forum, and Roche; grants from Research Foundation—Flanders (FWO) and KU Leuven; and nonfinancial support from GEHC. M.M.V. reported having served on an advisory board for Roche. F.R.J.V. reported having received compensation as a speaker and consultant for Nutricia Advanced Medical Food. P.J.V. reported having received research support from Biogen and grants from EU/EFPIA Innovative Medicines Initiative Joint Undertaking, EU Joint Programme—Neurodegenerative Disease Research (JPND), ZonMw, and Bristol-Myers Squibb; having served as member of the advisory board of Roche Diagnostics; and having received nonfinancial support from GE Healthcare. S.J.B.V. receives research support from Janssen Pharmaceutica N.V. and grants from ZonMw and EU/EFPIA Innovative Medicines Initiative Joint Undertaking. G.W. reported being a board member of the Lundbeck Foundation. D.A.W. reported having received personal fees from GE Healthcare and Piramal Pharma and grants from Avid Radiopharmaceuticals. H.Z. is a co-founder of Brain Biomarker Solutions in Gothenburg AB, a GU Venture-based platform company at the University of Gothenburg. The authors received compensation (i.e., salary) as employees of their respective organizations. No other disclosures were reported.

Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.jalz.2018.02.009>.

RESEARCH IN CONTEXT

1. Systematic review: Previous studies examining the prevalence of apolipoprotein E (*APOE*) $\epsilon 4$ in Alzheimer's disease have included patients based on clinical criteria, without using biomarker information. This may have led to an underestimation of the prevalence of *APOE* $\epsilon 4$ due to misdiagnosis.
2. Interpretation: Our results demonstrate that positron emission tomography or cerebrospinal fluid evidence for the presence of amyloid β is associated with a higher prevalence of *APOE* $\epsilon 4$ (66% vs. 50–60 in previous studies).
3. Future directions: Information on *APOE* $\epsilon 4$ status would improve algorithms to determine risk for amyloid β positivity, for example, to enrich clinical trials. Furthermore, similar studies in amyloid β positive subjects should be performed to determine the prevalence of other Alzheimer's disease susceptibility genes.

References

- [1] Scheltens P, Blennow K, Breteler MM, de Strooper B, Frisoni GB, Salloway S, et al. Alzheimer's disease. *Lancet* 2016;388:505–17.
- [2] Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. *Science* 1992;256:184–5.
- [3] Jack CR Jr, Holtzman DM. Biomarker modeling of Alzheimer's disease. *Neuron* 2013;80:1347–58.
- [4] Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC, et al. Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N Engl J Med* 2012;367:795–804.
- [5] Jansen WJ, Ossenkoppele R, Knol DL, Tijms BM, Scheltens P, Verhey FR, et al. Prevalence of cerebral amyloid pathology in persons without dementia: a meta-analysis. *JAMA* 2015;313:1924–38.
- [6] Jack CR Jr, Thorneau TM, Wiste HJ, Weigand SD, Knopman DS, Lowe VJ, et al. Transition rates between amyloid and neurodegeneration biomarker states and to dementia: a population-based, longitudinal cohort study. *Lancet Neurol* 2016;15:56–64.
- [7] Livingston G, Sommerlad A, Orgeta V, Costafreda SG, Huntley J, Ames D, et al. Dementia prevention, intervention, and Care. *Lancet* 2017;390:2673–734.
- [8] Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 1993;261:921–3.
- [9] Poirier J, Davignon J, Bouthillier D, Kogan S, Bertrand P, Gauthier S. Apolipoprotein E polymorphism and Alzheimer's disease. *Lancet* 1993;342:697–9.
- [10] Mahley RW. Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* 1988;240:622–30.
- [11] Raichlen DA, Alexander GE. Exercise, APOE genotype, and the evolution of the human lifespan. *Trends Neurosci* 2014;37:247–55.
- [12] Ringman JM, Coppola G. New genes and new insights from old genes: update on Alzheimer disease. *Continuum (Minneapolis Minn)* 2013;19:358–71.
- [13] Ward A, Crean S, Mercaldi CJ, Collins JM, Boyd D, Cook MN, et al. Prevalence of apolipoprotein E4 genotype and homozygotes (*APOE* $\epsilon 4/\epsilon 4$) among patients diagnosed with Alzheimer's disease: a systematic review and meta-analysis. *Neuroepidemiology* 2012;38:1–17.
- [14] Corbo RM, Scacchi R. Apolipoprotein E (*APOE*) allele distribution in the world. Is *APOE**4 a 'thrifty' allele? *Ann Hum Genet* 1999;63:301–10.
- [15] Beach TG, Monsell SE, Phillips LE, Kukull W. Accuracy of the clinical diagnosis of Alzheimer disease at National Institute on Aging Alzheimer Disease Centers, 2005–2010. *J Neuropathol Exp Neurol* 2012;71:266–73.
- [16] Ossenkoppele R, Prins ND, Pijnenburg YA, Lemstra AW, van der Flier WM, Adriaanse SF, et al. Impact of molecular imaging on the diagnostic process in a memory clinic. *Alzheimers Dement* 2013;9:414–21.
- [17] Palmqvist S, Zetterberg H, Mattsson N, Johansson P, Alzheimer's Disease Neuroimaging Initiative, Minthon L, Blennow K, et al. Detailed comparison of amyloid PET and CSF biomarkers for identifying early Alzheimer disease. *Neurology* 2015;85:1240–9.
- [18] Chetelat G, Ossenkoppele R, Villemagne VL, Perrotin A, Landeau B, Mezenge F, et al. Atrophy, hypometabolism and clinical trajectories in patients with amyloid-negative Alzheimer's disease. *Brain* 2016;139:2528–39.
- [19] Salloway S, Sperling R, Fox NC, Blennow K, Klunk W, Raskind M, et al. Two phase 3 trials of bapineuzumab in mild-to-moderate Alzheimer's disease. *N Engl J Med* 2014;370:322–33.
- [20] Andreasson U, Lautner R, Schott JM, Mattsson N, Hansson O, Herukka SK, et al. CSF biomarkers for Alzheimer's pathology and the effect size of *APOE* varepsilon4. *Mol Psychiatry* 2014;19:148–9.
- [21] Corneveaux JJ, Myers AJ, Allen AN, Pruzin JJ, Ramirez M, Engel A, et al. Association of CR1, CLU and PICALM with Alzheimer's disease in a cohort of clinically characterized and neuropathologically verified individuals. *Hum Mol Genet* 2010;19:3295–301.
- [22] Dubois B, Feldman HH, Jacova C, Hampel H, Molinuevo JL, Blennow K, et al. Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurol* 2014;13:614–29.
- [23] Albert MS, DeKosky ST, Dickson D, Dubois B, Feldman HH, Fox NC, et al. The diagnosis of mild cognitive impairment due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* 2011;7:270–9.
- [24] Sperling RA, Aisen PS, Beckett LA, Bennett DA, Craft S, Fagan AM, et al. Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* 2011;7:280–92.
- [25] Louwersheimer E, Wolfsgruber S, Espinosa A, Lacour A, Heilmann-Heimbach S, Alegret M, et al. Alzheimer's disease risk variants modulate endophenotypes in mild cognitive impairment. *Alzheimers Dement* 2016;12:872–81.
- [26] Jack CR Jr, Wiste HJ, Weigand SD, Rocca WA, Knopman DS, Mielke MM, et al. Age-specific population frequencies of cerebral beta-amyloidosis and neurodegeneration among people with normal cognitive function aged 50–89 years: a cross-sectional study. *Lancet Neurol* 2014;13:997–1005.
- [27] Morris JC, Roe CM, Xiong C, Fagan AM, Goate AM, Holtzman DM, et al. *APOE* predicts amyloid-beta but not tau Alzheimer pathology in cognitively normal aging. *Ann Neurol* 2010;67:122–31.

- [28] Papp KV, Rentz DM, Mormino EC, Schultz AP, Amariglio RE, Quiroz Y, et al. Cued memory decline in biomarker-defined preclinical Alzheimer disease. *Neurology* 2017;88:1431–8.
- [29] Vos SJ, Xiong C, Visser PJ, Jasielec MS, Hassenstab J, Grant EA, et al. Preclinical Alzheimer's disease and its outcome: a longitudinal cohort study. *Lancet Neurol* 2013;12:957–65.
- [30] Ossenkoppele R, Jansen WJ, Rabinovici GD, Knol DL, van der Flier WM, van Berckel BN, et al. Prevalence of amyloid PET positivity in dementia syndromes: a meta-analysis. *JAMA* 2015;313:1939–49.
- [31] Jansen WJ, Ossenkoppele R, Tijms BM, Fagan AM, Hansson O, Klunk WE, et al. Association of cerebral amyloid-beta aggregation with cognitive functioning in persons without dementia. *JAMA Psychiatry* 2018;75:84–95.
- [32] Jessen F, Amariglio RE, van Boxtel M, Breteler M, Ceccaldi M, Chetelat G, et al. A conceptual framework for research on subjective cognitive decline in preclinical Alzheimer's disease. *Alzheimers Dement* 2014;10:844–52.
- [33] Jansen WJ, Ossenkoppele R, Visser PJ. Amyloid pathology, cognitive impairment, and Alzheimer disease risk—reply. *JAMA* 2015;314:1177–8.
- [34] McKhann GM, Knopman DS, Chertkow H, Hyman BT, Jack CR Jr, Kawas CH, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement* 2011;7:263–9.
- [35] Zwan M, van Harten A, Ossenkoppele R, Bouwman F, Teunissen C, Adriaanse S, et al. Concordance between cerebrospinal fluid biomarkers and [11C]PIB PET in a memory clinic cohort. *J Alzheimers Dis* 2014;41:801–7.
- [36] Landau SM, Lu M, Joshi AD, Pontecorvo M, Mintun MA, Trojanowski JQ, et al. Comparing positron emission tomography imaging and cerebrospinal fluid measurements of beta-amyloid. *Ann Neurol* 2013;74:826–36.
- [37] Donohue MC, Sperling RA, Petersen R, Sun CK, Weiner MW, Aisen PS, et al. Association between elevated brain amyloid and subsequent cognitive decline among cognitively normal persons. *JAMA* 2017;317:2305–16.
- [38] Lim YY, Mormino EC. Alzheimer's Disease Neuroimaging I. APOE genotype and early beta-amyloid accumulation in older adults without dementia. *Neurology* 2017;89:1028–34.
- [39] Tilvis RS, Strandberg TE, Juva K. Apolipoprotein E phenotypes, dementia and mortality in a prospective population sample. *J Am Geriatr Soc* 1998;46:712–5.
- [40] Wang X, Lopez O, Sweet RA, Becker JT, DeKosky ST, Barmada MM, et al. Genetic determinants of survival in patients with Alzheimer's disease. *J Alzheimers Dis* 2015;45:651–8.
- [41] Beydoun MA, Beydoun HA, Kaufman JS, An Y, Resnick SM, O'Brien R, et al. Apolipoprotein E epsilon4 allele interacts with sex and cognitive status to influence all-cause and cause-specific mortality in U.S. older adults. *J Am Geriatr Soc* 2013;61:525–34.
- [42] Ossenkoppele R, Mattsson N, Teunissen CE, Barkhof F, Pijnenburg Y, Scheltens P, et al. Cerebrospinal fluid biomarkers and cerebral atrophy in distinct clinical variants of probable Alzheimer's disease. *Neurobiol Aging* 2015;36:2340–7.
- [43] van der Flier WM, Pijnenburg YA, Fox NC, Scheltens P. Early-onset versus late-onset Alzheimer's disease: the case of the missing APOE varepsilon4 allele. *Lancet Neurol* 2011;10:280–8.
- [44] Cohen ML, Kim C, Haldiman T, ElHag M, Mehndiratta P, Pichet T, et al. Rapidly progressive Alzheimer's disease features distinct structures of amyloid-beta. *Brain* 2015;138:1009–22.
- [45] Ossenkoppele R, van der Flier WM, Zwan MD, Adriaanse SF, Boellaard R, Windhorst AD, et al. Differential effect of APOE genotype on amyloid load and glucose metabolism in AD dementia. *Neurology* 2013;80:359–65.
- [46] Corrada MM, Paganini-Hill A, Berlau DJ, Kawas CH. Apolipoprotein E genotype, dementia, and mortality in the oldest old: the 90+ Study. *Alzheimers Dement* 2013;9:12–8.
- [47] Neu SC, Pa J, Kukull W, Beekly D, Kuzma A, Gangadharan P, et al. Apolipoprotein E genotype and sex risk factors for Alzheimer disease: a meta-analysis. *JAMA Neurol* 2017;74:1178–89.
- [48] Riedel BC, Thompson PM, Brinton RD. Age, APOE and sex: Triad of risk of Alzheimer's disease. *J Steroid Biochem Mol Biol* 2016;160:134–47.
- [49] Dumanis SB, Tesoriero JA, Babus LW, Nguyen MT, Trotter JH, Ladu MJ, et al. ApoE4 decreases spine density and dendritic complexity in cortical neurons in vivo. *J Neurosci* 2009;29:15317–22.
- [50] Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. *Neuron* 2009;63:287–303.
- [51] Liu Y, Yu JT, Wang HF, Han PR, Tan CC, Wang C, et al. APOE genotype and neuroimaging markers of Alzheimer's disease: systematic review and meta-analysis. *J Neurol Neurosurg Psychiatry* 2015;86:127–34.
- [52] Jagust WJ, Landau SMA. Alzheimer's Disease Neuroimaging I. Apolipoprotein E, not fibrillar beta-amyloid, reduces cerebral glucose metabolism in normal aging. *J Neurosci* 2012;32:18227–33.
- [53] Reiman EM, Chen K, Alexander GE, Caselli RJ, Bandy D, Osborne D, et al. Functional brain abnormalities in young adults at genetic risk for late-onset Alzheimer's dementia. *Proc Natl Acad Sci U S A* 2004;101:284–9.
- [54] Suri S, Heise V, Trachtenberg AJ, Mackay CE. The forgotten APOE allele: a review of the evidence and suggested mechanisms for the protective effect of APOE varepsilon2. *Neurosci Biobehav Rev* 2013;37:2878–86.
- [55] Norberg J, Graff C, Almkvist O, Ewers M, Frisoni GB, Frolich L, et al. Regional differences in effects of APOE epsilon4 on cognitive impairment in non-demented subjects. *Dement Geriatr Cogn Disord* 2011;32:135–42.
- [56] Arboleda GH, Yunis JJ, Pardo R, Gomez CM, Hedmont D, Arango G, et al. Apolipoprotein E genotyping in a sample of Colombian patients with Alzheimer's disease. *Neurosci Lett* 2001;305:135–8.
- [57] Kern S, Mehlig K, Kern J, Zetterberg H, Thelle D, Skoog I, et al. The distribution of apolipoprotein E genotype over the adult lifespan and in relation to country of birth. *Am J Epidemiol* 2015;181:214–7.
- [58] Insel PS, Palmqvist S, Mackin RS, Nosheny RL, Hansson O, Weiner MW, et al. Assessing risk for preclinical beta-amyloid pathology with APOE, cognitive, and demographic information. *Alzheimers Dement (Amst)* 2016;4:76–84.
- [59] Farrer LA, Cupples LA, Haines JL, Hyman B, Kukull WA, Mayeux R, et al. Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. *JAMA* 1997;278:1349–56.
- [60] Mattsson N, Insel PS, Palmqvist S, Stomrud E, van Westen D, Minthon L, et al. Increased amyloidogenic APP processing in APOE varepsilon4-negative individuals with cerebral beta-amyloidosis. *Nat Commun* 2016;7:10918.
- [61] Karch CM, Goate AM. Alzheimer's disease risk genes and mechanisms of disease pathogenesis. *Biol Psychiatry* 2015;77:43–51.