1	For: Journal of Economic Entomology Corr	responding author:
2	2 (Household and Structural Insects section)	
3	3 A	Chow-Yang Lee
4	4 Urban En	tomology Laboratory
5	5 Vector C	f Diological Sciences
0		arsiti Saine Malaysia
2 2	8 118	00 Penang Malaysia
9	9 Email:	chowyang@usm.mv
10	0	
11	1	
12	2	
13	3 Differences in climbing ability of <i>Cimex lectulariu</i>	s and Cimex
11	homintorus (Hemintera: Cimicidae)	
14	⁴ <i>nempterus</i> (Heimpteru: Chinetuae)	
16	6	
17	DAE-YUN KIM ¹ , JOHAN BILLEN ² , STEPHEN L. DOGGETT ³ AND C	CHOW-YANG
18	8 $LEE^{1,4}$	
19	9	
20	¹ Urban Entomology Laboratory, Vector Control Research Unit, School o	f Biological Sciences,
21	1 Universiti Sains Malaysia, 11800 Penang, Malaysia.	
22		
23	³ ⁻ KU Leuven, Zoological Institute, Naamsestraat 59, box 2466, B-3000 Le	euven, Belgium.
24	4	
25	³ Department of Medical Entomology, Westmead Hospital, Westmead NS	W 2145, Australia.
26	6 ⁴ Corresponding author. Email: chowyang@usm.my	
27	7	
28	8	
20	0	
29	9 Running title: Kim et al. Climbing ability of <i>Cimex lectularius</i> and <i>C</i> .	hemipterus
30	0	
31	1	
51	÷	

ABSTRACT The climbing abilities of two bed bug species, *Cimex lectularius* L. and *Cimex* 32 33 hemipterus (F.) were determined by evaluating their escape rates from smooth surface pitfall traps using four commercial bed bug monitors (Verifi™ Bed Bug Detector, ClimbUp® Insect 34 Interceptor, BlackOutTM Bed Bug Detector, and SenSci VolcanoTM Bed Bug Detector). All 35 detectors were used in the absence of lures or attractants. Unlike C. lectularius, adult C. 36 hemipterus were able to escape from all traps. On the other hand, no or low number nymphs 37 of both species escaped, depending on the evaluated traps. Examination on the vertical 38 friction force of adults of both species revealed a higher vertical friction force in C. 39 hemipterus than in C. lectularius. Scanning electron micrograph observation on the tibial pad 40 41 of adult bed bugs of C. hemipterus showed the presence of a greater number of tenent hairs on 42 the tibial pad than on that of adult C. lectularius. No tibial pad was found in the fourth and fifth instars of both species. Near the base of the hollow tenent hairs is a glandular epithelium 43 44 that is better developed in adult C. hemipterus than in adult C. lectularius. This study highlights significant morphological differences between C. lectularius and C. hemipterus, 45 which has profound implications in the monitoring and potential management of bed bug 46 infestations. 47

Key Words bed bug, tenent hair, vertical friction force, climbing, pitfall trap, monitoring,
management.

50

Introduction

The current global resurgence of bed bugs involves two species, namely the common bed bug, *Cimex lectularius* L. and the tropical bed bug, *Cimex hemipterus* (F.). *Cimex lectularius* is mainly found in the temperate and subtropical regions, while the latter species is distributed in the subtropical and tropical regions (Omori 1939, 1941, Usinger 1966, Doggett et al. 2003, How and Lee 2010, Zulaikha et al. 2016). In some regions such as in Africa (Newberry 1988,

1989), Australia (Doggett et al. 2003), Florida (Hixson 1943, Campbell et al. 2016), and
Taiwan (Lee 2013), both species can co-exist sympatrically.

58

The detection and monitoring of bed bugs is one of the most important aspects of integrated 59 pest management (IPM) against this insect (Wang & Copper 2011). Detection and monitoring 60 of bed bugs can be undertaken via visual inspections, through various traps and monitoring 61 62 devices, via canine detection (Pinto et al. 2007, Wang & Copper 2011, Boase & Naylor 2014), or any combination of these. Amongst the monitors available in the market are pitfall and 63 sticky traps, some with the addition of lures such as heat, carbon dioxide, and/or insect/host 64 65 odors (active monitors), while others contain no lure (passive monitors) and provide a harborage for bed bugs to enter. While many such products exist on the market, few have 66 been tested for efficacy via independent scientific tests. 67

68

Pitfall style traps of different designs have been found effective in detecting and monitoring C. 69 70 lectularius (Singh et al. 2013). These traps rely on a smooth inner wall surface to prevent trapped bed bugs from escaping. Such traps with smooth surfaces can also be used as a barrier 71 72 to prevent bed bugs from reaching the sleeping hosts (Doggett 2013). However, if the trapped 73 bugs were able to climb up the smooth surfaces and escape, this would seriously compromise monitoring efforts or their use as barriers. It is known that a buildup of dust and debris can 74 compromise the efficacy of pitfall traps against C. lectularius and hence maintenance 75 76 recommendations are usually included such as regular cleaning or the addition of talc (Doggett 2013). 77

78

Most of the trap products were originally developed and tested in the U.S. where C. *lectularius* is the major bed bug species. Many of these products have subsequently been

marketed in other parts of the world on the assumption that they also would work against the 81 82 species that were found there, such as C. hemipterus. In a preliminary laboratory investigation though, it was observed that adult C. hemipterus could escape from a new pitfall trap, which 83 otherwise effectively contained C. lectularius. This raised an important question: are all pitfall 84 traps that are effective in containing C. lectularius also effective against C. hemipterus? If 85 differences are found, this would have profound implications for the monitoring of the latter 86 species and for the potential use of barriers against C. hemipterus. Furthermore, it would raise 87 the question why such differences exist. The results of the initial investigation prompted the 88 following study. The escape rate was compared between C. lectularius and C. hemipterus in 89 90 four commercially available monitors that were previously or presently marketed for bed bug 91 detection and monitoring in the US. Following this, the vertical friction force was determined for both species on two types of surfaces (smooth and rough). Lastly, the leg morphology was 92 investigated to determine the possible reasons behind the differences in climbing ability of 93 both species. 94

95

96

Materials and Methods

Insects. Two strains for each species were used; C. lectularius (Monheim [MH] and Sydney 97 [SYD] strains), and C. hemipterus (Kuala Lumpur [KL] and Green lane [GL] strains) (Table 98 1). All strains were reared in the laboratory in glass jars (7 cm diameter x 9 cm height) and 99 provided with folded brown paper as harborage under environmental conditions of $26 \pm 2^{\circ}$ C, 100 $70 \pm 5\%$ RH, and a 12-h photo period. All the insects used in this study were blood-fed on a 101 human volunteer once a week. An alcohol-preserved sample of adult C. hemipterus collected 102 103 from Papua New Guinea in 1932 [PNG strain] was used in part of the scanning electron microscope (SEM) observation to examine if any morphological change could have been a 104 recent evolutionary adaptation due to human management interventions. 105

107 Trap escape test. The insects were evaluated in their ability to escape from four bed bug pitfall traps namely, VerifiTM bed bug detector (FMC Professional Solutions, Philadelphia, 108 109 PA), ClimbUp® insect interceptor (Susan McKnight Inc, Memphis, TN) (without the talc), BlackOut[™] bed bug detector (Protect-A-Bed, Wheeling, IL), and SenSci Volcano[™] bed bug 110 detector (Bedbug Central, Lawrenceville, NJ). All traps were cleaned with 70% ethanol and 111 dried in a 60°C incubator overnight before the test. While ClimbUp® traps are normally used 112 with the talc (as per manufacturer recommendation), they were evaluated in the absence of 113 talc in this study. Lures are available for use in conjunction with the Verifi and Volcano traps 114 115 to enhance their collection efficacy but they were not used in this study. One strain of each 116 species was tested: C. lectularius (SYD strain) and C. hemipterus (KL strain). The evaluation was carried out by introducing 5 adult males, 5 adult females and 5 fourth to fifth instars into 117 the trap. The tested insects were placed in the pitfall for both Verifi and Volcano traps, and in 118 the outer pitfall for the ClimbUp and Blackout traps. The traps were placed inside test arenas 119 (50 length x 30 cm width x 10 cm height) and kept in an environmental chamber (Binder 120 Model KBF 240, Binder GmbH, Tuttlingen, Germany) under conditions of $26 \pm 1^{\circ}$ C, $70 \pm 1^{\circ}$ 121 RH, and 24-h scotophase. All these insects were blood-fed 5 days prior to the test. The 122 123 experiments began at 2000 hour, and at 4, 24, 48 and 72 hour post-treatment, the number of 124 insects that remained in the trap, and the number that escaped were counted. Observations were made up to 72 hours for all traps except for the Volcano trap which was observed daily 125 126 for up to 10 days post-treatment. All experiments were replicated three times.

128 Vertical friction force measurement. To confirm that the differences in climbing ability of129 the insects within the pitfall traps was due to better holding ability on the smooth surface of

one species over the other, the vertical friction forces of both C. lectularius and C. hemipterus 130 were measured with a Sartorius ED224S (Sartorius, Göttingen, Germany) electronic balance 131 using modified methods described by Betz (2002) and Hottel et al. (2015). A data recording 132 program, Sartorius Weight Anchor Software (Sartorius, Göttingen, Germany) installed on a 133 computer, was connected to the electronic balance using a Sartorius RS232 cable (Sartorius, 134 Göttingen, Germany). A bed bug was tethered to a polyethylene bristle by gluing (UHU 135 Power Glue®, UHU GmbH & Co. KG, Bühl, Germany) its dorsal side to one tip of the bristle. 136 The other tip of the bristle was inserted into a 30-g ball of modelling clay (Tack-it, A.W. 137 138 Faber-Castell (India) Ltd, Mumbai, India). The clay ball, which was attached to the bristle, was placed on the balance with the tethered bed bug over the other end, and tared to zero. 139

140

141 Two surfaces were tested, namely a smooth surface (glass microscope cover slide), and a rough surface (filter paper). The test surface was moved to the tethered bed bug to permit the 142 insect's legs to come into contact. Once the insect had contacted and gripped onto the test 143 surface, the software started recording the changes in the mass of the clay ball for 5 min. The 144 data was then converted from mass (g) to force (μ N) using the formula F = ma, where m =145 mass (g), and $a = \text{acceleration} = -9.81 \text{ m}^2/\text{s}$. The mean vertical friction force (MVFF) was 146 calculated using the top five maximum readings. One strain of each species was tested: C. 147 lectularius (SYD strain), C. hemipterus (KL strain). Five adult males and five adult females 148 were used for each species. All these insects were blood-fed 5 days prior to the test. The 149 weight of the insects (g) was individually measured using the balance, and converted to µN 150 using the formula as stated above. The MVFF: insect weight ratio was calculated and used to 151 152 compare between both species, instead of using MVFF to equalize the effect of weight differences between the species. 153

155 SEM observation of the tibial pad (fossula spongiosa). Five adult males and five adult females of the two species (C. lectularius [MH and SYD strains], and C. hemipterus [KL, GL, 156 and PNG strains]) were used for SEM observation of the tibial pad. Two each of 4th instars 157 and 5th instar of *C. lectularius* (MH strain) were examined for the presence of the tibial pad. 158 Bed bugs were preserved in 70% ethanol, dehydrated and cleaned using acetone and a 159 160 Sonicator SC-52H (Sonicor Instrument Corp, New York, US). The fore-, mid- and hind-legs of the specimens were excised and mounted on an aluminium stub. The specimens were 161 coated with gold and observed under a scanning electron microscope (SEM) at 15 kV (Leo 162 163 Supra 50VP field emission SEM, Carl Zeiss SMT, Oberkochen, Germany). The number of tenent hairs (setae) on the tibial pad of each leg was counted from the SEM images taken 164 from different angles. 165

166

Histological examination of the tibial pad in C. lectularius and C. hemipterus. The tibiae 167 with attached tarsomeres of the six legs of three male and three female adults of both C. 168 *lectularius* (MH strain) and *C. hemipterus* (QSL strain) were cut off with microsurgery 169 scissors, and fixed in cold 2% glutaraldehyde, buffered with 50 mM Na-cacodylate and 150 170 mM saccharose. Tissues were postfixed in 2% osmium tetroxide in the same buffer, 171 dehydrated through a graded acetone series and embedded in Araldite resin. Serial semithin 172 173 sections with a thickness of 1 µm were made with a Leica EM UC6 ultramicrotome (Leica Biosystems, Nussloch, Germany), stained with methylene blue and thionin, and examined 174 with an Olympus BX-51 light microscope (Olympus Corporation, Tokyo, Japan). 175

176

Statistical Analysis. Data in percentage of escaped bed bugs from the traps were converted
to arc-sine values before the analyses. *T*-test was used to compare between the trap escape
rates of *C. lectularius* and *C. hemipterus*. Data on vertical friction force, and the number of
tenent hairs for adult males and females of both species were subjected to one-way Analysis
of Variance (ANOVA) and means were separated using Tukey's HSD at 95% confidence
level. All analyses were performed using the statistical program SPSS version 20 (IBM Corp.,
Armonk, NY).

184

Results

Trap escape test. Results showed that all stages tested of *C. hemipterus* were able to escape 185 from each of the traps, with mean combined escape rates (%) ranging from 24.4 ± 4.4 to 75.6 186 \pm 4.4 (Figure 1) per trap, at 72 hours post-treatment. In contrast, most traps were effective in 187 containing C. lectularius with only 0 to 2.2% escape rates, except for the ClimbUp insect 188 interceptor which registered a 26.7 + 10.2% mean escape rate. The escape rates from all traps 189 190 were significantly different (P < 0.05) between C. lectularius and C. hemipterus. For the 191 Volcano trap, an observation of up to 10 days revealed an escape rate of $55.0 \pm 27.5\%$ for C. hemipterus, while no C. lectularius escaped during the corresponding period. 192

193

When the monitors were tested against adults of *C. hemipterus*, the mean cumulative percentage of escaped insects in Verifi and ClimbUp traps were significantly higher (P < 0.05) than the BlackOut and the Volcano traps at 4 hours post-treatment (Figure 2). Regarding *C. lectularius*, significantly (P < 0.05) more adults escaped from the ClimbUp trap than from the other three evaluated traps. No nymphs of either species were able to escape from the Blackout and Volcano traps (Figure 2). For *C. lectularius*, no nymphs escaped from Verifi trap, but $13.3 \pm 6.7\%$ nymphs escaped from ClimbUp trap at 72 hour post-treatment. When

tested against adult *C. lectularius*, 3.3 ± 3.3 % and 33.3 ± 12.0 % adults escaped from Verifi and ClimbUp traps, respectively. Despite some nymph escapees, there were no significant differences (P > 0.05) in the mean cumulative percentage of escaped nymphal bed bugs of all four evaluated traps. Both Blackout and Volcano traps were effective in containing the *C*. *lectularius* adults with no escapees throughout the 72 hours evaluation period (as well as at 10 days post-treatment for Volcano trap).

207

Nymphs of *C. hemipterus* managed to escape from Verifi and ClimbUp traps at the rates of 26.7 \pm 13.3% and 20.0 \pm 11.5%, respectively at 72 hours. There were no nymph escapees for the Blackout and Volcano traps. Verifi was the least efficient trap against the adults of *C. hemipterus* with 100% escapees within 4 hours after the bed bugs were introduced into the trap. ClimbUp trap also showed a high escape rate (>90%) for *C. hemipterus* adults at 72 hour post-treatment. In contrast, 60.0 ± 15.3 and $36.7 \pm 6.7\%$ of adult *C. hemipterus* escaped from Blackout and Volcano traps, respectively, during the same 72-hour evaluation period.

215

216 Vertical friction force. Visual observation revealed that when bed bugs were climbing on the 217 rough surface, the tarsal claws were used, while tibial pads only were used while climbing on smooth surfaces (Figure 3). In this study, C. hemipterus demonstrated a significantly higher 218 219 vertical friction force (P < 0.05) of approximately 3 - 4x that of C. lectularius when tested on smooth glass microscope cover slides (Table 2). However, there appeared to be no differences 220 221 in vertical friction force in both species when the insects were tested on filter paper as a rough substrate. No differences in vertical friction force were found between adult males and 222 females of each species. 223

SEM observation on tibial pad. Tibial pads were present in the adults of both species, but 225 226 they were not present in the fourth instar (Figure 4) and fifth instar. The pads appear as an oval disk of approximately 120 x 40 µm at the ventrodistal portion of the tibia of the 6 legs 227 (Figure 5). The ventral side of the pad is covered with parallel tenent hairs that have a golf 228 229 club-like appearance. The hairs have a length between 30 and 40 μ m and a diameter around 2 μ m, the club-like distal portion having a length of approximately 10 μ m (Figure 5). The 230 231 number of tenent hairs on the tibial pad of C. hemipterus was significantly higher (P < 0.05) than in *C. lectularius* (Table 3, Figure 5). The number of tenent hairs appeared to be similar 232 among the different strains of each species (Table 3), however, the fore- and mid-legs showed 233 234 more tenent hairs than the hind-legs. The three strains of C. hemipterus have almost twice the 235 number of tenent hairs (P < 0.05) on the hind-legs, compared to C. lectularius. The adult male of C. lectularius had significantly more (P < 0.05) tenent hairs than the adult female of the 236 same species on their fore- and mid-legs, but this observation was not recorded for C. 237 hemipterus. The rank in total tenent hairs on the fore-, mid- and hind-legs for one side of the 238 body is as follows: C. hemipterus males = C. hemipterus females > C. lectularius males > C. 239 lectularius females. 240

241

242 Histological examination of tibial pad in *C. lectularius* and *C. hemipterus*. Serial

243 longitudinal sections through the distal tibial part revealed details of the internal anatomy of 244 the tibial pad region (Figure 6A – 6D). The tenent hairs are hollow with an internal diameter 245 around 1 μ m, and penetrate the cuticular plate that forms the base of the tibial pad. In all legs 246 of both males and females of *C. lectularius* and *C. hemipterus*, the tegumental epithelium of 247 the tibia is differentiated into a glandular epithelium. This epithelium in all legs is more 248 pronounced in *C. hemipterus*, where it reaches a thickness of 30 μ m, while in *C. lectularius* it 249 is approximately half as thick (Figure 6A – 6D). At its apical side, the epithelium shows a

subcuticular space in which its secretory products can be stored, and which is continuous with
the central space of the hollow tenent hairs. The epithelium is not associated with any
muscular tissue. The tibia contains two tendons, however, of which the ventral one passes in
the near vicinity of the glandular epithelium. Both tendons are connected with muscles in the
proximal part of the tibia and distally to the basitarsus, of which they direct the movements.

255

256

Discussion

Unlike C. lectularius, adult C. hemipterus were found to escape from all pitfall traps 257 258 evaluated. Examination of the vertical friction force of both species revealed higher vertical friction force in C. hemipterus, compared to C. lectularius. As bed bugs climb on rough 259 surface using their tarsal claws, and on smooth surfaces using the tibial pads, it was suspected 260 that the differences in climbing ability between the two species must be related to variations 261 in the tibial pad. Although Wigglesworth (1938) previously reported that the tibial pad did not 262 263 assist in the climbing of smooth surfaces by the bed bug, he was experimenting using only C. *lectularius.* Should he have worked on *C. hemipterus*, his conclusion would have been very 264 different. In the reduviid kissing bug (Triatoma spp.), it has been proposed that the tibial pads 265 enable the adult bugs to climb smooth surfaces (Gillett & Wigglesworth 1932, Weirauch 266 2007). Usinger (1966) described the presence of the tibial pad in C. lectularius, but did not 267 indicate its presence in C. hemipterus. 268

269

270 To date, the tibial pad had been reported as "fossula spongiosa" by Haridass &

Ananthakrishnan (1980), "apex of tibia" by Walpole (1987), and "tibial brush" by several

authors (Ferris & Usinger 1957, Baker et al. 2016). The tenent hairs (Haridass &

Ananthakrishnan 1980) were described as a "turf of hair" (Usinger 1966), "tibial brush setae"

274 (Baker et al. 2016), or "setae of tibial apex" (Walpole 1987).

276 SEM observation on the tibial pad revealed a greater number of tenent hairs in C. hemipterus than in C. lectularius. By incapacitating the function of tenent hairs on the tibial pad using 277 glue, it was found that this affected the mating process of C. hemipterus, which led to a 278 279 reduction in reproductive success (Kim & Lee, unpublished data). It is speculated that the tibial pad may play an important role in the reproduction of the bed bug, beyond that of 280 281 providing grip in the climbing of smooth surfaces. This argument is further supported by the absence of tibial pads in the nymphal stages, which explains their poor climbing ability as 282 documented in this study. Using SEM, Walpole (1987) too reported that the tibial pad could 283 284 only be found in the adults of C. lectularius and C. hemipterus, but failed to determine its function. In other insect orders such as Blattodea (Clemente & Federle 2008), Diptera 285 (Bauchhenss 1979, Gorb 1998, Gorb & Beutel 2001), Hymenoptera (Dirks & Federle 2011), 286 287 Coleoptera (Ishii 1987, Geiselhardt et al. 2011, Hosoda & Gorb 2011), as well as other families in the order Hemiptera (Gillett & Wigglesworth 1932, Edwards & Tarkanian 1970, 288 Haridass & Ananthakrishnan 1980), the use of the tenent hairs for surface adhesion has been 289 documented. Baker et al. (2016) described the tenent hairs on the tibia pad as "tibia brush 290 291 setae" and also suggested that these setae may be functional during the mating process and for 292 climbing of various surfaces.

293

Although the precise mechanism of how the tibial pad allows climbing on smooth vertical surfaces remains still unknown, it seems obvious that the hollow tenent hairs play a role in the ability to do so. Their central space is continuous with the subcuticular space of the glandular epithelium that is associated with the tibial pad (Baker et al. 2016), suggesting that a glandular secretion can be released through the hairs. With an internal diameter around 1 μ m, mere capillary action may be sufficient for this. In addition to capillarity, secretion may also

be pumped through the hollow hairs, although there is no direct muscular tissue associated 300 301 with the glandular epithelium. However, there is the ventral tibial tendon, that is situated in close proximity to the epithelium, and which may cause a pressure onto the epithelium and 302 303 hence also on the fluid in the subcuticular space. Such pulling action of the tendon may occur during climbing when the tarsomeres need to be moved in order to bring the tibial pad in the 304 right configuration with respect to the substrate. Increased pressure from the tendon onto the 305 306 glandular epithelium at the same time may then result in fluid being pumped into the tenent hairs. A somewhat comparable situation is known in ants, in which adhesion to smooth 307 surfaces is possible through the arolium that acts as an adhesive pad, which is situated at the 308 309 tip of the pretarsus of each leg. The arolium is associated with a sac-like epithelial gland, that 310 works as a hydraulic system by pumping liquid into the arolium upon contraction of the leg tendon (Federle et al. 2001). In a recent study comparing two ant species with very opposite 311 312 climbing ability, it was found that both species possessed an arolium gland, although it was only poorly developed in the non-climbing species (Billen et al. 2016). These reports are in 313 314 line with the observations herein that both Cimex species possess a tibial pad, but that the higher number of tenent hairs and the better developed glandular epithelium in C. hemipterus 315 allow this species to climb smooth vertical surfaces. 316

317

In the experiments described herein, the ClimbUp trap was evaluated without the addition of
talc, and *C. hemipterus* was able to easily escape from the monitor. When talc was applied (as
per manufacturer recommendations), the monitor was able to contain insects of both *C. lectularius* and *C. hemipterus* with minimal escapees (< 5%) over a 72 hour period (Kim &
Lee, unpublished data). Wang & Cooper (2011) reported that regular talc powder
reapplications on pitfall-type monitors are required to prevent bed bugs from escaping. Hottel
et al. (2015) found that talc powder reduced the vertical pulling force of *C. lectularius* on

glass surfaces and the ClimbUp inner wall. In addition, from our experience, the number of 325 326 trapped escapees could be minimized by placing a small piece of folded corrugated cardboard 327 inside the pitfall trap. However, this would require further efforts as the harbourage would need to be examined during monitoring programs. It is anecdotally known that the 328 329 performance of pitfall traps declines in use over time due to the build-up of dust and debris, and can even overcome the addition of talc (Singh et al. 2013, Wang et al. 2013, Cooper et al. 330 331 2016). A trial examining variably aged traps could provide insights into the required maintenance intervals to prevent escape of the respective species. 332

333

The present study demonstrated the need for the design of a more efficient bed bug trap (and the need to reconsider trap maintenance intervals) that could be used against both *C*. *lectularius* and *C. hemipterus*. New low friction surfaces for pitfall traps could also be explored, as long as the cost per unit item for the trap would not financially compromise any monitoring program. It remains unknown at this stage why these sympatric species of bed bugs could have these differences in leg morphology, and the possible biological trade-offs it may have on *C. hemipterus*.

341

342

Acknowledgments

We thank Jamilah Afandi and Johari Othman (Electron Microscopy Unit, Universiti Sains
Malaysia) for assistance with scanning microscopy, An Vandoren (KU Leuven) for making
the histological sections, and Annie Eow for helping with the field collection of *C. hemipterus*.
This study was supported by Bayer Environmental Science, Singapore.

347

348

References Cited

- 349 Baker, G.T., J. Goddard, and A. Lawrence. 2016. Structure of the tibial brush setae of the
- bedbug, *Cimex lectularius* L. (Hemiptera: Cimicidae). Poster presentation. Microscopy and
- 351 Microanalysis 2016. July 24 28, 2016. Columbus, Ohio, USA.
- 352 http://www.microscopy.org/MandM/2016/program/abstracts/PDP-24.pdf (retrieved on
- 353 August 8, 2016).
- **Bauchhenss, E. 1979.** Die Pulvillen von *Calliphora erythrocephala* (Diptera, Brachycera) als
- Adhäsionsorgane. Zoomorphologie 93: 99-123.
- 356 Betz, O. 2002. Performance and adaptive value of tarsal morphology in rove beetles of the
- 357 genus *Stenus* (Coleoptera, Staphylinidae). J. Exp. Biol. 205: 1097–1113.
- 358 Billen, J., M.S. Al-Khalifa and R.R. Silva. 2016. Pretarsus structure in relation to climbing
- ability in the ants *Brachyponera sennaarensis* and *Daceton armigerum*. Saudi J. Biol. Sci.
- 360 (in press) http://dx.doi.org/10.1016/j.sjbs.2016.06.007
- Boase, C., and R. Naylor. 2014. Bed bug management. pp. 8-22. In: Urban insect pests:
- sustainable management strategies (P. Dhang, ed.), CABI Press, London, UK.
- 363 Campbell, B.E., P.G. Koehler, L.J. Buss & R.W. Baldwin. 2016. Recent documentation of
- the tropical bed bug (Hemiptera: Cimicidae) in Florida since the common bed bug
- 365 resurgence. Fla. Entomol. 99: 549-551.
- 366 Clemente, C.J., and W. Federle. 2008. Pushing versus pulling: Division of labour between
- tarsal attachment pads in cockroaches. Proc. R. Soc. B: Biol. Sci. 275:1329-1236.
- 368 Cooper, R.A., C. Wang, and N. Singh. 2016. Evaluation of a model community-wide bed
- bug management program in affordable housing. Pest Manag. Sci. 72: 45 56.
- 370 Doggett, S.L. 2013. A Code of Practice for the Control of Bed Bug Infestations in Australia,
- 4th ed. Department of Medical Entomology, Westmead Hospital, Sydney, and The
- 372 Australian Environmental Pest Managers Association.

- 373 Doggett, S.L., M.J. Geary, W.J. Crowe, P. Wilson, and R.C. Russell. 2003. Has the
- tropical bed bug, *Cimex hemipterus* (Hemiptera: Cimicidae), invaded Australia? J. Environ.
 Health 3: 80-82.
- **Edwards, J.S., and M. Tarkanian. 1970.** The adhesive pads of Heteroptera: a reexamination.
- 377 Physiol. Entomol. 45: 1-5.
- 378 Dirks, J.-H., and W. Federle. 2011. Fluid based adhesion in insects principles and

379 challenges. Soft Mat. 7: 11047-11053.

- 380 Federle, W., E.L. Brainerd, T.A. McMahon and B. Hölldobler. 2001. Biomechanics of the
- movable pretarsal adhesive organ in ants and bees. Proc. Natl Acad. Sci. USA 98: 62156220.
- Ferris, G.F., and R.L. Usinger. 1957. Notes on and descriptions of Cimicidae (Hemiptera).
 Microentomol. 22: 1-37.
- 385 Geiselhardt, S.F., S. Geiselhardt, and K. Peschke. 2011. Congruence of epicuticular

hydrocarbons and tarsal secretions as a principle in beetles. Chemoecol. 21: 181-186.

- 387 Gillett, J.D., and V.B. Wigglesworth. 1932. The climbing organs of an insect, *Rhodnius*
- 388 *prolixus* Stal (Heteroptera-Reduviidae). Proc. R. Entomol. Soc. London B 111: 364–376.
- **Gorb, S., and R. Beutel. 2001.** Evolution of locomotory attachment pads of hexapods.
- 390 Naturwissenschaften 88: 530-534.
- Gorb, S.N. 1998. The design of the fly adhesive pad: distal tenent setae are adapted to the
 delivery of an adhesive secretion. Proc. R. Soc. B: Biol. Sci. 265: 747-752.
- 393 Haridass, E.T., and T.N. Ananthakrishnan. 1980. Functional morphology of the fossula
- spongiosa in some reduviids (Insecta Heteroptera Reduviidae). Proc. Ind. Acad. Sci. 89:
 457-466.
- 396 Hixson, H. 1943. The tropical bedbug established in Florida. Records of *Cimex hemiptera*, F.,
- in houses in several localities in 1938-42. Fla. Entomol. 26: 47.

Hosoda, N., and S.N. Gorb. 2011. Friction force reduction triggers feet grooming behaviour
in beetles. Proc. R. Soc. B: Biol. Sci. 278: 1748-1752.

400 Hottel, B., R. Pereira, S. Gezan, R. Qing, W. Sigmund, and P. Koehler. 2015. Climbing

- 401 ability of the common bed bug (Hemiptera: Cimicidae). J. Med. Entomol. 52: 289-295.
- 402 How, Y.F., and C.Y. Lee. 2010. Survey of bed bugs in infested premises in Malaysia and
- 403 Singapore. J. Vect. Ecol. 35: 89-94.
- 404 Ishii, S. 1987. Adhesion of a leaf feeding ladybird *Epilachna vigintioctomaculata*
- 405 (Coleoptera: Coccinellidae) on a vertically smooth surface. Appl. Entomol. Zool. 22: 222–
 406 228.
- 407 Lee, C.Y. 2013. Bed bugs in Asia Perspective from Southeast Asia, Global Bed Bug

408 Summit, 5–6 December 2013, Denver, Colorado.

- 409 Newberry, K. 1988. Production of a hybrid between the bedbugs *Cimex hemipterus* and
 410 *Cimex lectularius*. Med. Vet. Entomol. 2: 297-300.
- 411 Newberry, K. 1989. The effects on domestic infestations of *Cimex lectularius* bedbugs of
 412 interspecific mating with *Cimex hemipterus*. Med. Vet. Entomol. 3: 407-414.
- 413 Omori, N. 1939. Experimental studies on the cohabitation and crossing of two species of bed-
- 414 bugs (*Cimex lectularius* L. and *C. hemipterus* F.) and on the effects of interchanging of
- 415 males of one species for the other, every alternate days, upon the fecundity and longevity
- 416 of females of each species. Acta Jap. Medic. Tropic. 1: 127-154.
- 417 **Omori, N. 1941.** Comparative studies on the ecology and physiology of common and tropical
- 418 bed bugs, with special reference to the reactions to temperature and moisture. J. Med.
- 419 Assoc. Formosa 60: 555-729.
- 420 Pinto, L.J., R.A. Cooper, and S.K. Kraft. 2007. Bed bug handbook: The complete guide to
- 421 bed bugs and their control. Pinto & Associates, Inc. Mechanicsville, MD, USA.

- 422 Singh, N., C. Wang, and R.A. Cooper. 2013. Effect of trap design, chemical lure, carbon
- 423 dioxide release rate, and source of carbon dioxide on efficacy of bed bug monitors. J. Econ.

424 Entomol. 106: 1802-1811.

- Usinger, R.L. 1966. Monograph of Cimicidae (Hemiptera Heteroptera). Entomological
 Society of America, College Park, Maryland.
- Walpole, D. 1987. External morphology of the legs of two species of bed bugs (Hemiptera:
 Cimicidae). J. Entomol. Soc. South. Afr. 50: 193-201.
- 429 Wang, C. and R.A. Cooper. 2011. Environmentally sound bed bug management solutions.
- 430 pp. 44 63. In: P. Dhang (ed.), Urban Pest Management: An Environmental Perspective.

431 CABI, Oxon, UK.

- 432 Wang, C., N. Singh, R.A. Cooper, C.F. Liu, and G. Buczkowski. 2013. Evaluation of an
- 433 insecticide dust band treatment method for controlling bed bugs. J. Econ. Entomol. 106:
 434 347 352.
- 435 Weirauch, C. 2007. Hairy attachment structures in Reduviidae (Cimicomorpha, Heteroptera),
- with observations on the fossula spongiosa in some other Cimicomorpha. Zool. Anz. 246:155-175.
- 438 Wigglesworth, V. 1938. Climbing organs in insects. Nature 141: 974-975.

439 Zulaikha, Z., A.M.A. Hafiz, A.R.A. Hafis, and A.A. Hassan. 2016. A survey on the

- 440 infestation levels of tropical bed bugs in Peninsular Malaysia: Current updates and status
- 441 on resurgence of *Cimex hemipterus* (Hemiptera: Cimicidae). Asian Pac. J. Trop. Dis. 6:
- 442 40–45.

444 Table 1: The bed bug strains used in this study.

Species	Strain	Year established	Remarks
	Greenlane (GL)	2015	field collected
C	Kuala Lumpur (KL)	2005	lab colony
hemipterus	Queensland (QSL)	~2004	lab colony
	Papua New Guinea (PNG)	1932	curated specimens in 70% ethanol
C. lectularius	Monheim (MH)	~1970	lab colony
	Sydney (SYD)	2004	lab colony

Table 2. Mean vertical friction forces (MVFF) on smooth and rough surfaces, weights and MVFF: weight ratios of the adult male and female of *C. lectularius* and *C. hemipterus*.

450

		$MVFF \pm SE (\mu N)^1$			MVFF: weight ratio	
Species	Sex			weight (µN)		
		Cover glass	Filter paper	-	Cover glass	Filter paper
C heminterus	М	$156.8 \pm 9.2a$	$2476 \pm 140.5a$	$25.5 \pm 2.5 c$	6.3 ± 0.6a	$102 \pm 13.4a$
C. nemipierus	F	$184.8\pm37.8a$	$3203\pm579.5a$	$30.6 \pm 2.0 \text{bc}$	$6.5 \pm 1.7a$	$110 \pm 25.0a$
C loctularius	М	$48.7 \pm 10.2 b$	$2338\pm307.3a$	$37.9 \pm 2.4ab$	$1.3\pm0.3\text{b}$	$62 \pm 6.7a$
C. ieciularius	F	$42.2\pm11.8\text{b}$	3181 ± 516.0a	43.6 ± 4.1a	$1.0\pm0.2b$	77 ± 17.9a

451 ¹Means followed by different letters within the same column are significantly different (P < 0.05; 452 Tukey's HSD).

453

455 Table 3. Mean number of tenent hairs on tibial pad of the fore-, mid- and hind-legs of adult *C*.

hemipterus and *C. lectularius*.

Species	Sex	Strains	Fore leg ¹	Mid leg ¹	Hind leg ¹	Total ¹
		KL	$117.9 \pm 5.7a$	$107.1 \pm 6.0a$	$86.2 \pm 2.5a$	335.7 ± 13.6a
	М	GL	$101.8 \pm 2.6ab$	$97.6 \pm 3.4a$	$78.8 \pm 2.5a$	$292.2\pm9.2ab$
	IVI	PNG	$113.1 \pm 2.9a$	$106.4\pm2.7a$	$79.3 \pm 2.4a$	$318.2 \pm 8.1a$
C homintomus						
C. nemipierus	Б	KL	$100.0 \pm 8.1 \mathrm{ab}$	$112.7 \pm 3.1a$	$88.1 \pm 1.9a$	$309.7 \pm 11.8a$
		GL	$105.7 \pm 4.3a$	$110.3 \pm 3.4a$	$87.7 \pm 2.9a$	$328.6\pm9.9a$
	Г	PNG	$115.9 \pm 3.6a$	$115.7 \pm 4.2a$	$89.8 \pm 1.7a$	$340.6\pm10.0a$
	м	MH	$119.8 \pm 7.3a$	$99.3 \pm 5.4a$	$41.8\pm2.2b$	$247.6 \pm 10.4 bc$
	111	SYD	$114.2 \pm 5.3a$	$102.7 \pm 3.9a$	$49.2\pm2.0b$	$239.4 \pm 12.2c$
C. lectularius						
	F	MH	$66.8 \pm 0.7c$	$71.6 \pm 1.4b$	$43.2\pm2.4b$	$176.8\pm4.0d$
		SYD	$79.6 \pm 1.7 bc$	$78.0\pm3.9b$	$47.8 \pm 1.9 b$	$200.4\pm7.0cd$

458 ¹Means followed by different letters within the same column are significantly different (P<0.05,

459 Tukey's HSD).

Figure captions



Fig. 1. Cumulative percentage of combined escaped adults and nymphs of bed bugs (Mean \pm SEM) from the four traps after 72-h post treatment. Letters above each bar are assigned for comparison between the same trap and different letters denote significant difference (P<0.05, t-test).







Fig. 2. Mean cumulative percentage of escaped bed bugs \pm SE at 4, 24, 48 and 72-h post treatment (Top left: *C. hemipterus* adults; Top right: *C. hemipterus* nymphs; Bottom left: *C. lectularius* adults; Bottom right: *C. lectularius* nymphs). Letters above each bar are assigned for comparison between the traps and different letters denote significant difference (P = 0.05, Tukey's HSD).



Fig. 3. Detail of foreleg of *C. hemipterus* male climbing on plastic petri dish (smooth surface)
(A) and filter paper (rough surface) (B). Note that whitish tibial pad (arrow) is in contact with
substrate only on smooth surface. Observation was made using live bed bugs under an SZ61
stereomicroscope (Olympus, Tokyo, Japan) connected to a CCD camera.



Fig. 4. SEM micrographs showing the absence of tibial pad in 4th instar *C. lectularius* (A, B),
and presence of tibial pad in adult female *C. lectularius* (C, D). The framed area in A and C
indicate the parts that are enlarged in B and D, respectively. t: tarsomeres, Tb: tibia, TP: tibial
pad.



492

Fig. 5. SEM micrographs showing the tibial pad of the hind legs of an adult female in *C. lectularius* (A) and *C. hemipterus* (B). C. Detail of tenent hairs of male *C. hemipterus* mid leg.





Fig. 6. Longitudinal sections through the tibial pad of C. lectularius (A, male mid leg) and C. 496 *hemipterus* (**B**, male fore leg), showing the glandular epithelium (GE), the subcuticular space 497 (asterisks) and the hollow tenent hairs (th). Note the tibia contains two tibial tendons (TT), of 498 which the ventral is close to the glandular epithelium. The sectioning plane in C (C. 499 *lectularius* female hind leg) and **D** (*C. hemipterus* male mid leg) is perpendicular to that in A 500 and B, and is more or less parallel to the ventral surface of the tibia. LN: leg nerve, t: 501 tarsomeres, Tb: tibia. The histological organization in all legs is similar, the photographs 502 shown in this figure were selected as they were the best available images. 503