

1 **Effect of oxygen availability and pH on the furan concentration formed during**
2 **thermal preservation of plant-based foods**

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25 **Effects of oxygen availability and pH on the furan formation during thermal**
26 **preservation of plant-based foods**

27 Thermally treated fruit- and vegetable-based foods are important contributors to the furan exposure of
28 children and adults. Furan reduction by adding or removing precursors from the product has proven to
29 be challenging, because of major food constituents and interactions involved in the reaction pathways
30 leading to furan formation. Instead of intervening at the precursor level, it might be more feasible to
31 influence these formation pathways by adjusting the matrix properties of the product. As opposed to
32 many previous literature sources, the present study investigated the effects of oxygen availability
33 (normal vs. reduced) and pH (acid vs. low-acid) on the furan formation in a real food system. Different
34 combinations of both matrix properties were prepared in a reconstituted potato purée and subjected to a
35 thermal treatment with a pasteurization or sterilization intensity. Irrespective of addition of the furan
36 precursors ascorbic acid, fructose and fatty acids, a considerable furan reduction was observed for the
37 sterilized purées ($F_{121}^{10} = 15$ min) with either a reduced oxygen availability (0.1-1.8 mg/L) or at pH 3.
38 The effects of both matrix properties were less pronounced in the pasteurized purées ($P_{90}^{10} = 10$ min),
39 because of the lower furan concentrations. Even though the mechanisms of furan reduction for both
40 types of matrix properties could not be fully elucidated, the results showed that lowering the oxygen
41 concentration or the pH prior to thermal processing offers a powerful, additional strategy for furan
42 mitigation in thermally treated plant-based foods.

43

44 Keywords: furan; precursor; oxygen; pH; thermal pasteurization; thermal sterilization.

45 Word count: 7215 (main text incl. references)

46 **Introduction**

47 Furan (C₄H₄O) is a small organic molecule with high volatility. In 1995, furan was classified as
48 'possibly carcinogenic' to humans after it was proven to be carcinogenic in rats and mice (International
49 Agency for Research on Cancer (IARC) 1995). As a reaction, North American (US Food and Drug
50 Administration (FDA) 2009) and European (European Food Safety Authority (EFSA) 2011) food safety
51 authorities have analyzed a wide range of commercial foods for furan. The highest concentrations are
52 found in coffee products and canned or jarred foods. Shelf-stable fruit- and vegetable-based products
53 (e.g. jarred baby foods, ready-to-eat soups, sauces and juices) are important contributors to the furan
54 exposure of children and adults. Particular concerns exist about the furan exposure of babies and
55 toddlers, because of the sensitivity of these population groups in combination with a high food intake
56 per kg body weight (Lachenmeier et al. 2009; Owczarek-Fendor et al. 2010a). In anticipation of more
57 information on the toxicological effects of furan for humans, actions should be taken to keep the furan
58 levels in all these products as low as can reasonably be achieved by following good practices at all
59 stages of food production and distribution (Council of the European Communities 2009; Joint
60 FAO/WHO Codex Committee on Contaminants in Foods 2011). Modification of the processing
61 conditions for thermal preservation (Fan et al. 2008; Owczarek-Fendor et al. 2010a; Huang et al. 2011;
62 Sevenich et al. 2014; Palmers et al. 2014; Palmers et al. 2015b) or storage (Palmers et al. 2015a;
63 Palmers et al. 2015c) represents an effective way for furan reduction in fruit- and vegetable-based
64 products. However, a considerable furan reduction can sometimes be hampered by microbial safety
65 standards to comply with and/or economic reasons. For such products, mitigation strategies aiming at an
66 intervention in the reaction pathways for furan formation (e.g. by adding or removing substances from
67 the product) might offer an additional approach to reduce the furan concentration without adverse
68 effects on general food safety or quality attributes. In the literature, many possible reaction pathways
69 leading to the formation of furan have been reported, the major precursors being sugars (alone or in
70 combination with amino acids), ascorbic acid and unsaturated fatty acids, followed by amino acids and
71 carotenoids (Locas & Yaylayan 2004; Becalski & Seaman 2005; Fan 2005; Mark et al. 2006; Limacher

72 et al. 2007; Limacher et al. 2008; Owczarek-Fendor et al. 2010a; Owczarek-Fendor et al. 2011; Van
73 Lancker et al. 2011; Huang et al. 2011; Owczarek-Fendor et al. 2012). It is clear that most of these
74 precursors are major constituents of fruit- and vegetable-based foods, thus contributing to appreciated
75 quality attributes such as the nutritional value, rheology, appearance, etc. Instead of intervening at the
76 precursor level, it is therefore more desirable from an application point of view, to influence these
77 pathways by adjusting matrix properties such as the redox condition and the pH of the product.

78 In the literature, the effect of the redox condition has often been investigated by adding oxidizing or
79 reducing agents (e.g. ferric ions, butylated hydroxytoluene, sulfite, etc.) to model solutions of various
80 precursors (Becalski & Seaman 2005; Mark et al. 2006; Owczarek-Fendor et al. 2010a; Owczarek-
81 Fendor et al. 2011; Mogol & Gokmen 2013), however, with variable results depending on the applied
82 combination of the precursor and redox agentia. Surprisingly, the effect of oxygen on the furan
83 formation has rarely been investigated, even though this approach seems to be one of the most obvious
84 and feasible ways to control the redox condition of the food matrix (Johnson & Decker 2015; Larsen et
85 al. 2015). The effect of pH on the furan formation has been described in the context of ascorbic acid
86 (Fan 2005; Limacher et al. 2007; Fan et al. 2008; Owczarek-Fendor et al. 2010a; Nie et al. 2013a) and
87 sugar degradation (Fan 2005; Limacher et al. 2008; Huang et al. 2011; Owczarek-Fendor et al. 2012;
88 Nie et al. 2013a; Nie et al. 2013b). Like for the redox effect, the pH has variable effects, depending on
89 the precursor. Again, most of the studies are performed with model systems, using phosphate solutions
90 as a buffer for tempering the pH shift during the thermal treatment. However, phosphate ions have
91 recently been shown to enhance furan formation themselves (Fan et al. 2008; Huang et al. 2011). In real
92 food systems, the complexity is further increased, since the pH can have an indirect effect on the furan
93 formation through interactions with proteins, metal ions or even the structure of the matrix. It would
94 therefore be interesting to evaluate the effects of oxygen and pH on the furan formation in real food
95 systems. In this work, potato was selected based on food composition tables (Rijksinstituut voor
96 Volksgezondheid en Milieu 2013), as a vegetable matrix which is naturally low in furan precursors. In
97 addition, potato is a commonly used ingredient of many vegetable-based foods (e.g. jarred baby foods,

98 soups, sauces). By spiking a potato purée with fixed amounts of selected furan precursors (with and
99 without addition of ascorbic acid, fructose or fatty acids) and adjusting the oxygen availability (normal
100 vs. reduced) and the pH (acid vs. low-acid) to targeted levels, a range of well-defined research matrices
101 was obtained, as a compromise between pure model and real systems. The purée formulations were
102 subjected to a thermal treatment with a pasteurization or sterilization intensity and analyzed for furan, to
103 obtain an insight into the effects of precursor composition, pH, oxygen availability and thermal intensity
104 on the furan formation of fruit- and vegetable-based foods. A schematic overview of the experimental
105 approach of the current study is presented in **Figure 1**.

106

107 **Material and methods**

108 *Preparation of the reference and the spiked potato purées*

109 Potatoes (*Solanum tuberosum* ‘Challenger’) were bought at a local supplier and stored in a cold room
110 at 4 °C until further handling. They were first peeled and then cut into slices of approximately 1 cm
111 thickness, before vacuum-packing in low-density polyethylene bags. To assure that all the changes
112 observed during thermal processing were chemical, the potato slices were blanched at 95 °C for 8 min
113 in a water bath (WBU 45, Memmert, Schwabach, Germany). The blanching conditions were validated
114 using a qualitative and quantitative peroxidase test (Adebooye et al. 2008; Vervoort et al. 2012). After
115 blanching, the plastic bags were immediately cooled in iced water for 10 min, frozen in liquid nitrogen
116 and stored in a freezer at -40 °C. Prior to purée preparation, the potato slices were thawed overnight in a
117 cold room at 4°C. The slices were blended (B-400, BÜCHI, Flawil, Switzerland) with a standardized
118 amount of deionized water to obtain a homogeneous potato purée without addition of furan precursors
119 (further addressed to as ‘reference’). The composition of this reference purée was characterized in terms
120 of free sugars, vitamin C and fatty acids (all possible furan precursors) by certified analytical procedures
121 of Eurofins Food Testing Belgium (Brugge, Belgium). The concentration of free sugars (<0.1 g/100 g
122 purée) was found to be below the quantification limit. Also the vitamin C (3.7 mg/100 g purée) and fatty
123 acids (0.5 g/100 g purée, consisting of 7.1% monounsaturated fatty acids, 50.2% polyunsaturated fatty

124 acids and 42.7% saturated fatty acids) concentrations were relatively low, especially when compared
125 with other types of fruit- and vegetable-based foods (Rijksinstituut voor Volksgezondheid en Milieu
126 2013). To assess whether or not the effects of pH and oxygen availability on furan formation are
127 dependent on the type of precursors in the systems, three additional purée formulations were prepared
128 by spiking the blanched potato slices with fructose (5 g/100 g purée), ascorbic acid (150 mg/100 g
129 purée) or olive oil (3 g/100 g purée). The concentration levels of these precursors were selected as high,
130 but realistic levels for these groups of possible furan precursors in fruit- or vegetable-based products
131 (Rijksinstituut voor Volksgezondheid en Milieu 2013). The different purée formulations were prepared
132 by mixing the blanched potato slices with fixed amounts of precursor solutions. For this, stock solutions
133 of fructose (1 g/ml) ($\geq 99\%$, AppliChem, Darmstadt, Germany) and ascorbic acid (25 mg/ml) (99%,
134 Acros Organics, Geel, Belgium) were prepared in deionized water, while olive oil (consisting of 70.6%
135 monounsaturated fatty acids, 11.3% polyunsaturated fatty acids and 18.1% saturated fatty acids)
136 (Vandemoortele, Izegem, Belgium) was added as such. Analogous to the reference purée, the mixtures
137 were further diluted with deionized water to obtain a standardized total volume and blended (B-400,
138 BÜCHI, Flawil, Switzerland) to obtain homogeneous purées, ready for pH adjustment or modifying the
139 oxygen availability. In total, four different purée formulations regarding furan precursors were obtained;
140 (i) without additions (reference), (ii) spiked with fructose, (iii) spiked with ascorbic acid and (iv) spiked
141 with olive oil. All potato purées were prepared in duplicate.

142

143 ***Adjusting the pH and oxygen availability of the spiked potato purées***

144 After the purée preparation, the pH and oxygen availability of the different purée formulations were
145 adjusted to the desired levels. The initial pH values of the potato purées ranged between 5.37 (spiked
146 with ascorbic acid) and 6.10 (spiked with fructose), depending on the purée composition. The pH levels
147 of the purée formulations were adjusted to pH 3 and 7 with 1 M HCl (6.3-6.9 ml/100 g purée) or NaOH
148 (1.5-2.3 ml/100 g purée). Opposite to the pH, it is rather difficult to control the oxygen concentration at
149 a controlled, standardized level. Therefore, the purée formulations were manipulated in such a way that

150 oxygen was removed as much as possible ('reduced oxygen availability') or on the contrary, completely
151 allowed ('normal oxygen availability'). For the purées with reduced oxygen availability, the air was
152 removed with a rotary vane vacuum pump (Type RE 2, Vacuubrand, Werdheim, Germany). Next, the
153 sample holders were transferred into a closed environment (Pyramid portable glove bag, Erlab, Val-de-
154 Reuil, France), which was filled with nitrogen to avoid reintroduction of air. In this oxygen-free
155 environment, the samples were flushed with nitrogen for 10 min to continue the removal of dissolved
156 oxygen from the systems. For the purées with normal oxygen availability, no specific measures were
157 taken to avoid oxygen from entering the systems. On the contrary, the purées were intensively stirred for
158 10 min under normal atmospheric conditions. Due to their inert nature, glass jars (100 ml volume, 95
159 mm height and 45 mm diameter) were used as sample holders for the thermal treatments. Depending on
160 the targeted oxygen availability, the jars were filled with 85 ± 0.5 g of purée under nitrogen atmosphere
161 or normal atmospheric conditions and then closed with metal lids. The efficiency of the procedures for
162 adjusting the oxygen availability inside the potato purées was validated by non-invasive, optical oxygen
163 measurements (O2xyDot, OxySense, Dallas, Texas) in the purée varieties of the reference, and will be
164 discussed in more detail in the Results and discussion section. In total, four possible combinations of pH
165 and oxygen conditions were obtained for each purée formulation.

166

167 *Thermal treatments of the potato purées*

168 The purée formulations were subjected to a thermal treatment for obtaining shelf-stable products, with
169 a thermal intensity depending on the pH of the systems. The potato purées at pH 3 were subjected to a
170 severe pasteurization treatment with a process value of $P_{90}^{10} = 10$ min, the purées at pH 7 were sterilized
171 with a process value of $F_{121}^{10} = 15$ min. The targeted process values for both treatments were indeed
172 much higher than the theoretical values for microbial safety of the respective types of products, but to
173 avoid product recall and to account for non-uniform impact distributions, process values of this order
174 are frequently applied in food industry. In addition to the pasteurization treatment, the potato purées at
175 pH 3 were also sterilized to investigate the effect of pH on the furan formation. The thermal treatments

176 were performed in a static Steriflow pilot retort (Barriquand, Roanne, France). The glass jars filled with
177 potato purée were loaded into the retort, which was heated (come-up time of 8-10 min) at the processing
178 temperature of 90 (pasteurization) or 121 °C (sterilization). The holding time (± 25 min at 90 °C or ± 32
179 min at 121 °C) was calculated in advance to obtain the desired process value in the coldest point of the
180 product. The temperature profiles in the retort and in the coldest point of the product were recorded
181 using type T thermocouples (Ellab, Hilleroed, Denmark) (not shown). After the thermal treatment, the
182 glass jars were immediately transferred to iced water to slow down further chemical reactions. The
183 thermally treated potato purées were emptied in a cold room at 4 °C, frozen in liquid nitrogen and stored
184 in a freezer at -80 °C until analysis.

185

186 ***Quantitation of furan***

187 Quantitation of furan was performed via an isotope dilution assay as described by Palmers et al.
188 (2014), using furan-d₄ as an internal standard. For sample preparation, 2.5 g of the thermally treated
189 purées was weighed into a 10 ml headspace vial with a PTFE/silicone septum seal. The purée was
190 diluted with 2.5 ml of a saturated NaCl solution, 100 μ l of furan-d₄ (98%, Sigma-Aldrich, Saint Louis,
191 Missouri) working solution (ca. 0.05 μ g/ml in deionized water) and deionized water to obtain a
192 standardized total volume of 6 ml. Furan was extracted by solid phase microextraction (SPME), using a
193 75 μ m carboxen/polydimethylsiloxane fiber (Supelco, Bellefonte, Pennsylvania) which was exposed to
194 the headspace of the samples at 30 °C for 15 min. The analyses were carried out using an Agilent
195 7890A gas chromatograph and an Agilent 5975C mass spectrometer (Keysight Technologies, Santa
196 Rosa, California), equipped with a HP-PLOT Q column (30 m \times 320 μ m, 20 μ m film thickness,
197 Keysight Technologies, Santa Rosa, California) and using helium as the carrier gas at a constant flow
198 rate of 2 ml/min. Mass spectra were obtained by electron ionization (EI) at 70 eV, in the combined
199 SCAN and SIM mode. The selected ions monitored were m/z 68 (quantifier) and 39 (qualifier) for furan
200 and m/z 72 (quantifier), 44 and 42 (both qualifier) for furan-d₄. Each sample was analyzed in triplicate.
201 For quantitation, a calibration curve of furan (>99%, Sigma-Aldrich, Saint Louis, Missouri) was

202 prepared in the reference purée without additions, covering the concentration range of 0-50 ng/g purée.
203 The decision limit and the detection capability of the procedure (in accordance with the definitions of
204 European Commission Decision 2002/657/EC) were 1.15 ng/g purée and 1.86 ng/g purée, respectively.
205 Both parameters were established by the calibration curve procedure.

206

207 *Statistical data-analysis*

208 The statistical data analysis was performed with the SAS statistical software package (SAS Enterprise
209 Guide 4.3, Cary, North Carolina). A mixed model was applied, with fixed factors to estimate the effects
210 of the controllable factors (composition, pH level, oxygen availability, thermal intensity) on the furan
211 concentration of the thermally treated potato purées, and random factors to correct for the correlation
212 between samples that were prepared, thermally treated or analyzed together. Significant differences
213 among the fixed factors were examined using the post-hoc Tukey test at a significance level of 0.05.

214

215 **Results and discussion**

216 The effects of oxygen availability and pH on the furan concentration of fruit- and vegetable-based
217 foods were investigated in a potato purée without additions (reference) and in potato purées with the
218 addition of the possible furan precursors fructose, ascorbic acid and fatty acids. For each purée
219 formulation, four different combinations of the selected matrix properties were prepared and subjected
220 to a thermal treatment with a pasteurization or sterilization intensity. An overview of the experimental
221 approach of this study is presented in **Figure 1**.

222

223 *Relative importance of precursors and reaction conditions for furan formation*

224 As can be seen from **Figure 1**, the experimental approach of the present study resulted in a large
225 number of purée formulations (in total 24 combinations consisting of different conditions of precursors,
226 oxygen availability, pH and thermal intensity) to be prepared in duplicate, thermally treated and
227 analyzed for furan. In this section, the relative importance of the selected factors for furan formation in

228 the thermally treated potato purées is investigated. The purée formulations which are illustrative for the
229 effects of oxygen availability and pH, will be discussed in more detail in the following sections. Overall,
230 the furan concentrations of the thermally treated purées varied from not detectable (ND) to 203 ng/g
231 purée. However, the majority of the potato purées had a furan concentration in the lower range, which is
232 indicated by the median and mean concentrations of 18 and 29 ng/g purée, respectively. Even though a
233 comparison with commercially available fruit- and vegetable-based foods is hampered by the variety in
234 matrix properties and the different intensities of the thermal treatment, the furan concentrations of the
235 present study were very similar to results of the reports by the EFSA (e.g. jarred baby foods, with
236 median and mean furan concentrations of 24 and 31-32 ng/g purée, and a maximum value of 233 ng/g
237 purée) (European Food Safety Authority (EFSA) 2011) and by the FDA (only results of individual food
238 products reported) (US Food and Drug Administration (FDA) 2009). For the statistical analysis of the
239 present results, a mixed model regression approach was applied. Given the large number of purée
240 formulations, the potato purées were divided into different groups for the sample preparation (purée
241 preparation and thermal treatment) and furan analysis. The experimental factors (precursor composition,
242 oxygen availability, pH and thermal intensity) are included in the model as fixed main effects, with
243 random effects to capture the possible correlation between the samples that were prepared, thermally
244 treated and/or analyzed together. Interaction effects are omitted from the model, to avoid overfitting of
245 the data and because there is no mechanistic base for most of these effects. The mixed effects model is
246 presented in equation (1):

247

$$248 \quad Y_{klm} = \mu + \sum_{i=1}^4 \beta_{1i} x_{1i} + \sum_{i=1}^2 \beta_{2i} x_{2i} + \sum_{i=1}^2 \beta_{3i} x_{3i} + \sum_{i=1}^2 \beta_{4i} x_{4i} + a_k + b_l + c_m + \varepsilon \quad (1)$$

249

250 where Y_{klm} is the observed furan concentration at the m^{th} repeated observation ($m = 1,2,3$) from the k^{th}
251 preparation group ($k = 1,2,3,4$) and the l^{th} analysis group ($l = 1,2,\dots,12$). μ represents the intercept, β_{1i} ,
252 β_{2i} , β_{3i} and β_{4i} are the regression coefficients of the main effects and x_{1i} , x_{2i} , x_{3i} and x_{4i} are the i^{th} levels of
253 the precursor composition, oxygen availability, pH and thermal intensity, respectively. a_k , b_l and c_m are

254 the random effects of the k^{th} preparation group, the l^{th} analysis group and the m^{th} repetition of furan
255 analysis. The mixed effects model was estimated using generalized least squares for the effects of the
256 experimental factors and restricted maximum likelihood (REML) for the variances of the random
257 effects. The estimates of the regression coefficients are presented in **Table 1**.

258

259 Whether or not the selected furan precursors (i.e., fructose, ascorbic acid or fatty acids) were added to
260 the potato purée, had no significant effect on the furan concentration after thermal treatment (individual
261 estimates for β_1 and type III test, results not shown). However, the mixed effects model showed highly
262 significant effects ($p < 0.0001$) for the oxygen availability, pH and thermal intensity. The potato purées
263 with a normal oxygen availability had a clearly higher furan concentration than the purées with a
264 reduced oxygen availability (positive estimate for β_2). Similarly, the potato purées at pH 3 had a
265 significantly lower furan concentration than the purées at pH 7 (negative estimate for β_3), and the
266 pasteurized potato purées had a significantly lower concentration than the sterilized purées (negative
267 estimate for β_4). As mentioned above, the effects of the oxygen availability and pH will be discussed in
268 more detail in the following sections. Nevertheless, it should be noted that the effects of the matrix
269 properties oxygen availability and pH were of a similar importance as the effect of the thermal intensity.
270 The impact of the thermal processing conditions on the extent of furan formation is well-established in
271 the literature. High furan concentrations are also found in food products that are subjected to an
272 intensive thermal treatment for sterilization or roasting purposes (US Food and Drug Administration
273 (FDA) 2009; European Food Safety Authority (EFSA) 2011). However, the potential for decreasing the
274 thermal impact by adjusting the thermal processing conditions, is often limited by microbial safety
275 standards to comply with and/or technological requirements (e.g. the heating rates delivered by the
276 equipment), especially in the case of thermal sterilization of low-acid, conduction-heated foods like
277 vegetable-based products. The results of the present study seemed to demonstrate that adjusting the
278 selected matrix properties prior to the thermal preservation step can be an interesting, additional strategy
279 for furan mitigation in fruit- and vegetable-based foods, next to optimizing the processing conditions

280 themselves. The degradation of the selected furan precursors is strongly influenced by matrix properties
281 such as the oxygen availability and pH. Since the experimental setup of this study was specifically
282 designed for obtaining insight into the effects of both matrix properties on the furan formation, it was
283 not surprising that these effects dominated the effect of adding precursors. To elucidate the specific role
284 of the furan precursors (which can depend on the concentration, conversion efficiency and possible
285 interactions), a specific experimental setup is needed, which was outside the scope of the present study.

286

287

288 *Effect of oxygen availability on furan formation in the thermally treated potato purées*

289 To investigate the effect of oxygen on the furan formation in fruit- and vegetable-based food products,
290 two types of purée formulations were included in this study (cf. **Figure 1**). The first type ('normal
291 oxygen availability') was prepared under normal atmospheric conditions, in order to obtain a realistic
292 degree of oxidative degradation reactions during thermal processing. The second type of purées
293 ('reduced oxygen availability') was subjected to vacuum conditions and flushed with nitrogen, to
294 remove the dissolved oxygen as much as possible before the pasteurization or the sterilization treatment.
295 The effectiveness of both procedures for setting the oxygen availability, was checked by measuring the
296 oxygen concentration in the different formulations of the reference purées (i.e., the six combinations of
297 oxygen availability, pH and intensity of the thermal treatment). The results of these measurements
298 showed a clear difference between the oxygen concentrations of the purées with a normal oxygen
299 availability (7.4-7.9 mg/L) and the purées with a reduced oxygen availability (0.1-1.8 mg/L),
300 immediately after the purée preparation. After the pasteurization or the sterilization treatments, the
301 oxygen concentrations of all the reference purées were strongly decreased to maximum levels of 0.3
302 mg/L. A large part of the oxygen might be expelled from the glass jars because of the thermal expansion
303 of air. The remaining oxygen was consumed in thermal oxidation reactions of various food constituents,
304 such as the possible furan precursors ascorbic acid and fatty acids. As shown by results of the mixed
305 effects model (**Table 1**), this resulted in significantly lower furan concentrations for the purées with a

306 reduced oxygen availability as compared to the purées with a normal oxygen availability. For a better
307 understanding of this effect in specific formulations, the furan concentrations of the pasteurized purées
308 at pH 3 and the sterilized purées at pH 7 are presented in **Figure 2**. Both are interesting reactions
309 conditions from the perspective of food processing, because they closely resemble the pH and
310 corresponding thermal intensity of shelf-stable fruit- and vegetable-based products. From **Figure 2**, it
311 can be seen that the furan concentrations of the pasteurized purées were low (ND-26 ng/g purée) as
312 compared to the concentrations of the sterilized purées (40-114 ng/g purée). The effect of oxygen on the
313 furan formation was more pronounced in the sterilized purées, where a reduced oxygen availability
314 resulted in a considerable decrease in the furan concentrations of the reference purée (60%), as well as
315 in the other purées with addition of ascorbic acid (58%), fructose (35%) or fatty acids (33%). This is a
316 very interesting observation, because it shows the potential for furan reduction by lowering the oxygen
317 concentration of food products with a various composition.

318

319 Literature studies directly investigating the effect of oxygen on the furan formation in food or model
320 systems are scarce. Nevertheless, it is generally acknowledged that the furan precursors ascorbic acid
321 and fatty acids are prone to oxidative degradation (Joint FAO/WHO Codex Committee on Contaminants
322 in Foods 2011). Ascorbic acid can also be degraded in a non-oxidative way, but the oxidative pathways
323 (via dehydroascorbic acid and hydrolysis to diketogulonic acid) are faster and dominating in the
324 presence of oxygen (Verbeyst et al. 2013). Under pyrolytic conditions (dry heating to 220 °C) and after
325 replacing air inside the reaction vessel by nitrogen, Mark et al. (2006) have observed a considerable
326 decrease (30%) in the furan amounts from ascorbic acid. This is in accordance with the results of the
327 present study, which seems to confirm the importance of the oxidative degradation pathways for the
328 furan formation from ascorbic acid. Oxygen can also act as an initiator for the thermal oxidation of fatty
329 acids. In studies on model systems, the furan concentration is usually higher as the degree of fatty acid
330 unsaturation is higher and the susceptibility to oxidation increases (e.g. for linoleic and linolenic acid)
331 (Becalski & Seaman 2005; Mark et al. 2006). Discussion is ongoing whether or not unrealistically high

332 degrees of fatty acid oxidation are necessary to form furan in real foods (Owczarek-Fendor et al.
333 2010b). In the present study, fatty acid oxidation seemed to have only a small contribution to the furan
334 formation. Despite the high, but realistic amounts of polyunsaturated fatty acids in the spiked olive oil
335 (11.3% of the total fatty acids content, or a concentration of ca. 340 mg/100 g purée), a small difference
336 was observed between the furan concentrations of the different purée formulations with addition of fatty
337 acids. Next to the purées discussed above, a strong decrease in the furan concentration was observed for
338 the reference purée (without addition of precursors) and the purée with addition of fructose. Since the
339 latter formulations contained low concentrations of vitamin C (3.7 mg/100 g purée) and fatty acids (0.5
340 g/100 g purée) (cf. Material and methods), these observations seemed to demonstrate that oxygen has a
341 larger effect on the furan formation in the potato purées than only acting as an initiator for the oxidative
342 degradation reactions. The observed furan reduction might be linked to sugar degradation, which mainly
343 occurs through Maillard reactions under the current conditions (pH 7, high water content). In a very
344 similar research matrix (starch solution at pH 6 with addition of furan precursors), Owczarek-Fendor et
345 al. (2012) have observed a strong synergistic effect on the furan formation between fructose and
346 oxidized soybean oil. Conversely, the addition of ascorbic acid to a fructose-containing solution resulted
347 in a lower than expected furan concentration. Both results indicate that sugar degradation can be
348 affected by changes in the reaction conditions on the redox level, which might be an important part of
349 the explanation for the observed furan reduction in the present study.

350

351 *Effect of pH on furan formation in the thermally treated potato purées*

352 To investigate the effect of pH on the furan formation in fruit- and vegetable-based foods, purée
353 formulations with different pH levels of 3 and 7 were prepared and subjected to a thermal treatment (cf.
354 **Figure 1**). The selected pH values closely resemble the natural pH of fruit- and vegetable-based food
355 products, respectively. Contrary to most other studies (Fan et al. 2008; Huang et al. 2011; Nie et al.
356 2013b), the pH of the purée formulations was adjusted using concentrated HCl or NaOH solutions (1 M)
357 instead of using phosphate buffers. This way, the present study utilized the natural buffering capacity of

358 the potato matrix to stabilize the pH during the thermal treatments. Measurements of the pH after the
359 treatments indicated that the pH shift, which was induced by thermal reactions inside the product, was
360 limited to maximally 0.32 pH unit. For a fair comparison of the pH effect, the potato purées at pH 3 and
361 7 were both subjected to a sterilization treatment (121 °C, $F_0 = 15$ min). For each purée composition, the
362 pH effect was investigated at reduced and normal oxygen availability, even though no interactions were
363 expected between both matrix characteristics. The furan concentrations for the different formulations of
364 the sterilized purées are presented in **Figure 3**. The effect of pH was very similar for the purées with a
365 reduced or a normal oxygen availability. Irrespective of the purée composition (i.e., type of spiked
366 precursor) or the oxygen availability, a pH level of 7 resulted in higher furan concentrations as
367 compared to pH 3. This is again a very interesting observation in the context of furan mitigation,
368 because the observed differences in the furan concentrations of fruit- and vegetable-based foods from
369 monitoring experiments (US Food and Drug Administration (FDA) 2009; European Food Safety
370 Authority (EFSA) 2011) are usually explained by the different thermal intensities of the preservation
371 treatments (e.g. pasteurization vs. sterilization). However, the results of the present study indicate that
372 the lower furan concentrations for fruit-based foods might be partly explained by the acid pH of the
373 products. It should also be noted that the addition of the furan precursors ascorbic acid, fructose or olive
374 oil, did not result in a clear furan increase as compared with the reference purées (see section “Relative
375 importance of precursors and reaction conditions for furan formation”). Possibly, the furan formation
376 from the various precursors was limited by the effects of the thermal intensity, the investigated matrix
377 properties (i.e., oxygen availability and pH) or even other properties (e.g. the amino acid fraction),
378 rather than by the concentrations of the precursors themselves. By spiking the potato purées with high,
379 but realistic concentrations of each precursor, the relative importance of that specific reaction pathway
380 is favored as compared with other pathways. Nevertheless, the observed effect of the investigated matrix
381 property on the furan formation, is always a combination of the effects on all the constituting reaction
382 steps in the potato purée under consideration, as will be explained in the next paragraph.

383

384 As mentioned in the Introduction, the effect of pH on furan formation is dependent on several matrix
385 characteristics such as the precursors under consideration and the type of buffer solution. In real food
386 systems, the complexity is further increased, because the pH might have additional, indirect effects on
387 the furan formation through interactions with proteins (unfolding and/or reaction availability) (Cui et al.
388 2013; Makinen et al. 2016), metal ions (solubility and pro-oxidant properties) (Sorensen et al. 2008) or
389 even the structure of the matrix (Fraeye et al. 2007; Thongkaew et al. 2015). The complexity of the
390 matrix makes it virtually impossible to provide a comprehensive explanation for the effects of pH on the
391 course of a reaction with multiple precursors, such as the furan formation. However, by using a potato
392 purée as a research matrix, the effect of pH on the furan formation could be directly investigated in a
393 commonly used ingredient of various vegetable-based foods. The reduced furan concentration at pH 3
394 compared to pH 7 was consistently observed for all the different purée compositions included in this
395 study (potato purée with and without addition of ascorbic acid, fructose or olive oil). Given the high
396 starch content of potato purée, it can be hypothesized that the acid pH resulted in a partial hydrolysis of
397 the starch. This could have affected its gelating properties (not characterized), but would also provide a
398 supplementary amount of precursors (i.e., glucose) to the matrices. However, since the reference purées
399 (without spiking additional precursors) showed an increased furan formation at pH 7, the furan increase
400 can most probably be explained by an enhanced degradation of the intrinsic precursors ascorbic acid,
401 sugars and fatty acids at pH 7. Ascorbic acid contains an acidic hydroxyl group with a pK_a value slightly
402 higher than 4 (Davey et al. 2000; Belitz et al. 2009) and is therefore prevalent in the stable, non-ionized
403 form in the purée formulations at pH 3. At pH 7, however, it exists as a monovalent anion, which is
404 more reactive and prone to oxidative degradation (and furan formation) (see section “Effect of oxygen
405 availability on furan formation in the thermally treated potato purées”). An analogous reasoning can be
406 set up for the monosaccharides fructose and glucose, which are most stable around pH 3-4 (Belitz et al.
407 2009). Sugars are degraded to furan through Maillard reactions and to a lesser extent also in the absence
408 of amino acids (Locas & Yaylayan 2004; Limacher et al. 2008; Van Lancker et al. 2011). Both reactions
409 are enhanced at pH 7, which has been observed before and is often linked to the favorable conditions for

410 sugar enolization and fragmentation (Limacher et al. 2008; Owczarek-Fendor et al. 2012). Furan
411 formation from fatty acids is a free radical-induced reaction, that is rarely affected by pH. Nevertheless,
412 Fan et al. (2008) and Shen et al. (2015) have observed an optimal furan formation in linoleic and
413 linolenic acid emulsions at pH 6-7 as compared to a lower (pH 3 and 4.55, respectively) or a higher pH
414 (pH 8.68). It is stated that acidic or alkaline model systems can restrain the oxidation of polyunsaturated
415 fatty acids and, therefore, reduce the furan formation (in this case at pH 3). Irrespective of the provided
416 explanation for the pH effect, lowering the pH of the product for furan mitigation should be done with
417 the highest care. As mentioned above, adjusting the pH level can have a large impact on various quality
418 attributes (microbial safety, taste and structure, among others). Reducing the oxygen concentration (see
419 section “Effect of oxygen availability on furan formation in the thermally treated potato purées”) might
420 therefore be considered a more feasible approach for furan reduction in shelf-stable, low-acid foods
421 such as jarred baby foods, soups and sauces.

422

423 **Conclusions**

424 Furan reduction by adding or removing precursors from the product has proven to be challenging,
425 because of major food constituents and interactions involved in the reaction pathways leading to furan
426 formation. Instead of intervening at the precursor level, it might be more feasible from an application
427 point of view, to influence the formation pathways by adjusting the matrix properties of the product. To
428 investigate the effects of the oxygen availability (normal vs. reduced) and the pH (acid vs. low-acid) on
429 furan formation in fruit- and vegetable-based foods, different combinations of both matrix properties
430 were prepared in a potato purée, with and without the addition of the furan precursors ascorbic acid,
431 fructose or fatty acids. Irrespective of the addition of these precursors, a considerable decrease in the
432 furan concentration was observed for the purée formulations with a reduced oxygen availability and the
433 purées at pH 3. In the literature, commercially available, fruit-based food products tend to have a lower
434 furan concentration than vegetable-based foods, which is usually explained by the lower intensity of the
435 thermal treatment for preservation (pasteurization vs. sterilization). Based on the results of this study,

436 the difference in furan concentration can partly be attributed to the acid pH of the fruit-based products.
437 In the context of furan mitigation, priority should be given to the optimization of the thermal
438 preservation process for fruit- and vegetable-based foods (i.e., sterilization of low-acid, conduction-
439 heated foods). If this thermal optimization is hindered by microbial safety standard to comply with or an
440 extended shelf life, lowering the oxygen concentration and/or the pH of the products might offer an
441 additional, powerful strategy for furan mitigation. However, since the mechanisms of furan reduction
442 for both matrix characteristics could not be fully established, the adjustments of the product composition
443 should always be accompanied by a thorough evaluation of the effects on other food safety and quality
444 attributes (e.g. structure, taste, color). Ideally, various mitigation strategies should be brought together in
445 an industrial toolbox for furan reduction, as already done for acrylamide by FoodDrinkEurope (2014).
446 By providing measures at the different steps of the food chain (product formulation, thermal processing
447 and storage), such a toolbox could allow food processors to reduce the furan concentrations of various
448 fruit- and vegetable-based foods, with limited adverse side effects on other important food attributes.

449

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454

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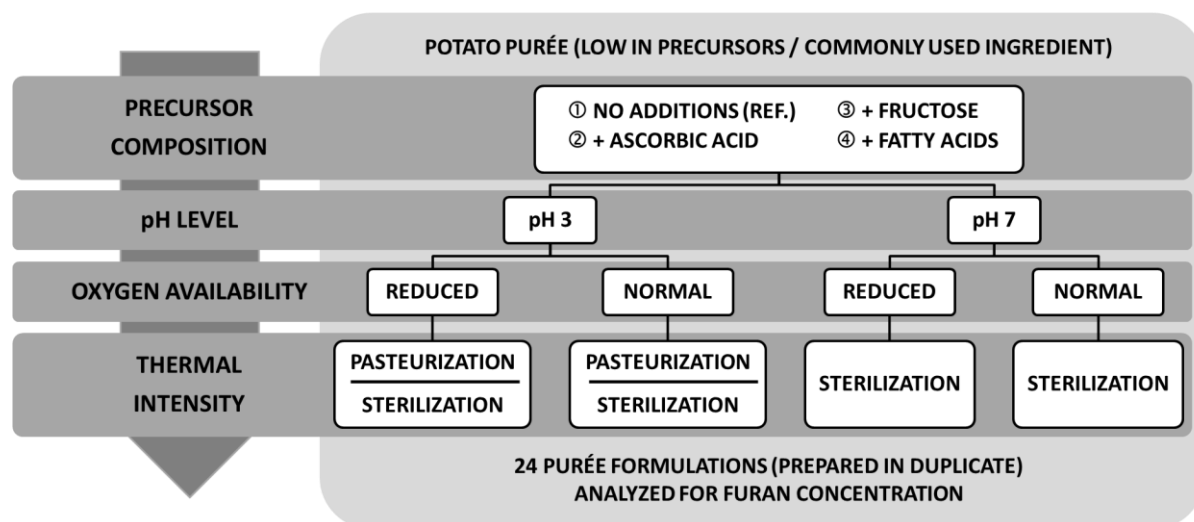
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- 564
- 565

568 **Table 1.** Estimates of the regression coefficients and significance tests for the mixed effects model
 569 describing the furan concentrations in the thermally treated potato purées as a function of precursor
 570 composition, oxygen availability, pH and thermal intensity.

Effect	Value	Estimate	Standard error	DF	t Ratio	p Value
Intercept (μ)		46.89				
Precursor (β_1)	Reference	4.77	25.76	124	0.19	0.8533
Precursor (β_1)	Fructose	-4.33	9.10	124	-0.48	0.6352
Precursor (β_1)	Ascorbic acid	20.53	12.74	124	1.61	0.1096
Oxygen availability (β_2)	Normal	21.97	3.35	124	6.56	<0.0001
pH level (β_3)	3	-40.39	5.15	124	-7.84	<0.0001
Thermal intensity (β_4)	Pasteurization	-21.97	5.15	124	-4.26	<0.0001

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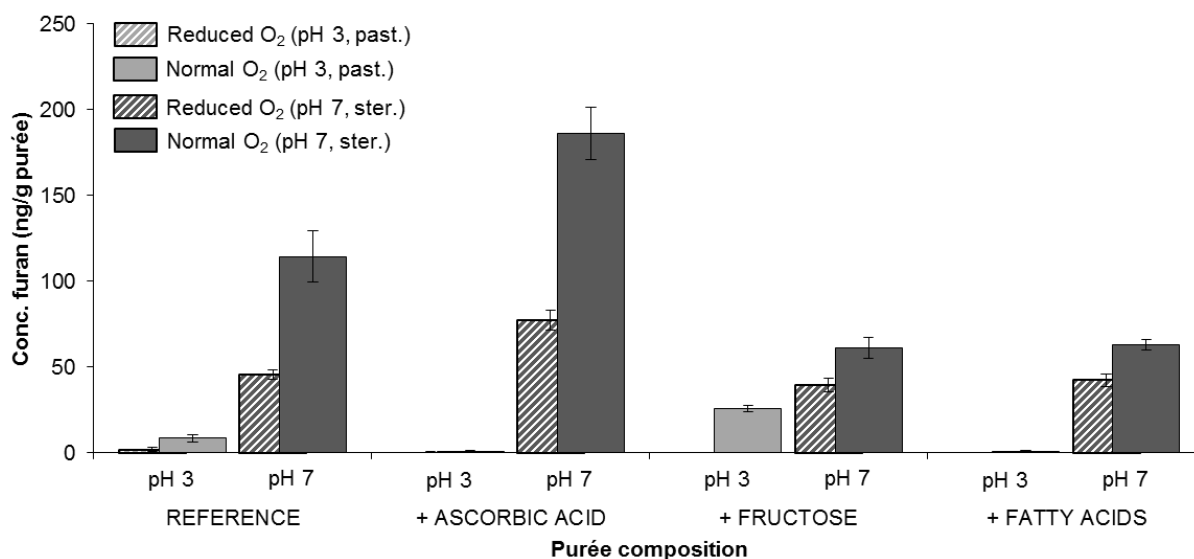


574

575 **Figure 1.** Schematic overview of the experimental approach for purée preparation. All the purée

576 formulations were prepared in duplicate, thermally treated and analyzed for furan.

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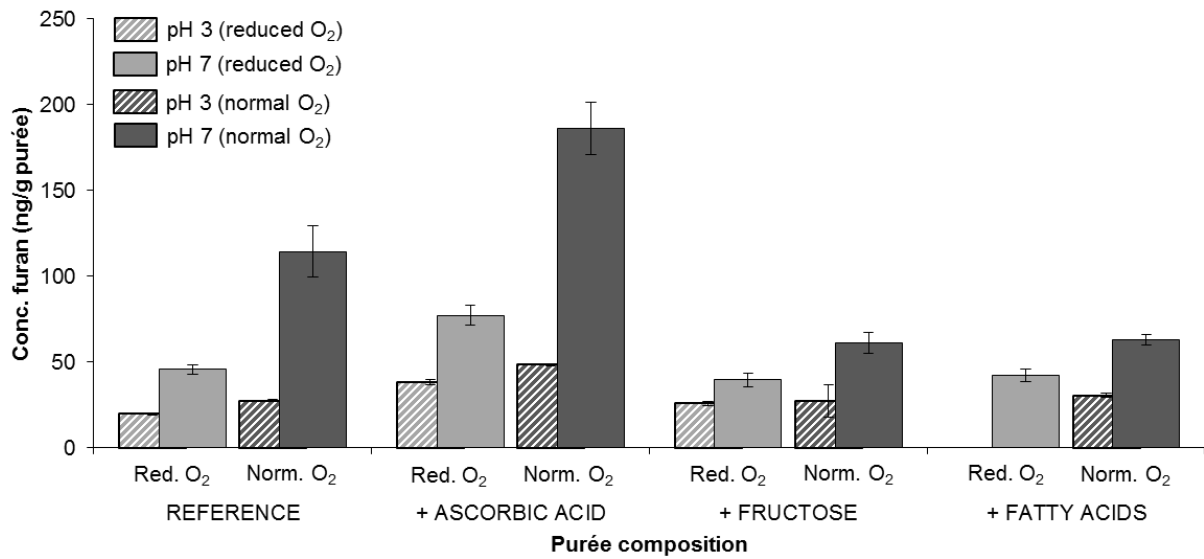
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579 **Figure 2.** Effect of oxygen availability on the furan concentration of the thermally treated, spiked

580 potato purées (past. = pasteurization, ster. = sterilization).

581

582



583

584 **Figure 3.** Effect of pH on the furan concentration of the thermally sterilized, spiked potato purées.

585